P1 819731

THE UNITED STATES OF AMERICA

TO ALL TO WHOM THESE PRESENTS SHALL COME:

UNITED STATES DEPARTMENT OF COMMERCE
United States Patent and Trademark Office

June 12, 2002

THIS IS TO CERTIFY THAT ANNEXED HERETO IS A TRUE COPY FROM THE RECORDS OF THE UNITED STATES PATENT AND TRADEMARK OFFICE OF THOSE PAPERS OF THE BELOW IDENTIFIED PATENT APPLICATION THAT MET THE REQUIREMENTS TO BE GRANTED A FILING DATE.

APPLICATION NUMBER: *09/826,115*
FILING DATE: *April 04, 2001*
RELATED PCT APPLICATION NUMBER: *PCT/US02/10764*

By Authority of the
COMMISSIONER OF PATENTS AND TRADEMARKS

T. WALLACE
Certifying Officer

PRIORITY DOCUMENT
SUBMITTED OR TRANSMITTED IN COMPLIANCE WITH RULE 17.1(a) OR (b)

J102 U.S. PTO 04/04/01

CONTINUING APPLICATION TRANSMITTAL FORM
(37 C.F.R. § 1.53(b))

ANTICIPATED CLASSIFICATION OF THIS APPLICATION:	PRIOR APPLICATION: 09/701,536	
CLASS: SUBCLASS:	EXAMINER: Unassigned	ART UNIT: Unassigned

JC968 U.S. PTO 09/826615 04/04/01

To the Assistant Commissioner for Patents:

This is a request for filing a continuation-in-part application under 37 C.F.R. § 1.53(b), of pending prior application Serial No. 09/701,536, filed on November 29, 2000, which is a national phase application filed from international patent application PCT/US99/12298, filed June 3, 1999, which claims priority to Provisional application Serial No. 60/087,908, filed June 4, 1998 of Gwong-Jen J. Chang for "NUCLEIC ACID VACCINES FOR PREVENTION OF FLAVIVIRUS INFECTION.

If any extension of time is necessary for the filing of this application, including any extension of time necessary in the prior application for maintaining copendency between the prior application and this application, and such extension has not otherwise been requested, Applicant hereby petitions for such an extension in the prior application.

1. Enclosed is a specification as prescribed by 35 U.S.C. § 112 containing a description pursuant to § 1.71 and at least one claim pursuant to § 1.75, and any drawing required by § 1.81(a).

2. The inventor of the invention being claimed in this application is: Gwong-Jen J. Chang

3A. [] This application is a continuation or divisional application (not a continuation-in-part) that:

(i) names as inventors the same or fewer than all of the inventors named in the prior application; and
(ii) contains no matter that would have been new matter in the prior application; and
(iii) a copy of the executed oath or declaration filed in the prior application, showing the signature or an indication thereon that it was signed:

[] is enclosed.
[] will follow.

[] This application is being filed by less than all the inventors named in the prior application. In accordance with 37 C.F.R.§1.53(d)(4), the Commissioner is requested to delete the name(s) of the following persons who are not inventors of the invention being claimed in this application:

--OR--

3B. [X] This application is a continuation-in-part application, or a continuation or divisional application naming an inventor not named in the prior application. A newly executed oath or declaration:

[X] is enclosed.
[] will follow.

4. [] Amend the Title of the Invention as follows:

5. [] Amend the specification by inserting before the first line the sentence:

6. [] Priority of foreign application number is claimed under 35 U.S.C. § 119.

[] The certified copy has been filed in prior application Serial No. , filed on .

7. [] Cancel in this application original claims of the prior application before calculating the filing fee. (At least one original independent claim must be retained for filing purposes.)

8. [] A preliminary amendment is enclosed.

9. [X] New formal drawings are enclosed.

10. [X] Also enclosed is/are: Sequence listing (46 pages); Sequence listing diskette; and form entitled Authorization to Treat Reply Requiring Extension of Time as Incorporating Petition for Extension of Time and Payment of Extension of Time Fee.

11. [] The prior application is assigned of record to:

12. [] Applicant claims small entity status. See 37 C.F.R. § 1.27.

13. The filing fee is calculated as follows:

TOTAL CLAIMS	43 - 20 =	23	x $18.00	$414.00
INDEPENDENT CLAIMS	1 - 3 =	0	x $80.00	$0.00
MULTIPLE DEPENDENT CLAIM(S) (IF APPLICABLE) =			+ $270.00	$0.00
BASIC FEE =				$710.00
TOTAL OF ABOVE CALCULATIONS =				$1,124.00
REDUCTION BY ½ FOR FILING BY SMALL ENTITY (NOTE 37 C.F.R. § 1.9, § 1.27, § 1.28)				$
TOTAL NATIONAL FEE =				$1,124.00

14. [X] A check in the amount of $1,124.00 is enclosed.

15. [X] The Commissioner is hereby authorized to charge any fees which may be required under 37 C.F.R. § 1.16 and § 1.17, or credit any overpayment to Deposit Account No. 14-0629.

16. [X] The Power of Attorney in the prior application is to: Needle & Rosenberg, P.C.

a. [X] The Power of Attorney appears in the original papers in the prior application.

b. [] Since the Power does not appear in the original papers, a copy of the power in the prior application is enclosed.

c. [X] Address all future correspondence to: (May only be completed by applicant, or attorney or agent of record.)

Mary L. Miller, Esq.
NEEDLE & ROSENBERG, P.C.
Suite 1200, The Candler Building
127 Peachtree Street, N.E.
Atlanta, GA 30303-1811

17. I hereby verify that all statements made herein of my own knowledge are true; and further that these statements were made with the knowledge that willful false statements and the like are made punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Mary L. Miller
REG. NO. 39,303

Mary L. Miller
SIGNATURE

April 4, 2001
DATE

[] Inventor(s)

[] Assignee of complete interest

[X] Attorney or agent of record

[] Filed under 37 C.F.R. § 1.34(a)
(Registration No. is acting under 37 C.F.R. § 1.34(a):

EXPRESS MAIL NO. EL 491887575 US

CERTIFICATE OF EXPRESS MAILING

I hereby certify that this correspondence is being deposited with the United States Postal Service as Express Mail Invoice No. EL 491887575 US in an envelope addressed to: BOX PATENT APPLICATION, Assistant Commissioner for Patents, Washington, D.C. 20231, on the date shown below.

Everardo McFarlane

4-4-01
DATE

EXPRESS MAIL NO. EL491587108US

ATTORNEY DOCKET NO. 14114.0332U3

PATENT

TO ALL WHOM IT MAY CONCERN:

Be it known that I, Gwong-Jen J. Chang, a citizen of the United States of America, residing at 4237 Beaver Creek Drive, Ft. Collins, CO 80526, U.S.A., have invented new and useful improvements in

NUCLEIC ACID VACCINES FOR PREVENTION OF FLAVIVIRUS INFECTION

for which the following is a specification.

NUCLEIC ACID VACCINES FOR PREVENTION OF FLAVIVIRUS INFECTION

This application is a continuation-in-part of, and claims the benefit of, U.S. application Serial No. 09/701,536, filed November 29, 2000, which status is pending and which is a national stage application of international application Serial No. PCT/US99/12298, filed June 3, 1999 from U.S. provisional application Serial No. 60/087908, filed June 4, 1998, which applications are hereby incorporated herein in their entirety by reference.

Field of the Invention

This invention relates to novel vaccines, diagnostics and methods of using both in the treatment and prevention of the diseases caused by flaviviruses. In particular, the vaccines are recombinant nucleic acids which contain genes for structural proteins of flaviviruses, such as Japanese encephalitis virus (JEV), West Nile virus (WNV) or related flaviviruses. These vaccines serve as a transcriptional unit for the biosynthesis of the virus protein antigens when administered *in vivo*. The diagnostics are compositions containing antigens produced from the recombinant nucleic acids that can be used to detect flavivirus infection.

Background of the Invention

Flaviviruses are members of the genus *Flavivirus*, which is classified within the family Flaviviridae. The flaviviruses are largely pathogenic to humans and other mammals. Flaviviruses that inflict disease upon humans and animals include Alfuy, Apoi, Aroa, Bagaza, Banzi, Batu Cave, Bouboui, Bukalasa bat, Bussuquara, Cacipacore, Carey Island, Cowbone Ridge, Dakar bat, Dengue (serotypes 1, 2, 3 and 4), Edge Hill, Entebbe bat, Gadgets Gully, Iguape, Ilheus, Israel turkey

meningoencephalitis, Japanese encephalitis, Jugra, Jutiapa, Kadam, Karshi, Kedougou, Kokobera, Koutango, Kunjin, Kyasanur Forest disease, Langat, Meaban, Modoc, Montana myotis leukoencephalitis, Murray Valley encephalitis, Naranjal, Negishi, Ntaya, Omsk hemorrhagic fever, Phnom Penh bat, Potiskum, Powassan, Rio Bravo, Rocio, Royal Farm, Russian spring summer encephalitis, Saboya, Sal Vieja, San Perlita, Saumarez Reef, Sepik, Sokuluk, Spondweni, St. Louis encephalitis, Stratford, Tick-borne encephalitis - central European subtype, Tick-borne encephalititis - far eastern subtype, Tembusu, THCAr, Tyuleniy, Uganda S, Usutu, West Nile, Yaounde, Yellow fever, Yokose, Ziki, Cell fusing agent and other related flaviviruses, as listed in Kuno et al. (*J. Virol.* 72: 73-83 (1998)).

The flaviviruses contain the following three structural proteins: prM/M, the premembrane and membrane protein; E, the envelope protein; and C, the capsid protein. (Monath, in Virology (Fields, ed.), Raven Press, New York, 1990, pp. 763-814; Heinz and Roehrig, in Immunochemistry of Viruses II: The Basis for Serodiagnosis and Vaccines (van Regenmortel and Neurath, eds.), Elsevier, Amsterdam, 1990, pp. 289-305). M has a molecular weight (MW) of about 7-8 kilodaltons (kDa) and E has a MW of about 55-60 kDa. M is synthesized as a larger precursor termed prM. The pr portion of prM is removed when prM is processed to form M protein in mature virions. M and E are located in the membrane of the flavivirus particle, and so have long been considered to constitute important immunogenic components of the viruses.

The flaviviruses are RNA viruses comprising single stranded RNA having a length, among the various species, of about 10 kilobases (kb). The C protein, with a MW of 12-14 kDa, complexes with the RNA to form a nucleocapsid complex. Several nonstructural proteins are also encoded by the RNA genome which are termed NS1, NS2A, NS2B, NS3, NS4A, NS4B and NS5. The genome is translated within the host cell as a polyprotein, then processed co- or post-translationally into the individual gene products by viral- or host-specific proteases (Figure 1).

The nucleotide sequences of the genomes of several flaviviruses are known, as summarized in U. S. Patent No. 5,494,671. That for JEV is provided by Sumiyoshi et al. (*Virology* 161: 497-510 (1987)) and Hashimoto et al. (*Virus Genes* 1: 305-317 (1988)). The nucleotide sequences of the virulent strain SA-14 of JEV and the attenuated strain SA-14-14-2, used as a vaccine in the People's Republic of China, are compared in the work of Nitayaphan et al. (*Virology* 177: 541-552 (1990)).

Nucleotide sequences encoding the structural proteins of other flavivirus species are also known. In many cases, the sequences for the complete genomes have been reported. The sequences available include dengue serotype 1 virus, dengue serotype 2 virus (Deubel et al., *Virology* 155: 365-377 (1986); Gruenberg et al., *J. Gen. Virol.* 69: 1391-1398 (1988); Hahn et al. *Virology* 162: 167-180 (1988)), dengue serotype 3 virus (Osatomi et al., *Virus Genes* 2: 99-108 (1988)), dengue serotype 4 virus (Mackow et al., *Virology* 159: 217-228 (1987), Zhao et al., *Virology* 155: 77-88 (1986)), West Nile virus (Lanciotti et al., *Science* 286: 2331-2333 (1999)), Powassan virus (Mandl et al., *Virology* 194: 173-184 (1993)) and yellow fever virus (YFV) (Rice et al., *Science* 229: 726-733 (1985)).

Many flaviviruses, including St. Louis encephalitis virus (SLEV), WNV and JEV, are transmitted to humans and other host animals by mosquitoes. They therefore occur over widespread areas and their transmission is not easily interrupted or prevented.

West Nile fever is a mosquito-borne flaviviral infection that is transmitted to vertebrates primarily by various species of *Culex* mosquitoes. Like other members of the Japanese encephalitis (JE) antigenic complex of flaviviruses, including JE, SLE and Murray Valley encephalitis (MVE) viruses, WNV is maintained in a natural cycle between arthropod vectors and birds. The virus was first isolated from a febrile human in the West Nile district of Uganda in 1937 (Smithburn et al., *Am. J. Trop. Med. Hyg.* 20: 471-492 (1940)). It was soon recognized as one of the most widely distributed

flaviviruses, with its geographic range including Africa, the Middle East, Western Asia, Europe and Australia (Hubalek et al., *Emerg. Infect. Dis.* 5: 643-50 (1999)). Clinically, West Nile fever in humans is a self-limited acute febrile illness accompanied by headache, myalgia, polyarthropathy, rash and lymphadenopathy (Monath and Tsai, in Clinical Virology, (Richman, Whitley and Hayden eds.), Churchill-Livingtone, New York, 1997, pp. 1133-1186). Acute hepatitis or pancreatis has been reported on occasion and cases of WNV infection in elderly patients are sometimes complicated by encephalitis or meningitis (Asnis et al., *Clin. Infect. Dis.* 30: 413-418 (2000)). Thus, infection by WNV is a serious health concern in many regions of the world.

The geographical spread of the disease, particularly the introduction of WNV into the U.S. in 1999, has greatly increased awareness of the human and animal health concerns of this disease. Between late August and early September 1999, New York City and surrounding areas experienced an outbreak of viral encephalitis, with 62 confirmed cases, resulting in seven deaths. Concurrent with this outbreak, local health officials observed increased mortality among birds (especially crows) and horses. The outbreak was subsequently shown to be caused by WNV, based on monoclonal antibody (Mab) mapping and detection of genomic sequences in human, avian and mosquito specimens (Anderson et al., *Science* 286: 2331-2333 (1999); Jia et al., *Lancet* 354: 1971-1972 (1999); Lanciotti et al., *Science* 286: 2333-2337 (1999)). Virus activity detected during the ensuing winter months indicated that the virus had established itself in North America (*Morb. Mortal. Wkly. Rep.* 49: 178-179 (2000); Asnis et al., *Clin. Infect. Dis.* 30: 413-418 (2000); Garmendia et al., *J. Clin. Micro.* 38: 3110-3111 (2000)). Surveillance data reported from the northeastern and mid-Atlantic states during the year 2000 confirmed an intensified epizootic/epidemic transmission and a geographic expansion of the virus with documentation of numerous cases of infection in birds, mosquitoes and horses, as well as cases in humans (*Morb. Mortal. Wkly. Rep.* 49: 820-822 (2000)).

Currently, no human or veterinary vaccine is available to prevent WNV infection and mosquito control is the only practical strategy to combat the spread of the disease.

Japanese encephalitis virus (JEV) infects adults and children and there is a high mortality rate among infants, children and the elderly in areas of tropical and subtropical Asia (Tsai et al., in Vaccines (Plotkin, ed.) W.B. Saunders, Philadelphia, Pa, 1999, pp. 672-710). Among survivors, there are serious neurological consequences, related to the symptoms of encephalitis, that persist after infection. In more developed countries of this region, such as Japan, the Republic of China (Taiwan) and Korea, JEV has been largely controlled by use of a vaccine of inactivated JEV. Nevertheless, it is still prevalent in other countries of the region.

Vaccines available for use against JEV infection include live virus inactivated by such methods as formalin treatment, as well as attenuated virus (Tsai et al., in Vaccines (Plotkin, ed.) W.B. Saunders, Philadelphia, Pa, 1994, pp. 671-713). Whole virus vaccines, although effective, do have certain problems and/or disadvantages. The viruses are cultivated in mouse brain or in cell culture using mammalian cells as the host. Such culture methods are cumbersome and expensive. Furthermore, there is the attendant risk of incorporating antigens from the host cells, i.e., the brain or other host, into the final vaccine product, potentially leading to unintended and undesired allergic responses in the vaccine recipients. There is also the risk of inadvertent infection among workers involved in vaccine production. Finally, there is the risk that the virus may not be fully or completely inactivated or attenuated and thus, the vaccine may actually cause disease.

Dengue fever and dengue hemorrhagic fever (DF/DHF) are caused by dengue virus, which is also a mosquito-borne flavivirus. There are four antigenically related, but distinct, dengue virus serotypes, (DEN-1, DEN-2, DEN-3 and DEN-4), all of which can cause DF/DHF. Symptoms of DF, the mild form of dengue-related disease, include

fever, rash, severe headache and joint pain. Mortality among those subjects suffering from DF is low; however, among those subjects suffering from DHF, mortality can be as high as 5%. From available evidence, more than 3 million cases of DHF and 58,000 deaths have been attributed to DHF over the past 40 years, making DHF a major emerging disease (Halstead, in Dengue and Dengue Hemorrhagic Fever (Gubler and Kuno, eds.) CAB International, New York, NY, (1997) pp 23-44). Nevertheless, despite decades of effort, safe and effective vaccines to protect against dengue virus infection are not yet available.

Yellow fever is prevalent in tropical regions of South America and sub-Saharan Africa and is transmitted by mosquitos. Infection leads to fever, chills, severe headache and other pains, anorexia, nausea and vomiting, with the emergence of jaundice. A live virus vaccine, 17D, grown in infected chicken embryos, is considered safe and effective. Nevertheless, there remains a need for a vaccine that is stable under adverse conditions, such as are commonly encountered in the tropical regions of Africa and the Americas where the vaccine is most needed.

A recombinant flavivirus which is a chimera between two flaviviruses is disclosed in PCT publication WO 93/06214. The chimera is a construct fusing non-structural proteins from one "type," or serotype, of dengue virus or a flavivirus, with structural proteins from a different "type," or serotype, of dengue virus or other flavivirus.

Several recombinant subunit and viral vaccines have been devised in recent years. U. S. Patent No. 4,810,492 describes the production of the E glycoprotein of JEV for use as the antigen in a vaccine. The corresponding DNA is cloned into an expression system in order to express the antigen protein in a suitable host cell such as *E. coli*, yeast, or a higher organism cell culture. U. S. Patent No. 5,229,293 discloses recombinant baculovirus harboring the gene for JEV E protein. The virus is used to

infect insect cells in culture such that the E protein is produced and recovered for use as a vaccine.

U. S. Patent No. 5,021,347 discloses a recombinant vaccinia virus genome into which the gene for JEV E protein has been incorporated. The live recombinant vaccinia virus is used as the vaccine to immunize against JEV. Recombinant vaccinia viruses and baculoviruses in which the viruses incorporate a gene for a C-terminal truncation of the E protein of dengue serotype 2, dengue serotype 4 and JEV are disclosed in U.S. Patent 5,494,671. U. S. Patent 5,514,375 discloses various recombinant vaccinia viruses which express portions of the JEV open reading frame extending from prM to NS2B. These pox viruses induced formation of extracellular particles that contain the processed M protein and the E protein. Two recombinant viruses encoding these JEV proteins produced high titers of neutralizing and hemagglutinin-inhibiting antibodies, and protective immunity, in mice. The extent of these effects was greater after two immunization treatments than after only one. Recombinant vaccinia virus containing genes for the prM/M and E proteins of JEV conferred protective immunity when administered to mice (Konishi et al., *Virology* 180: 401-410 (1991)). HeLa cells infected with recombinant vaccinia virus bearing genes for prM and E from JEV were shown to produce subviral particles (Konishi et al., *Virology* 188: 714-720 (1992)). Dmitriev et al. reported immunization of mice with a recombinant vaccinia virus encoding structural and certain nonstructural proteins from tick-borne encephalitis virus (*J. Biotechnology* 44: 97-103 (1996)).

Recombinant virus vectors have also been prepared to serve as virus vaccines for dengue fever. Zhao et al. (*J. Virol.* 61: 4019-4022 (1987)) prepared recombinant vaccinia virus bearing structural proteins and NS1 from dengue serotype 4 and achieved expression after infecting mammalian cells with the recombinant virus. Similar expression was obtained using recombinant baculovirus to infect target insect cells (Zhang et al., *J. Virol.* 62: 3027-3031(1988)). Bray et al. (*J. Virol.* 63: 2853-2856 (1989)) also reported a recombinant vaccinia dengue vaccine based on the E protein

gene that confers protective immunity to mice against dengue encephalitis when challenged. Falgout et al. (*J. Virol* 63: 1852-1860 (1989)) and Falgout et al. (*J. Virol.* 64: 4356-4363 (1990)) reported similar results. Zhang et al. (*J. Virol* 62: 3027-3031 (1988)) showed that recombinant baculovirus encoding dengue E and NS1 proteins likewise protected mice against dengue encephalitis when challenged. Other combinations in which structural and nonstructural genes were incorporated into recombinant virus vaccines failed to produce significant immunity (Bray et al., *J. Virol.* 63: 2853-2856 (1989)). Also, monkeys failed to develop fully protective immunity to dengue virus challenge when immunized with recombinant baculovirus expressing the E protein (Lai et al. (1990) pp. 119-124 in F. Brown, R. M. Chancock, H. S. Ginsberg and R. Lerner (eds.) Vaccines 90: Modern approaches to new vaccines including prevention of AIDS, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY).

Immunization using recombinant DNA preparations has been reported for SLEV and dengue-2 virus, using weanling mice as the model (Phillpotts et al., *Arch. Virol.* 141: 743-749 (1996); Kochel et al., *Vaccine* 15: 547-552 (1997)). Plasmid DNA encoding the prM and E genes of SLEV provided partial protection against SLEV challenge with a single or double dose of DNA immunization. In these experiments, control mice exhibited about 25% survival and no protective antibody was detected in the DNA-immunized mice (Phillpotts et al., *Arch. Virol.* 141: 743-749 (1996)). In mice that received three intradermal injections of recombinant dengue-2 plasmid DNA containing prM, 100% developed anti-dengue-2 neutralizing antibodies and 92% of those receiving the corresponding E gene likewise developed neutralizing antibodies (Kochel et al., *Vaccine* 15: 547-552 (1997)). Challenge experiments using a two-dose schedule, however, failed to protect mice against lethal dengue-2 virus challenge.

The vaccines developed to date for immunizing against infection by JEV, SLEV, dengue virus and other flaviviruses have a number of disadvantages and problems attending their use. Inactivated vaccine is costly and inconvenient to prepare. In addition, any such vaccine entails the risk of allergic reaction originating from

proteins of the host cell used in preparing the virus. Furthermore, such vaccines present considerable risk to the workers employed in their production. Candidate attenuated JEV vaccines are undergoing clinical trials, but as of 1996 have not found wide acceptance outside of the People's Republic of China (Hennessy et al., *Lancet* 347: 1583-1586 (1996)).

Recombinant vaccines based on the use of only certain proteins of flaviviruses, such as JEV, produced by biosynthetic expression in cell culture with subsequent purification or treatment of antigens, do not induce high antibody titers. Also, like the whole virus preparations, these vaccines carry the risk of adverse allergic reaction to antigens from the host or to the vector. Vaccine development against dengue virus and WNV is less advanced and such virus-based or recombinant protein-based vaccines face problems similar to those alluded to above.

There is therefore a need for vaccines or improved vaccines directed against flaviviruses such as yellow fever virus, dengue virus, JEV, SLEV and WNV which are inexpensive to prepare, present little risk to workers involved in their manufacture, carry minimal risk of adverse immunological reactions due to impurities or adventitious immunogenic components and are highly effective in eliciting neutralizing antibodies and protective immunity. There is furthermore a need for a vaccine against JEV, WNV and related flaviviruses that minimizes the number of immunizing doses required.

Many of the shortcomings of the current art as described in detail for the production of vaccines also apply to the production of antigens and antibodies to be used for the production of immunodiagnostics. Particularly, the concurrent risks and costs involved in the production of antigens from viruses and the failure of most currently available recombinantly expressed antigens to elicit effective immune responses are paralleled in the field of immunodiagnostics by the same risks, high costs and a corresponding lack of sensitivity. Thus, because of the high costs, risk of accidental infection with live virus and the lower than desired levels of sensitivity of

the previously available tests, there exists a need for rapid, simple and highly sensitive diagnostic tests for detecting flavivirus infection and/or contamination.

The present invention meets these needs by providing highly immunogenic recombinant antigens for use in diagnostic assays for the detection of antibodies to selected flaviviruses. The present invention further provides for the use of recombinant antigens derived from flaviviruses, flavivirus genes or mimetics thereof in immunodiagnostic assays for the detection of antibodies to flavivirus proteins.

Summary of the Invention

The present invention provides a nucleic acid molecule which contains a transcriptional unit (TU) for an immunogenic flavivirus antigen. The TU directs a host cell, after being incorporated within the cell, to synthesize the antigen. In an important aspect of the invention, the flavivirus can be yellow fever virus (YFV), dengue serotype 1 virus (DEN-1), dengue serotype 2 virus (DEN-2), dengue serotype 3 virus (DEN-3), dengue serotype 4 virus (DEN-4), St. Louis encephalitis virus (SLEV), Japanese encephalitis virus (JEV), West Nile virus (WNV), Powassan virus or any other flavivirus. In important embodiments of the present invention, the antigen can be the flavivirus prM/M protein, the E protein, or both. In particular, when the TU includes both the prM/M and E proteins, the host cell secretes subviral particles containing the prM/M and E antigens. In a further important aspect of the invention, the nucleic acid is a DNA molecule. In additional significant embodiments, the nucleic acid TU includes a control sequence disposed appropriately such that it operably controls the expression of the prM/M and E antigens and this control sequence can be the cytomegalovirus immediate early promoter. In an additional embodiment, the nucleotide sequence of the TU is engineered to optimize eukaryotic translation by minimizing large hairpin structures in the 5'-end untranslated region of an mRNA produced by the TU and/or the inclusion of a Kozak consensus sequence at the

translational start site of an mRNA produced by the TU. In an additional embodiment, the transcriptional unit also includes a poly-A terminator.

The present invention further provides a host cell comprising a nucleic acid molecule which includes a transcriptional unit for an immunogenic flavivirus antigen that directs the host cell to synthesize the immunogenic antigen. The flavivirus may be YFV, DEN-1, DEN-2, DEN-3, DEN-4, SLEV, JEV, WNV, Powassan virus or other flavivirus. In important embodiments, the antigen may be the prM/M protein, the E protein, or both the prM/M and the E proteins. In the latter case, the cell secretes subviral particles containing the prM/M and E antigens.

Additionally, the invention provides a composition for vaccinating a subject against a flavivirus containing a nucleic acid molecule that includes a transcriptional unit for an immunogenic flaviviral antigen. The transcriptional unit directs a cell within the body of the subject, after being incorporated therein, to synthesize the immunogenic antigen. The composition further includes a pharmaceutically acceptable carrier. In significant embodiments, the flavivirus may be YFV, DEN-1, DEN-2, DEN-3, DEN-4, SLEV, JEV, WNV, Powassan virus or other flavivirus. Furthermore, the antigen may be the prM/M protein, the E protein, or both the prM/M and the E proteins. In the latter instance, the cell secretes subviral particles comprising the flavivirus prM/M and E antigens. These subviral particles are also referred to as noninfectious recombinant antigen (NRA). In important embodiments, the nucleic acid molecule is a DNA molecule. In further significant embodiments, the transcriptional unit additionally contains a control sequence disposed appropriately such that it operably controls the synthesis of the prM/M and E antigens when the nucleic acid is introduced into the cell of the subject. This control sequence can be the cytomegalovirus immediate early promoter. In a still further embodiment, the transcriptional unit can also include a poly-A terminator.

The invention provides still further a method of immunizing a subject against infection by a flavivirus. The method involves administering to the subject an effective amount of a vaccinating composition that contains a nucleic acid molecule which includes a transcriptional unit for an immunogenic flavivirus antigen. The transcriptional unit directs a cell within the body of the subject, after being taken up by the cell, to synthesize the immunogenic antigen. The composition additionally includes a pharmaceutically acceptable carrier. In significant embodiments of the method, the flavivirus may be YFV, DEN-1, DEN-2, DEN-3, DEN-4, SLEV, JEV, WNV, Powassan virus or other flavivirus. In yet other important aspects of the method, the antigen may be the prM/M protein, the E protein, or both the prM/M and the E proteins. When the antigen is both the prM/M and the E proteins, the cell within the body of the subject, after incorporating the nucleic acid within it, secretes subviral particles comprising the flaviviral prM/M and E antigens. Additionally, in significant embodiments of the method, the vaccinating composition is administered to the subject in a single dose, via a parenteral route. In yet a further aspect of the method, the nucleic acid is a DNA molecule. In yet additional embodiments of the method, the transcriptional unit further includes a control sequence disposed appropriately such that it operably controls the synthesis of the prM/M and E antigens and in a significant aspect of this embodiment, the control sequence is the cytomegalovirus immediate early promoter. Furthermore, the transcriptional unit may include a poly-A terminator.

These aspects and embodiments of the invention are the basis for its distinct attributes and advantages. Being a nucleic acid construct involving only portions of the flavivirus genome rather than the sequence encompassing the complete genome, the nucleic acid TU-containing vaccine is completely nonviable. It therefore poses no danger of infection by the flavivirus to those involved in its manufacture or to subjects receiving the vaccine. The nucleic acid vaccine is easy to prepare and easy to administer and is stable in storage prior to use. Unexpectedly it has been found that the nucleic acid vaccine of the invention is essentially 100% successful in conferring protective immunity in mammals after administering only a single dose. A further

unexpected result is that the nucleic acid TU is able to engender immunity to a flavivirus in a female mammal which can be transmitted to its progeny through the milk. Without wishing to be limited by theory, the inventor believes that a possible mechanism for the success of the nucleic acid in conferring protective immunity is that a host cell harboring the nucleic acid, such as the cell of a subject to whom the vaccine is administered, produces subviral particles containing the flaviviral prM/M and E antigens. These particles mimic the immunogenic attributes of native flavivirus virions.

The present invention also provides noninfectious antigenic polypeptides, antigenic polypeptide fragments and NRA comprising the prM/M and/or E proteins of flaviviruses, wherein the transmembrane signal sequence is derived from a first flavivirus and the M and/or E proteins are derived from a second flavivirus. Further, the prM/M protein can comprise amino acid sequences from both the first and the second flaviviruses. "Chimeric" as used herein means any protein or nucleic acid comprising sequence from more than one flavivirus. As used herein, "non-virulent" means the antigen or vaccine of this invention is incapable of causing disease. More particularly, the recombinant protein antigens are free of contaminating genomic material from flaviviruses that is necessary for flavivirus infection, replication and pathogenesis.

The polypeptides of the present invention can comprise the amino acid sequences defined herein, or that are known in the art, of the prM, M and/or E proteins of selected flaviviruses. The nucleic acids of this invention can comprise nucleotide sequence that encodes the prM, M and/or E proteins of selected flaviviruses.

The antigens of the present invention can be unconjugated, or they can be conjugated to a carrier molecule that facilitates placement of the antigen on a solid phase. A carrier molecule is one to which antigens can be conjugated and which will

not react with antibodies in human serum. An example of such a carrier is bovine serum albumin (BSA).

The antigens of the present invention can also be recombinant proteins obtained by expressing nucleic acids encoding the antigen in an expression system capable of producing the antigen.

The amino acid sequences of the present antigens can contain an immunoreactive portion of the prM, M and/or E antigen. These antigens may further be attached to sequences designed to provide for some additional property, such as to remove/add amino acids capable of disulfide bonding to increase the reactivity of an epitope by providing a more rigid secondary structure, to increase its bio-longevity or to alter its cytotoxicity or to prevent infection. In any case, the antigen must possess immunoreactivity and/or immunogenicity.

Brief Description of the Drawings

Figure 1 is a schematic representation of flaviviral polyprotein processing. The central horizontal region provides a schematic representation of the viral genome. The lines denote the 5' and 3' non-translated regions and the boxed regions represent the open reading frame for structural (left and top) and non-structural (right and bottom) proteins. Cleavage by host cell signalase occurs simultaneously with translation at the E protein C-terminus, separating structural and non-structural regions. A subtilase-like cellular enzyme, furin, may be responsible for prM cleavage. Potential transmembrane domains of viral polyprotein are indicated by shaded areas.

Figure 2 is a map of the JEV genome (top) and the DNA sequence of oligonucleotides used in a reverse transcriptase-polymerase chain reaction (RT-PCR) (center) to construct the transcription unit for the expression of prM-E protein coding regions (bottom). Potential transmembrane domains of viral polyprotein are indicated by shaded areas.

Figure 3 shows a schematic representation of the plasmid vectors, pCDNA3, pCBamp, and pCIBamp, and the relationship between them. These plasmids include the CMV (cytomegalovirus) promoter/enhancer element, BGHp(A) (bovine growth hormone polyadenylation signal and transcription termination sequence), ampicillin resistance gene and ColE1 origin of replication for selection and maintenance in *E. coli*. The f1 origin of replication for single-stranded rescue in *E. coli* cells, SV40 origin of replication (SV40 ORI), neomycin resistance coding region and SV40p(A) sequences were deleted from pCDNA3 to generate pCBamp. An intron sequence was inserted in the NcoI-KpnI site of pCBamp to generate plasmid pCIBamp.

Figure 4 shows SDS-PAGE-immunoblot analyses of the sucrose gradient purified subviral particles from JE-4B COS-1 culture fluid (4B, right lane of each pair). The density gradient purified JE virion from JEV infected C6/36 cell culture was used as a positive control (JEV, left lane of each pair). JE HIAF (hyperimmune ascitic fluid); 4G2, anti-E monoclonal antibody; JM01, anti-M monoclonal antibody; NMAF (normal mouse ascitic fluid).

Figure 5 shows a profile of the E antigen in a rate zonal sucrose gradient analysis prepared from the PEG precipitate of JE-4B cell culture medium with or without Triton X-100 treatment.

Detailed Description of the Invention

The invention encompasses nucleic acid transcriptional units which encode flaviviral antigenic proteins, such as the prM/M and E protein antigens. The nucleic acids function to express the prM/M and E protein antigens when the nucleic acid is taken up by an appropriate cell, especially when the cell is the cell of a subject. The invention also encompasses a vaccine whose active agent is the nucleic acid transcriptional unit (TU). The invention further encompasses cells containing a TU. The invention in addition encompasses a method of immunizing a subject against

flaviviral infection by administering to the subject an effective amount of a vaccine containing the nucleic acid TU molecules.

The invention provides an isolated nucleic acid comprising a transcriptional unit encoding a signal sequence of a structural protein of a first flavivirus and an immunogenic flavivirus antigen of a second flavivirus, wherein the transcriptional unit directs the synthesis of the antigen. The invention further encompasses the use of the nucleic acid transcriptional unit (TU) to generate flaviviral antigens and the flaviviral antigens produced by the nucleic acid TU. The invention still further encompasses the use of the flaviviral antigens encoded by the TU of the invention to produce flavivirus-specific antibodies and to detect the presence of flavivirus-specific antibodies.

In one embodiment, the isolated nucleic acid of this invention can comprise a transcriptional unit encoding a Japanese encephalitis virus signal sequence.

In another embodiment, the transcriptional unit of this invention can encode an immunogenic flavivirus antigen which can be from one or more of the following flaviviruses: yellow fever virus, dengue serotype 1 virus, dengue serotype 2 virus, dengue serotype 3 virus, dengue serotype 4 virus, Japanese encephalitis virus, Powassan virus and West Nile virus.

In a particular embodiment, the nucleic acid of this invention can encode a signal sequence of Japanese encephalitis virus and an M protein and an E protein of West Nile virus, SLEV, YFV and/or Powassan virus. The nucleic acid can also encode an immunogenic antigen which can be an M protein of a flavivirus, an E protein of a flavivirus, both an M protein and an E protein of a flavivirus, a portion of an M protein of a flavivirus, a portion of an E protein of a flavivirus and/or both a portion of an M protein of a flavivirus and a portion of an E protein of a flavivirus. In a preferred embodiment, the isolated nucleic acid encodes both the M protein and the E protein of the flavivirus. Further, the nucleic acid of the invention can be DNA and can comprise

nucleotide sequence SEQ ID NO:15, SEQ ID NO:19, SEQ ID NO:21 or SEQ ID NO:23.

The transcriptional unit of this invention can also comprise a control sequence disposed appropriately so that it operably controls the synthesis of the antigen. The control sequence can be, for example, the cytomegalovirus immediate early promoter. The nucleic acid of this invention can also comprise a Kozak consensus sequence located at a translational start site for a polypeptide comprising the antigen encoded by the transcriptional unit. The transcriptional unit of this invention can also comprise a poly-A terminator.

The present invention further provides a cell comprising the nucleic acid of this invention.

Also provided is a composition comprising a pharmaceutically acceptable carrier and nucleic acid or cell or antigen of this invention. The present invention additionally provides a method of immunizing a subject against infection by a flavivirus, comprising administering to the subject an effective amount of a composition of this invention. In a particular embodiment, the composition used to immunize a subject directs the synthesis of both the M protein and the E protein of a flavivirus and a cell within the body of the subject, after incorporating the nucleic acid within it, secretes subviral particles comprising the M protein and the E protein. Alternatively, the composition can comprise an M protein and/or E protein of a flavivirus or subviral particles comprising the M protein and E protein. In the methods of this invention, the immunizing composition can be administered to the subject in a single dose and can be administered via a parenteral route.

This invention further provides the antigens produced from the isolated nucleic acids of this invention. As an example, the antigen from the second flavivirus encoded by the nucleotide sequence of TU can be the M protein which can be, for example, from

West Nile virus. The antigen can also be protein from dengue virus, St. Louis encephalitis virus, Japanese encephalitis virus, Powassan virus and/or yellow fever virus. In a further embodiment, the antigen comprises a prM/M protein comprising the transmembrane signal sequence from a first flavivirus and further amino acid sequence comprising the remainder of the prM/M protein from a second flavivirus, which can be from SLEV, JEV, YFV, WNV and/or Powassan virus.

The antigen encoded by the nucleotide sequence of the TU can be West Nile virus antigen, dengue virus antigen, St. Louis encephalitis virus antigen, Japanese encephalitis virus antigen, Powassan virus antigen and/or yellow fever virus antigen.

The antigen encoded by the nucleotide sequence of the TU can also be the E protein, which can be the E protein from West Nile virus, dengue virus, St. Louis encephalitis virus, Japanese encephalitis virus, Powassan virus and/or yellow fever virus.

Additionally, the antigen encoded by the nucleotide sequence of the TU can be the M protein and the E protein, which can be from West Nile virus, dengue virus, St. Louis encephalitis virus, Japanese encephalitis virus, Powassan virus and/or yellow fever virus.

As used herein, "M protein" or "pr/M protein" or "prM/M protein" means a flavivirus M protein or flavivirus prM protein. Examples include, but are not limited to, prM proteins comprising amino acid sequence from one or more flavivirus prM proteins, M proteins comprising no additional amino acid sequence and proteins comprising additional amino acid sequences which are processed *in vitro* or *in vivo* to generate the mature M protein.

As used herein, "nucleic acid transcriptional unit" or "nucleic acid transcriptional unit molecule" means a nucleic acid encoding one or more specified

proteins. The TU has biological activity such that, after having been introduced into a suitable cell, the nucleic acid induces the synthesis of one or more specified gene products encoded by the nucleic acid. The gene product(s) is(are) other biological macromolecules, such as proteins, not chemically related to the TU. The nucleic acid TU induces the cell to employ its cellular components to produce the specific gene product or products encoded by the nucleic acid of the TU. Although any nucleic acid may serve as a TU, in a preferred embodiment, the TU is the DNA of a plasmid or similar vector, wherein the plasmid or vector comprises coding sequences of marker genes or other sequence constructions that facilitate use of the TU for experimentation and biosynthesis.

As used herein, a "control sequence" is a regulatory nucleotide sequence incorporated within a TU which interacts with appropriate cellular components of the cell and leads to enhanced or activated biosynthesis of the gene products encoded by the TU. Thus a suitable control sequence is one with which the components of the cell have the capability to interact, resulting in synthesis of the gene product. When operably disposed in a nucleic acid with respect to a specified coding sequence, a control sequence effectively controls expression of the specified nucleic acid to produce the gene product.

As used herein, a "promoter" is a nucleotide sequence in a TU which serves as a control sequence.

As used herein, a "Kozak sequence" or "Kozak consensus sequence" is a nucleotide sequence at the translational start site which optimizes translation of eukaryotic mRNAs (Kozak, *Mol. Cell. Biology* 9: 5134-5142 (1989)).

As used herein, a "terminator" is an extended nucleotide sequence which acts to induce polyadenylation at the 3' end of a mature mRNA. A terminator sequence is found after, or downstream from, a particular coding sequence.

As used herein, a "cell" is a prokaryotic or eukaryotic cell comprising a TU coding for one or more gene products, or into which such a TU has been introduced. Thus, a cell harbors a foreign or heterologous substance, the TU, which is not naturally or endogenously found in the cell as a component. A suitable cell is one which has the capability for the biosynthesis of the gene products as a consequence of the introduction of the TU. In particular, a suitable cell is one which responds to a control sequence and to a terminator sequence, if any, that may be included within the TU. In important embodiments of the present invention, the cell is a mammalian cell. In particularly important embodiments of this invention, the cell is a naturally occurring cell in the body of a human or nonhuman subject to whom (which) the TU has been administered as a component of a vaccine. Alternatively, in analytical, or diagnostic applications, including preparation of antigen for use as a vaccine or in immunodiagnostic assays, or for demonstrative purposes, the cell may be a human or nonhuman cell cultured *in vitro*.

As used herein, a "vaccine" or a "composition for vaccinating a subject" specific for a particular pathogen means a preparation, which, when administered to a subject, leads to an immunogenic response in a subject. As used herein, an "immunogenic" response is one that confers upon the subject protective immunity against the pathogen. Without wishing to be bound by theory, it is believed that an immunogenic response may arise from the generation of neutralizing antibodies (i.e., a humoral immune response) or from cytotoxic cells of the immune system (i.e., a cellular immune response) or both. As used herein, an "immunogenic antigen" is an antigen which induces an immunogenic response when it is introduced into a subject, or when it is synthesized within the cells of a host or a subject. As used herein, an "effective amount" of a vaccine or vaccinating composition is an amount which, when administered to a subject, is sufficient to confer protective immunity upon the subject. Historically, a vaccine has been understood to contain as an active principle one or more specific molecular components or structures which comprise the pathogen, especially its surface. Such structures may include surface components such as

proteins, complex carbohydrates, and/or complex lipids which commonly are found in pathogenic organisms.

As used herein, however, it is to be stressed that the terms "vaccine" or "composition for vaccinating a subject" extend the conventional meaning summarized in the preceding paragraph. As used herein, these terms also relate to the TU of the instant invention or to compositions containing the TU. The TU induces the biosynthesis of one or more specified gene products encoded by the TU within the cells of the subject, wherein the gene products are specified antigens of a pathogen. The biosynthetic antigens then serve as an immunogen. As already noted, the TU, and hence the vaccine, may be any nucleic acid that encodes the specified immunogenic antigens. In a preferred embodiment of this invention, the TU of the vaccine is DNA. The TU can include a plasmid or vector incorporating additional genes or particular sequences for the convenience of the skilled worker in the fields of molecular biology, cell biology and viral immunology (See Molecular Cloning: A Laboratory Manual, 2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989; and Current Protocols in Molecular Biology, Ausubel et al., John Wiley and Sons, New York 1987 (updated quarterly), which are incorporated herein by reference).

The TU molecules of the instant invention comprise nucleic acids, or derivatives of nucleic acids, having nucleotide sequences that encode specific gene products related to antigens of flaviviruses such as, but not limited to, WNV, JEV, dengue virus, yellow fever virus and SLEV. Although any nucleic acid may serve as a TU, in an important embodiment, the TU is DNA. Alternatively, the nucleic acids may be RNA molecules. They may also be any one of several derivatives of DNA or RNA having a backbone of phosphodiester bonds that have been chemically modified to increase the stability of the TU as a pharmaceutical agent. Modifications so envisioned include, but are not limited to, phosphorothioate derivatives or phosphonate derivatives.

These and other examples of derivatives are well known to persons skilled in the field of nucleic acid chemistry.

The genome of JEV has been characterized and sequenced (Figures 1 and 2). The M structural protein is expressed as a portion of the polyprotein which includes a pre-M sequence (pr). This pr sequence, immediately amino terminal to the M protein sequence, prevents conformational problems in the processing of the polyprotein. In particular, the presence of the pr sequence is important in preventing misfolding of the E protein. Thus, the presence of prM allows for assembly of JEV particles. Once the virion or particle is formed, the pr sequence can be cleaved from the prM protein to yield mature virus particles containing M proteins, although cleavage of the prM protein to yield M protein is not necessary to produce infectious particles. The prM sequences from many different, related flaviviruses are cleaved to but a low extent, but the flaviviruses themselves are nonetheless, infectious. Examples of such related flaviviruses with similar genomic structures and functions include, but are not limited to WNV, YFV, dengue virus and SLEV.

In one embodiment, the TU encoding flaviviral M and E proteins in the instant invention is DNA. In accord with the discussion in the preceding paragraph, this DNA comprises a nucleotide sequence which encodes the M protein, comprising the pre-M sequence, and a nucleotide sequence encoding the E protein. In this way, the intended gene products are enabled to form subviral particles within the cell. The pre-M sequence can then be cleaved in a fashion analogous to that which occurs with respect to replete virions.

In order to function effectively *in vivo* as a vaccine, it is advantageous to include within the TU a control sequence that has the effect of enhancing or promoting the transcription of the nucleotide sequences encoding the antigens. Use of such promoters is well known to those of skill in the fields of molecular biology, cell biology and viral immunology (See Molecular Cloning: A Laboratory Manual, 2nd Ed., Sambrook,

Fritsch and Maniatis, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989; and Current Protocols in Molecular Biology, Ausubel et al., John Wiley and Sons, New York 1987 (updated quarterly)). When the TU is used as a vaccine in a mammalian host, the promoter to be employed is preferably one which operates effectively in mammalian cells. Such a promoter is disposed with respect to the coding sequences from which transcription is to be promoted, at a position at which it may operably promote such transcription. In a significant embodiment of the instant invention, this promoter is the cytomegalovirus early promoter. In addition, in a further preferred embodiment of the invention, the coding sequences are followed, in the TU nucleic acid, by a terminator sequence (Sambrook et al.). Particular embodiments of the invention relate to both prokaryotic and eukaryotic cells. Many promoter sequences are known that are useful in either prokaryotic or eukaryotic cells. (See Sambrook et al.)

The nucleic acids of the invention may further include DNA sequences known to those of skill in the art to act as immunostimulatory elements. Examples of such elements include, but are not limited to, certain CpG motifs in bacterial DNA (Sato et al., *Science* 273: 352-354 (1996); Klinman et al., *Vaccine* 17: 19-25 (1998)).

Preparation of the TU of the invention is readily accomplished by methods well known to workers of skill in the field of molecular biology. Procedures involved are set forth, for example, in Molecular Cloning: A Laboratory Manual, 2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989 and Current Protocols in Molecular Biology, Ausubel et al., John Wiley and Sons, New York 1987 (updated quarterly). The flaviviral RNA molecule may be isolated from a sample of live virus by methods widely known among virologists familiar with flaviviruses, for example, and with other groups of viruses as well. Methods used with JEV are summarized in Kuno et al. (*J. Virol.* 72: 73-83 (1998)). The RNA is used as a template for the synthesis of cDNA using reverse transcriptase. From the cDNA, a fragment containing the pre-M through E coding region (Figure 2) is obtained by digestion with restriction nucleases known to cleave the cDNA

appropriately to provide such fragments. Examples of restriction digestion of JEV are provided in Nitayaphan et al. (1990) and Konishi et al. (1991). Incorporation of promoters, such as the cytomegalovirus promoter, sequences to promote efficient translation, such as the Kozak sequence, and of the polyadenylation signal, is likewise well known to skilled practitioners in molecular biology and recombinant DNA engineering (Kozak, *Mol. Cell. Biology* 9: 5134-5142 (1989); Azevedo et al., *Braz. J. Med. Biol. Res.* 32: 147-153 (1999)). When a nucleic acid comprising a TU containing the desired coding sequences and control sequences is prepared, it may be obtained in larger quantities by methods that amplify nucleic acids. Such methods are widely known to workers skilled in molecular biology and recombinant DNA engineering. Examples of these methods include incorporation of the nucleic acid into a plasmid for replication by culturing in a cell such as a prokaryotic cell and harvesting the plasmid after completing the culture, as well as amplification of the nucleic acid by methods such as PCR and other amplification protocols, as are well known in the art. These examples are not intended to limit the ways in which the nucleic acid containing the TU may be obtained.

The TU-containing nucleic acid molecules of the instant invention may be introduced into appropriate cells in many ways well known to skilled workers in the fields of molecular biology and viral immunology. By way of example, these include, but are not limited to, incorporation into a plasmid or similar nucleic acid vector which is taken up by the cells, or encapsulation within vesicular lipid structures such as liposomes, especially liposomes comprising cationic lipids, or adsorption to particles that are incorporated into the cell by endocytosis.

In general, a cell of this invention is a prokaryotic or eukaryotic cell comprising a TU, or into which a TU has been introduced. The TU of the present invention induces the intracellular biosynthesis of the encoded prM/M and E antigens. A suitable cell is one which has the capability for the biosynthesis of the gene products as a consequence of the introduction of the nucleic acid. In particular embodiments of the

invention, a suitable cell is one which responds to a control sequence and to a terminator sequence, if any, which may be included within the TU. In order to respond in this fashion, such a cell contains within it components which interact with a control sequence and with a terminator and act to carry out the respective promoting and terminating functions. When the cell is cultured *in vitro*, it may be a prokaryote, a single-cell eukaryote or a multicellular eukaryote cell. In particular embodiments of the present invention, the cell is a mammalian cell. In these cases, the synthesized prM/M and E protein gene products are available for use in analytical, or diagnostic applications, including preparation of antigen for use as a vaccine or in immunodiagnostic assays, or for demonstrative purposes.

In some circumstances, such as when the cell is a cultured mammalian cell, the prM/M and E antigens are secreted in the form of subviral particles. These are aggregates of prM/M and E proteins resembling live virus in surface ultrastructural morphology and immunogenic properties. Since the TU of the invention does not include the remainder of the flaviviral genome, however, there is no capsid incorporated, and most importantly, no infectious viral RNA.

In another important embodiment of this invention, the cell is a natural cellular component of the subject to whom the TU has been administered as a vaccine. The TU, when administered to the subject, is taken up by the cells of the subject. The subject's cells have the capability of responding to any promoter sequences, and terminator, if present. In any case, the TU induces the subject's cells to synthesize flaviviral prM/M and E gene products. Without wishing to be constrained by theoretical considerations, it is believed that the subject's cells produce subviral particles *in vivo* consisting of the prM/M and E antigens, just as has been found to occur with cultured mammalian cells *in vitro*. Such subviral particles, it is believed, then serve as the *in vivo* immunogen, stimulating the immune system of the subject to generate immunological responses which confer protective immunity on the subject. Again without wishing to be limited by theory, the resulting protective immunity may

arise via either humoral or cellular immunity, i.e., via either an MHC class II- or class I-restricted mechanism, respectively, or by both mechanisms.

According to the invention, subjects are immunized against infection by flaviviruses, such as JEV, YFV, dengue virus, SLEV, WNV or other flaviviruses by administering to them an effective amount of a TU comprising nucleic acid which encodes the prM and/or E antigens. The nucleic acid, after being incorporated into the cells of the subject, leads to the synthesis of the flaviviral prM/M and/or E antigens.

In order to administer the TU to the subject, it is incorporated into a composition which comprises a pharmaceutically acceptable carrier. The term "pharmaceutically acceptable" means a material that is not biologically or otherwise undesirable, i.e., the material may be administered to an subject along with the immunogenic material (i.e., recombinant flavivirus protein antigens or portions thereof) without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the vaccine in which it is contained. Examples of pharmaceutically acceptable carriers, or components thereof, include water, physiological saline and common physiological buffers (for further examples, see Arnon, R. (Ed.) Synthetic Vaccines I: pp. 83-92, CRC Press, Inc., Boca Raton, Florida, 1987).

It is understood by those skilled in the art that the critical value in describing a vaccination dose is the total amount of immunogen needed to elicit a protective response in a host which is subject to infectious disease caused by virulent or wild-type flavivirus infection. The number and volume of doses used can be varied and are determined by the practitioner based on such parameters as, age, weight, gender, species, type of vaccine to be administered, mode of administration, overall condition of the subject, et cetera, as well as other important factors recognized by those of skill in the art.

The TU may be administered to a subject orally, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, transdermally, extracorporeally, intranasally, topically or the like. Delivery can also be directly to any area of the respiratory system (e.g., lungs) via intubation. The exact amount of the TU required will vary from subject to subject, depending on the species, age, weight and general condition of the subject, the immunogenicity of the vaccine used, the strain or species of flavivirus against which the subject is being immunized, the mode of administration and the like. Thus, it is not possible to specify an exact amount for every embodiment of the present invention. However, an appropriate amount can be determined by one of ordinary skill in the art using only routine experimentation given the teachings herein and what is available in the art.

Parenteral administration of the vaccine of the present invention, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution or suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant dosage is maintained. See, e.g., U.S. Patent No. 3,610,795, which is incorporated by reference herein.

For solid compositions, conventional nontoxic solid carriers include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talc, cellulose, glucose, sucrose, magnesium carbonate, and the like. Liquid pharmaceutically administrable compositions can, for example, be prepared by dissolving, dispersing, etc. an active compound as described herein and optional pharmaceutical adjuvants in an excipient, such as, for example, water, saline, aqueous dextrose, glycerol, ethanol, and the like, to thereby form a solution or suspension. If desired, the pharmaceutical composition to be administered may also contain minor amounts of nontoxic auxiliary substances such as wetting or emulsifying agents, pH buffering agents and the like, for example, sodium acetate, sorbitan monolaurate,

triethanolamine sodium acetate, triethanolamine oleate, etc. Actual methods of preparing such dosage forms are known, or will be apparent, to those skilled in this art; for example, see Remington's Pharmaceutical Sciences (Martin, E.W. (ed.), latest edition, Mack Publishing Co., Easton, PA).

In one embodiment, the TU of this invention can be administered to the subject by the use of electrotransfer mediated *in vivo* gene delivery, wherein immediately following administration of the TU to the subject, transcutaneous electric pulses are applied to the subject, providing greater efficiency and reproducibility of *in vivo* nucleic acid transfer to tissue in the subject (Mir et al., *Proc. Nat. Acad. Sci USA* 96: 4262-4267 (1999)).

In the methods of the present invention which describe the immunization of a subject by administering a vaccine of this invention to a subject, the efficacy of the immunization can be monitored according the clinical protocols well known in the art for monitoring the immune status of a subject.

An effective amount of a vaccinating composition is readily determined by those of skill in the art to be an amount which, when administered to a subject, confers protective immunity upon the subject. In order to undertake such a determination, the skilled artisan can assess the ability to induce flaviviral prM/M- and E-specific antibodies and/or flaviviral prM/M- and E-specific cytotoxic T lymphocytes present in the blood of a subject to whom the vaccine has been administered. One can also determine the level of protective immunity conferred upon an experimental subject by challenge with live flavivirus corresponding to the antigenic composition used to immunize the experimental subject. Such challenge experiments are well known to those of skill in the art.

In general, in order to immunize a subject against infection by WNV, JEV, YFV, dengue virus, SLEV, or other flaviviruses according to the present invention, and

recognizing that the TUs employed in such methods may have differing overall sizes, doses ranging from about 0.1 μg/kg body weight to about 50 μg/kg body weight can be used.

It has unexpectedly been found that a TU of the present invention which is a DNA confers protective immunity at a level of effectiveness approximating 100% after administration of only a single effective dose of the TU by i.m. injection or by electrotransfer. This is in contrast to many immunization methods carried out using conventional vaccines (as described above), which require one or more booster vaccinations and which may not confer protective immunity to an effectiveness near 100%.

It has further been found unexpectedly that protective immunity may be transmitted from a vaccinated female subject to the offspring of the subject. A significant proportion of neonatal mice was shown to be protected against viral challenge after the mothers were vaccinated using the TU DNA of the invention. Without wishing to be limited by theory, it is known that passive immunity may be conferred on neonatal mammals due to the presence in maternal milk of neutralizing antibodies specific for various pathogens. It is possible that the protective immunity against JEV found within the neonates was transmitted to them in this way.

In another embodiment of the invention, the TU encodes a signal sequence of a structural protein of a first flavivirus and an immunogenic flavivirus antigen of a second flavivirus. Thus, in one embodiment, for example, the signal sequence of structural protein of a first flavivirus is replaced by a signal sequence of structural protein of a second flavivirus, which results in proper folding of the nascent polypeptide, proper processing in a host, and/or proper folding of the processed protein. In another embodiment of the invention, the TU may encode an immunogenic flavivirus antigen wherein the antigen comprises sequence from one or more than one flavivirus.

The present invention further provides immunogenic compositions comprising the polypeptides of this invention in a pharmaceutical acceptable carrier for use as a protein vaccine. Antigens produced from the transcriptional units of the present invention can be used to elicit effective immune responses in a subject. Antigens for this purpose can comprise flavivirus prM protein, flavivirus M protein, flavivirus E protein or any combination thereof, including immunogenic fragments of the proteins. A particularly preferred embodiment is the use of the NRA described herein. A further preferred embodiment is a chimeric protein comprising the signal sequence of one flavivirus and the structural protein(s) of one or more different flaviviruses. In a particularly preferred embodiment, the signal sequence of the antigen is the Japanese encephalitis virus signal sequence.

In other embodiments, the protein vaccine of this invention further comprises a suitable adjuvant. As used herein, an "adjuvant" is a potentiator or enhancer of the immune response. The term "suitable " is meant to include any substance which can be used in combination with the vaccine immunogen (i.e., flavivirus prM protein, flavivirus M protein, flavivirus E protein, or any combination thereof) to augment the immune response, without producing adverse reactions in the vaccinated subject. Effective amounts of a specific adjuvant may be readily determined so as to optimize the potentiation effect of the adjuvant on the immune response of a vaccinated subject. In a preferred embodiment, adjuvanting of the vaccines of this invention is a 2 – stage process, utilizing first a 2% aluminum hydroxide solution and then a mineral oil. In specific embodiments, suitable adjuvants can be chosen from the following group: mineral, vegetable or fish oil with water emulsions, incomplete Freund's adjuvant, *E. coli* J5, dextran sulfate, iron oxide, sodium alginate, Bacto-Adjuvant, certain synthetic polymers such as Carbopol (BF Goodrich Company, Cleveland, Ohio), poly-amino acids and co-polymers of amino acids, saponin, carrageenan, REGRESSIN (Vetrepharm, Athens, GA), AVRIDINE (N, N-dioctadecyl-N',N'-bis(2-hydroxyethyl)-propanediamine), long chain polydispersed ß (1,4) linked mannan polymers interspersed with O-acetylated groups (e.g. ACEMANNAN), deproteinized highly

purified cell wall extracts derived from non-pathogenic strain of Mycobacterium species (e.g. EQUIMUNE, Vetrepharm Research Inc., Athens GA), Mannite monooleate, paraffin oil and muramyl dipeptide.

In another aspect, this invention provides a method for immunizing subjects with immunogenic amounts of the protein vaccine of the invention to elicit an effective immune response in the subject. Immunization can be carried out orally, parenterally, intranasally, intratracheally, intramuscularly, intramammarily, subcutaneously, intravenously and/or intradermally. The vaccine containing the flavivirus prM protein, flavivirus M protein and/or the flavivirus E protein can be administered by injection, by inhalation, by ingestion, or by infusion. A single dose can be given and/or repeated doses of the vaccine preparations, i.e. "boosters," can be administered at periodic time intervals to enhance the initial immune response or after a long period of time since the last dose. The time interval between vaccinations can vary, depending on the age and condition of the subject.

The term " immunogenic amount" means an amount of an immunogen, or a portion thereof, which is sufficient to induce an immune response in a vaccinated subject and which protects the subject against disease caused by wild-type or virulent flavivirus infections upon exposure thereto or which has a therapeutic or commercially beneficial effect that lessens the effect of flavivirus infection on the vaccinated subject.

The invention further provides an antibody produced in response to immunization by the antigen of this invention. The antibodies of the present invention can include polyclonal and monoclonal antibodies which can be intact immunoglobulin molecules, chimeric immunoglobulin molecules, "humanized antibodies," or Fab or $F(ab')_2$ fragments. Such antibodies and antibody fragments can be produced by techniques well known in the art which include those described in Harlow and Lane (Antibodies: A Laboratory Manual. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989) and Kohler et al. (*Nature* 256:495-97, 1975) and U.S. Patents

5,545,806, 5,569,825 and 5,625,126, incorporated herein by reference. The antibodies can be of any isotype IgG, IgA, IgD, IgE and IgM.

The present invention can also include single chain antibodies (ScFv), comprising linked V_H and V_L domains and which retain the conformation and specific binding activity of the native idiotype of the antibody. Such single chain antibodies are well known in the art and can be produced by standard methods. (see, e.g., Alvarez et al., *Hum. Gene Ther.* 8: 229-242 (1997)).

Antibodies can be produced against the antigens of this invention which are synthesized from nucleic acid sequences encoding immunogenic amino acid sequences of the prM, M and/or E antigens of one or more flaviviruses and the signal sequence of a different flavivirus (e.g., JEV). Immunogenic peptides synthesized from the use of these chimeric constructs can easily be identified by use of methods well known in the art for identifying immunogenic regions in an amino acid sequence and used to produce the antibodies of this invention.

Conditions whereby an antigen/antibody complex can form, as well as assays for the detection of the formation of an antigen/antibody complex and quantitation of the detected protein, are standard in the art. Such assays can include, but are not limited to, Western blotting, immunoprecipitation, immunofluorescence, immunocytochemistry, immunohistochemistry, fluorescence activated cell sorting (FACS), fluorescence *in situ* hybridization (FISH), immunomagnetic assays, ELISA, ELISPOT (Coligan et al., eds. 1995. Current Protocols in Immunology. Wiley, New York.), agglutination assays, flocculation assays, cell panning, etc., as are well known to the artisan.

As used herein, the term "bind" means the well characterized binding of antibody to antigen as well as other nonrandom association with an antigen. "Specifically bind" as used herein describes an antibody or other ligand that does not

cross react substantially with any antigen other than the one specified, which in this case, is an antigen of this invention.

The antibody or ligand of this invention can be bound to a substrate (e.g., beads, tubes, slides, plates, nitrocellulose sheets, etc.) or conjugated with a detectable moiety or both bound and conjugated. The detectable moieties contemplated for the present invention can include, but are not limited to, an immunofluorescent moiety (e.g., fluorescein, rhodamine), a radioactive moiety (e.g., ^{32}P, ^{125}I, ^{35}S), an enzyme moiety (e.g., horseradish peroxidase, alkaline phosphatase), a colloidal gold moiety and a biotin moiety. Such conjugation techniques are standard in the art (for example, Harlow and Lane, Antibodies: A Laboratory Manual. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1989); Yang et al., *Nature* 382: 319-324 (1996)).

The present invention further provides a method of detecting flavivirus antibody in a sample, comprising contacting the sample with the flavivirus antigen of the present invention, under conditions whereby an antigen/antibody complex can form; and detecting formation of the complex, thereby detecting flavivirus antibody in the sample.

The present invention further provides a method of detecting flavivirus antigen in a sample, comprising contacting the sample with an antibody of this invention under conditions whereby an antigen/antibody complex can form; and detecting formation of the complex, thereby detecting flavivirus antigen in the sample.

The method of detecting flavivirus antigen in a sample can be performed, for example, by contacting a fluid or tissue sample from a subject with an antibody of this invention and detecting binding of the antibody to the antigen. It is contemplated that the antigen will be on an intact flavivirus virion, will be a flavivirus-encoded protein displayed on the surface of a flavivirus-infected cell expressing the antigen, or will be a fragment of the antigen. A fluid sample of this method can comprise any biological fluid which could contain the antigen or a cell containing the antigen, such as

cerebrospinal fluid, blood, bile, plasma, serum, saliva and urine. Other possible examples of body fluids include sputum, mucus and the like.

The method of detecting flavivirus antibody in a sample can be performed, for example, by contacting a fluid or tissue sample from a subject with an antigen of this invention and detecting the binding of the antigen to the antibody. A fluid sample of this method can comprise any biological fluid which could contain the antibody, such as cerebrospinal fluid, blood, bile, plasma, serum, saliva and urine. Other possible examples of body fluids include sputum, mucus and the like.

Enzyme immunoassays such as immunofluorescence assays (IFA), enzyme linked immunosorbent assays (ELISA) and immunoblotting can be readily adapted to accomplish the detection of flavivirus antibodies according to the methods of this invention. An ELISA method effective for the detection of the antibodies can, for example, be as follows: (1) bind the antigen to a substrate; (2) contact the bound antigen with a fluid or tissue sample containing the antibody; (3) contact the above with a secondary antibody bound to a detectable moiety which is reactive with the bound antibody (e.g., horseradish peroxidase enzyme or alkaline phosphatase enzyme); (4) contact the above with the substrate for the enzyme; (5) contact the above with a color reagent; and (6) observe/measure color change or development.

Another immunologic technique that can be useful in the detection of flavivirus antibodies uses monoclonal antibodies (MAbs) for detection of antibodies specifically reactive with flavivirus antigens in a competitive inhibition assay. Briefly, sample is contacted with an antigen of this invention which is bound to a substrate (e.g., an ELISA 96-well plate). Excess sample is thoroughly washed away. A labeled (e.g., enzyme-linked, fluorescent, radioactive, etc.) monoclonal antibody is then contacted with any previously formed antigen-antibody complexes and the amount of monoclonal antibody binding is measured. The amount of inhibition of monoclonal antibody binding is measured relative to a control (no antibody), allowing for detection and

measurement of antibody in the sample. The degree of monoclonal antibody inhibition can be a very specific assay for detecting a particular flavivirus variety or strain, when based on monoclonal antibody binding specificity for a particular variety or strain of flavivirus. MAbs can also be used for direct detection of flavivirus antigens in cells by, for example, immunofluorescence assay (IFA) according to standard methods.

As a further example, a micro-agglutination test can be used to detect the presence of flavivirus antibodies in a sample. Briefly, latex beads, red blood cells or other agglutinable particles are coated with the antigen of this invention and mixed with a sample, such that antibodies in the sample that are specifically reactive with the antigen crosslink with the antigen, causing agglutination. The agglutinated antigen-antibody complexes form a precipitate, visible with the naked eye or measurable by spectrophotometer. In a modification of the above test, antibodies of this invention can be bound to the agglutinable particles and antigen in the sample thereby detected.

The present invention further provides a method of diagnosing a flavivirus infection in a subject, comprising contacting a sample from the subject with the antigen of this invention under conditions whereby an antigen/antibody complex can form; and detecting antigen/antibody complex formation, thereby diagnosing a flavivirus infection in a subject.

The present invention further provides a method of diagnosing a flavivirus infection in a subject, comprising contacting a sample from the subject with the antibody of this invention under conditions whereby an antigen/antibody complex can form; and detecting antigen/antibody complex formation, thereby diagnosing a flavivirus infection in a subject.

In the diagnostic methods taught herein, the antigen of this invention can be bound to a substrate and contacted with a fluid sample such as blood, serum, urine or saliva. This sample can be taken directly from the patient or in a partially purified

form. In this manner, antibodies specific for the antigen (the primary antibody) will specifically react with the bound antigen. Thereafter, a secondary antibody bound to, or labeled with, a detectable moiety can be added to enhance the detection of the primary antibody. Generally, the secondary antibody or other ligand, which is reactive, either specifically with a different epitope of the antigen or nonspecifically with the ligand or reacted antibody, will be selected for its ability to react with multiple sites on the primary antibody. Thus, for example, several molecules of the secondary antibody can react with each primary antibody, making the primary antibody more detectable.

The detectable moiety allows for visual detection of a precipitate or a color change, visual detection by microscopy, or automated detection by spectromctry, radiometric measurement or the like. Examples of detectable moieties include fluorescein and rhodamine (for fluorescence microscopy), horseradish peroxidase (for either light or electron microscopy and biochemical detection), biotin-streptavidin (for light or electron microscopy) and alkaline phosphatase (for biochemical detection by color change).

Particular embodiments of the present invention are set forth in the examples which follow. These examples are not intended to limit the scope of the invention as disclosed in this specification.

Examples

General methods utilizing molecular biology and recombinant DNA techniques related to preparing and expressing the nucleic acid TU molecules of the invention are set forth in, for example, Current Protocols in Molecular Biology, Ausubel et al., John Wiley and Sons, New York 1987 (updated quarterly), and Molecular Cloning: A Laboratory Manual 2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989.

Example 1. Preparation of recombinant plasmids containing the transcriptional unit encoding JEV prM and E antigens. Genomic RNA was extracted from 150 μL of JEV strain SA 14 virus seed grown from mouse brain using a QIAamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA). RNA, adsorbed on a silica membrane, was eluted in 80 μL of nuclease-free water, and used as a template for the amplification of JEV prM and E gene coding sequences. Primer sequences were obtained from the work of Nitayaphan et al. (*Virology* 177: 541-552 (1990)). A single cDNA fragment containing the genomic nucleotide region 389-2478 was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction sites KpnI and XbaI, the consensus Kozak ribosomal binding sequence, and the translation initiation site were engineered at the 5' terminus of the cDNA by amplimer 14DV389 (nucleotide sequence, SEQ ID NO:1; amino acid sequence, SEQ ID NO:2). An in-frame translation termination codon, followed by a NotI restriction site, was introduced at the 3' terminus of the cDNA by amplimer c14DV2453 (SEQ ID NO:3) (Figure 2). One-tube RT-PCR was performed using a Titan RT-PCR Kit (Boehringer Mannheim, Indianapolis, IN). 10 μL of viral RNA was mixed with 1 μL each of 14DV389 (50 μM) and c14DV2453 (50 μM) and 18 μL of nuclease-free water and the mixture was heated at 85°C for 5 min and then cooled to 4°C. 75 μL of reaction mix [20 μL 5x buffer, 2 μL of dNTP mixture (10 mM each), 5 μL of dithiothreitol (0.1 mM), 0.5 μL of RNasin™ (40 U/μL, Boehringer Mannheim), 2 μL of polymerase mixture, and 45.5 μL of nuclease-free water] was added and RT-PCR performed as follows: 1 cycle (50°C for 30 min, 94°C for 3 min, 50°C for 30 s, 68°C for 2.5 min), 9 cycles (94°C for 30 s, 50°C for 30 s, 68°C for 2.5 min), 20 cycles (94°C for 30 s, 50°C for 30 s, 68°C for 2.5 min in the first cycle, with an increment of 5 s per cycle thereafter), and a final extension at 68°C for 15 min. The RT-PCR product was purified by a QIAquick™ PCR Purification Kit (Qiagen) and eluted with 50 μL of 1 mM Tris-HCl, pH 7.5.

All vector constructions and analyses were carried out by using standard techniques (Sambrook et al., 1989). RT-PCR amplified cDNA, digested with KpnI and NotI nucleases, was inserted into the KpnI-NotI site of eukaryotic expression plasmid

vector (pCDNA3, Invitrogen, Carlsbad, CA). Electroporation-competent *Escherichia coli* XL1-Blue cells (Stratagene, La Jolla, CA) were transformed by electroporation (Gene Pulser™, Bio-Rad, Hercules, CA) and plated onto LB agar plates containing 100 μg/mL carbenicillin (Sigma Chemical Co., St. Louis, MO). Clones were picked and inoculated into 3 mL LB broth containing 100 μg/mL carbenicillin. Plasmid DNA was extracted from a 14 h culture using a QIAprep™ Spin Miniprep Kit (Qiagen). Automated DNA sequencing was performed as recommended (Applied Biosystems/Perkin Elmer, Foster City, CA). Both strands of the cDNA were sequenced and shown to be identical to the sequence for the original SA14 strain (Nitayaphan et al., 1990).

The fragment of plasmid pCDNA3 (Invitrogen, Carlsbad, CA) from nucleotide (nt) 1289 to nt 3455, containing f1 ori, SV40 ori, the neomycin resistance gene, and SV40 poly(A) elements was deleted by PvuII digestion and then ligated to generate the pCBamp plasmid. The vector pCIBamp, containing a chimeric intron insertion at the NcoI/KpnI site of the pCBamp was constructed by excising the intron sequence from pCI (Promega, Madison, WI) by digestion with NcoI and KpnI. The resulting 566-bp fragment was cloned into pCBamp by digesting with NcoI-KpnI to replace its 289-bp fragment. Figure 3 presents the relationships between the plasmids pCDA3, pCBamp, and pCIBamp.

Plasmids containing the transcriptional unit encoding JEV prM and E proteins were prepared from these plasmids. The cDNA fragment containing the JEV prM and E coding regions in the recombinant plasmid pCDJE2-7 (nucleotide sequence, SEQ ID NO:10; amino acid sequence, SEQ ID NO:11), derived from the pCDNA3 vector, was excised by digestion with NotI and KpnI or XbaI and cloned into the KpnI-NotI site of pCBamp, pCIBamp, pCEP4 (Invitrogen, Carlsbad, CA), or pREP4 (Invitrogen, Carlsbad, CA), or into the SpeI-NotI site of pRc/RSV (Invitrogen, Carlsbad, CA) expression vector to create pCBJE1-14 (nucleotide sequence, SEQ ID NO:17; amino acid sequence, SEQ ID NO:18), pCIBJES14, pCEJE, pREFE, and pRCJE, respectively.

Both strands of the cDNA from clones of each plasmid were sequenced and recombinant clones with the correct nucleotide sequence were identified. Plasmid DNA for use in the in vitro transformation of mammalian cells or mouse immunization experiments was purified by anion exchange chromatography using an EndoFree™ Plasmid Maxi Kit (Qiagen).

Example 2. Evaluation of JEV prM and E proteins expressed by various recombinant plasmids using an indirect immunofluorescent antibody assay. The expression of JEV specific gene products by the various recombinant expression plasmids was evaluated in transiently transfected cell lines of COS-1, COS-7 and SV-T2 (ATCC, Rockville MD; 1650-CRL, 1651-CRL, and 163.1-CCL, respectively) by indirect immunofluorescent antibody assay (IFA). The SV-T2 cell line was excluded from further testing since a preliminary result showed only 1-2% of transformed SV-T2 cells were JEV antigen positive. For transformation, cells were grown to 75% confluence in 150 cm^2 culture flasks, trypsinized, and resuspended at 4°C in phosphate buffered saline (PBS) to a final cell count 5 x 10^6 per mL. 10 μg of plasmid DNA was electroporated into 300 μL of cell suspension using a BioRad Gene Pulse™ (Bio-Rad) set at 150 V, 960 μF and 100 Ω resistance. Five minutes after electroporation, cells were diluted with 25 mL fresh medium and sceded into a 75 cm^2 flask. 48 h after transformation the medium was removed from the cells, and the cells were trypsinized and resuspended in 5 mL PBS with 3% normal goat serum. 10 μL aliquots were spotted onto slides, air dried and fixed with acetone at -20°C for 20 min. IFA was performed with acetone-fixed plasmid-transformed cells using fluorescein isothiocyanate-conjugated goat anti-mouse immunoglobulin G (Sigma Chemical Co.) and JEV HIAF.

To determine the influence of various promoter and poly(A) elements on the JEV prM and E protein expression, COS-1 and COS-7 cell lines were transiently transformed by an equal amount of pCDJE2-7 (SEQ ID NO:10), pCEJE, pREJE, or pRCJE plasmid DNA. JEV antigens were expressed in both cell lines transformed by

all four recombinant plasmids, thus confirming that the CMV or RSV (rous sarcoma virus) promoter and BGH or SV40 poly(A) elements were functionally active. However, the percentage of transformed cells and the level of JEV antigens expressed, as determined by the number of IFA positive cells and IFA intensity, respectively, differed greatly among the various plasmids (Table 1). A significantly high percentage of COS-1 cells transformed by pCDJE2-7 (SEQ ID NO:10), pCBJE1-14 (SEQ ID NO:17) and pCIBJES14 expressed the JEV antigens, and the level of the expressed proteins was compatible with JEV-infected cells. Cells transfected with pCEJE, pREJE, or pRCJE vectors, on the other hand, had a low percentage of antigen-expressing cells, as well as a low intensity of fluorescence, indicating weak expression of the antigens.

In order to ascertain whether the enhanced expression of JEV proteins by pCDJE2-7 (SEQ ID NO:10) was influenced by the SV40-encoded eukaryotic origin of replication, the plasmid pCBJE1-14 (SEQ ID NO:17) was constructed so that a 2166-bp fragment, containing f1 ori, SV40 ori, the neomycin resistance gene and SV40 poly(a) elements from pCDJE2-7, was deleted. A chimeric intron was then inserted into pCBJE1-14 to generate pCIBJES14. The pCIBJES14 plasmid was used to determine if the expression of JEV proteins could be enhanced by the intron sequence. Following transformation, cells harboring both pCBJE1-14 and pCIBJES14 vectors expressed a level of JEV antigens similar to that observed with pCDJE2-7 (Table 1). This result indicates that expression of JEV prM and E antigens by recombinant vectors is influenced only by the transcriptional regulatory elements. Neither the eukaryotic origin of replication nor the intron sequence enhanced JEV antigen expression in the cells used. Vectors containing the CMV promoter and BGH poly(A) (Figure 3) were selected for further analysis.

Example 3. Selection of an in vitro transformed, stable cell line constitutively expressing JEV specific gene products. COS-1 cells were transformed with 10 μg of pCDJE2-7 DNA by electroporation as described in the previous example. After a 24 hr

incubation in non-selective culture medium, cells were treated with neomycin (0.5 mg/mL, Sigma Chemical Co.). Neomycin-resistant colonies, which became visible after 2-3 weeks, were cloned by limited dilution in neomycin-containing medium. Expression of vector-encoded JEV gene products was initially screened by IFA using JEV HIAF. One JEV-IFA positive clone (JE-4B) and one negative clone (JE-5A) were selected for further analysis and maintained in medium containing 200 µg/mL neomycin.

Authenticity of the JEV E protein expressed by the JE-4B clone was demonstrated by epitope mapping by IFA using a panel of JEV E-specific murine monoclonal antibodies (Mab) (Kimura-Kuroda et al., *J. Virol.* 45: 124-132 (1983); Kimura-Kuroda et al., *J. Gen. Virol.* 67: 2663-2672 (1986); Zhang et al., *J. Med. Virol.* 29: 133-138 (1989); and Roehrig et al., *Virol.* 128: 118-126 (1983)). JEV HIAF and normal mouse serum were used as positive and negative antibody controls, respectively. Four JEV-specific, six flavivirus-subgroup specific, and two flavivirus-group reactive Mabs reacted similarly with the 4B clone or JEV-infected COS-1 cells (Table 2).

Example 4. Antigenic properties and immunological detection of subviral particles secreted by the JE-4B COS-1 cell line.

a. Preparation of subviral particles. JE-4B COS-1 cells were grown and maintained in medium containing 200 µg/mL of neomycin. The cultured medium was routinely harvested and stored at 4°C, and replenished twice weekly, and the cells were split 1:5 every 7-10 days. Culture medium was clarified by centrifugation at 10,000 rpm for 30 min in a Sorvall F16/250 rotor at 4°C, and centrifuged further for 4 hr at 39,000 rpm in a Sorvall TH641 rotor at 4°C through a 5% sucrose cushion (w/w, prepared with 10 mM Tris HCl, pH 7.5, 100 mM NaCl (TN buffer)). The pellet containing subviral particles was resuspended in TN buffer and stored at 4°C. Alternatively, 7% or 10% PEG-8000 (w/v) was added to the clarified culture medium. The mixture was stirred at 4°C for at least 2 hr, and the precipitated particles were

collected by centrifugation at 10,000 rpm for 30 min. The precipitate was resuspended in TN buffer and stored at 4°C. The subviral particles were purified from both pelleted and PEG-precipitated preparations by rate zonal centrifugation in a 5-25% continuous sucrose gradient in TN at 38,000 rpm at 4°C for 90 min. 1-mL fractions were collected from the top of the gradient, tested by antigen capture ELISA (see below), and the positive fractions loaded onto a 25-50% sucrose gradient in TN. This was centrifuged overnight in an equilibrium density centrifugation at 35,000 rpm at 4°C. 0.9-mL fractions from the equilibrium gradients were collected from the bottom. They were tested by antigen-capture ELISA and assessed for hemagglutination (HA) activity at pH 6.6. An aliquot of 100 µL of each fraction was weighed precisely to determine its density. The ELISA-positive fractions were pooled and pelleted at 39,000 rpm at 4°C for 3-4 hr and the pellet resuspended in TN buffer. Antigen-capture ELISA and HA titers were determined on the pelleted samples. JEV-infected COS-1 cell supernatant was also subjected to similar purification protocols as detailed above and used as a positive control for the gradient analysis. JE virions were also purified from infected C6/36 cells 5-6 days postinfection by sedimentation in a glycerol/tartrate equilibrium gradient.

b. Western blots of subviral particles. Gradient-purified samples of the subviral particles were mixed with electrophoresis sample buffer and run on 10 or 12.5% sodium dodecyl sulfate-containing polyacrylamide gels (SDS-PAGE) as described by Laemmli (*Nature* 277: 680-685 (1970)). Proteins were transferred to a nitrocellulose membrane and immunochemically detected with polyclonal JEV HIAF, flavivirus cross-reactive anti-E Mab 4G2 (Henchal et al., *Amer. J. Trop. Med. Hyg.* 31: 830-836 (1982)), or mouse anti-prM peptide hyperimmune serum (JM01). Figure 4 shows a comparison of the M and E proteins produced by JEV infected C6/36 and JE-4B COS-1 cells. Some nonspecific reactivity to E protein was observed in the normal mouse ascitic fluid and Jmo1 anti-peptide serum. Proteins identical in size to M and E were secreted in the subviral particles and could be detected by E-specific Mab 4G2 and prM-specific JM01 antiserum, respectively.

c. Density gradient detection of JEV subviral particles in culture medium. For ELISA, antigen-capture antibody (4G2) was diluted in 0.1 M sodium carbonate buffer, pH 9.6, and used to coat 96-well microtiter plates (Immulon II, Dynatech. Chantilly, VA) by overnight incubation at 4° C. After blocking with 3% normal goat serum in PBS, two-fold serially-diluted samples were added to the 4G2-coated plate and incubated 1.5 hours at 37°C. Captured antigen was detected by horseradish peroxidase-conjugated 6B6C-1 Mag, and incubated for 1 hour at 37°C. The enzyme activity on the solid phase was then detected with TMB (3,3',5,5'-tetramethylbenzidine)-ELISA (Life Technologies, Grand Island, NY).

Approximately 500 mL of cell culture medium from 15 X 150 cm^2 flasks of JE-4B cells was collected four days after cells were seeded. PEG-precipitated subviral particles were resuspended in 2 mL of TN buffer, pH 7.5; a 0.7 mL aliquot of this resuspended pellet was loaded onto a 5-25% sucrose gradient. Triton X-100, which disrupts subviral particles, was added to another 0.7 mL aliquot to a final concentration of 0.1% and this was loaded onto a 5-25% sucrose gradient prepared in TN buffer containing 0.1% Triton X-100. A definite opaque band was observed approximately 2.5 cm from the top of the gradient containing Triton X-100, but not in the gradient without detergent. Fractions (1 mL) were collected from top to bottom for each gradient (Figure 5). Each collected fraction was analyzed by antigen capture ELISA. Antigen was detected in fractions 4-6, indicating relatively rapid sedimentation characteristic of subviral particles. Treatment of the PEG precipitate from JE-4B culture medium with Triton X-100 shifted the position of ELISA-reactive material to the top of the gradient. Thus treatment with Triton X-100 produces only slow-sedimenting molecules. A similar finding was reported by Konishi et al. (*Virol.* 188: 714-720 (1992)). These results show that rapidly sedimenting subviral particles containing prM/M and E could be disrupted by detergent treatment.

Hemagglutination (HA) activity was determined in the pH range from 6.1 to 7.0 by the method of Clarke and Casals (*Amer. J. Trop. Med. Hyg.* 7: 561-573 (1958)).

The subviral particle secreted by JE-4B cells and the virion particle produced by JEV infected COS-1 cells had a similar HA profile with the optimum pH determined to be 6.6.

Example 5. Comparison of the immune response in mice vaccinated with pCDJE2-7 nucleic acid vaccine of the invention and commercial JEV vaccine. Groups of five 3-week-old female, ICR outbred mice were injected intramuscularly in the left and right quadriceps with 100 μg of pCDJE2-7 plasmid in 100 μL of dH_2O or were given doses of JE-VAX (manufactured by the Research Foundation for Microbial Disease of Osaka University and distributed by Connaught Laboratories, Swiftwater, PA.) subcutaneously that are one-fifth the dose given to humans. The plasmid pCDNA3/CAT (Invitrogen), which encodes and expresses an unrelated protein, was used as the negative vaccination control. Except for one group of pCDJE2-7-vaccinated mice, all animals were boosted 3 weeks later with an additional dose of plasmid or JE-VAX. Mice were bled from the retroorbital sinus at 3, 6, 9, 23, 40 and 60 weeks after inoculation. JEV antibody titers were determined by enzyme-linked imunosorbent assay (ELISA) against purified JEV or by plaque reduction neutralization tests (PRNT) (Roehrig et al., *Virol.* 171: 49-60 (1989); and Hunt and Calisher, *Amer. J. Trop. Med. Hyg.* 28: 740-749 (1979)).

The pCDJE2-7 nucleic acid vaccine and JE-VAX provided 100% seroconversion within three weeks after the first vaccination in all three groups of mice (Table 3). The JEV ELISA and PRNT antibody titers reached the highest level at week 6 and week 9, respectively, after immunization. Mice receiving 1 dose of DNA vaccine had similar antibody responses as those receiving 2 doses. Comparable ELISA antibody titers were maintained in DNA-vaccinated groups up to 60 weeks, after which the experiment was terminated. However, only one of four mice in the JE-VAX group was JEV antibody positive at 60 weeks post-inoculation. The pCDNA3/CAT control group did not have any measurable JEV antibody. These results demonstrate that a

single dose of JEV-specific nucleic acid vaccine is more effective in maintaining JEV antibody in mice than the commercial, FDA-approved JE-VAX vaccine.

Example 6. Comparison of various nucleic acid vaccine constructs of the invention and commercial JEV vaccine for effectiveness of vaccination at different ages. A similar level of JEV protein was expressed by COS-1 cells transformed by either pCDJE2-7, pCBJE1-14, or pCIBJES14. JEV antibody induction by these nucleic acid constructs was compared to JE-VAX commercial vaccine at two different ages at vaccination. Three-day (mixed sex) or 3-week-old (female) ICR outbred mice, 10 per group, were vaccinated intramuscularly with 50 or 100 μg of plasmid DNA, or subcutaneously with doses of JE-VAX that are one-tenth or one-fifth the dose given to humans. Serum specimens were collected at 3 and 7 weeks after immunization and tested at a 1:1600 dilution by ELISA using purified JEV as an antigen. Results are shown in Table 4.

Plasmid pCBJE1-14 provided the highest extent of seroconversion, i.e., antibody titer greater than 1:1600, achieving 80-100% at both ages of vaccination. Administration of pCDJE2-7 or pCIBJES14 provided moderate seroconversion by 7 weeks when 3-day old mice were vaccinated (60% for each), but weaker seroconversion (40% and 10%, respectively) when measured 3 weeks after vaccination. When these plasmids were administered at the age of 3 weeks, however, seroconversions of 90% or 100% were attained at both 3 weeks and 7 weeks after vaccination. In contrast, the commercial vaccine, JE-VAX, conferred no seroconversion when administered at 3 days of age, and 100% when given at 3 weeks of age. Thus the nucleic acid TU's for JEV prM and E provided an extent of seroconversion better than a very high dose of the commercial vaccine, and unexpectedly high seroconversion in both young and more mature animals.

Example 7. Protective immunity conferred by the nucleic acid vaccine of the invention. Three-day old vaccinated groups from Example 6 were challenged 7 weeks

after vaccination by intraperitoneal injection of 50,000 pfu/100 μL of the mouse-adapted JEV strain SA14 and observed for 3 weeks. 100% protection was achieved in groups that received various nucleic acid TU-containing vaccine constructs for up to 21 days (Table 5). In contrast, 60% of the JE-VAX-vaccinated mice, as well as 70% of the pCDNA3/CAT-vaccinated negative controls, did not survive virus challenge by 21 days. These results indicate that the nucleic acid TU's of the invention confer unexpectedly effective protection on vaccinated mice. This suggests the possibility of employing the nucleic acid vaccine of the invention as an early childhood vaccine for humans. In contrast, JE-VAX, the inactivated human vaccine currently used, does not appear to be effective in young animals.

Example 8. Passive protection of neonatal mice correlated with the maternal antibody titer. Female ICR mice at the age of 3 weeks were vaccinated with either one dose or two doses spaced two days apart of pCDJE2-7 plasmid DNA, at 100 μg/100 μL, or with two doses of JE-VAX that were one-fifth the dose given to humans. The negative control group received two doses of 100 μg/100 μL of pCDNA-3/CAT plasmid. Passive protection by maternal antibody was evaluated in pups resulting from matings of experimental females with non-immunized male mice that occurred nine weeks following the first vaccination or 6 weeks following the second vaccination. Pups were challenged between 3-15 days after birth by intraperitoneal administration of 5,000 pfu/100 μL of mouse-adapted SA14 virus and observed daily for 3 weeks (Table 6). The survival rates correlated with the maternal neutralizing antibody titers. 100% of pups nursed by mothers with a PRNT of 1:80 survived viral infection, whereas none of the pups from the control mother survived (Table 6). Partial protection of 45% and 75% was observed in older pups that were nursed by mothers with a PRNT titer of 1:20 and 1:40, respectively. The survival rates also correlated with the length of time that pups were nursed by the immune mother. As just indicated, 13-15 day old pups had high survival rates. None of the 3-4 day old pups, however, survived virus challenge when the mother had a PRNT titer of 1:20 or 1:40. Thus maternal antibody provides

partial to complete protective immunity to the offspring. In addition, JEV antibody was detected by ELISA in the sera of 97% (29/30) of the post-challenge pups.

Mice were inoculated intramuscularly with 1 or 2, 100 µg doses of plasmid DNA, or subcutaneously with two, 1/5 human doses of JE-VAX vaccine. Sera were collected 9 weeks post-vaccination for PRNT testing prior to mating with non-immune male.

Example 9. Preparation of recombinant plasmids containing the transcriptional unit encoding WNV prM and E antigens. Genomic RNA was extracted from 150 µL of Vero cell culture medium infected with NY 99-6480 strain, an strain isolated from the outbreak in New York 1999, using the QIAamp ™ Viral RNA Kit (Qiagen, Santa Clarita, CA). Extracted RNA was eluted and suspended in 80 µl of nuclease-free water, and used as a template for the amplification of WNV prM and E gene coding sequences. Primer sequences were obtained from the work of Lanciotti et al. (*Science* 286: 2333-2337 (1999)). A cDNA fragment containing the genomic nucleotide region was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction sites BsmBI and KasI were engineered at the 5' terminus of the cDNA by using amplimer WN466 (nucleotide sequence, SEQ ID NO:12). An in-frame translation termination codon , followed by a NotI restriction site was introduced at the 3' terminus of the cDNA by using amplimer cWN2444 (SEQ ID NO:13). The RT-PCR product was purified by a QIAquick™ PCR Purification Kit (Qiagen).

The double-stranded amplicon produced by use of the two amplimers above (SEQ ID NO:12 and SEQ ID NO:13) was digested with KasI and NotI enzymes to generate a 998 bp (nt-1470 to 2468) fragment of DNA was inserted into the KasI and NotI sites of a pCBJESS vector to form an intermediate plasmid, pCBINT. The pCBJESS was derived from the pCBamp plasmid, that contained the cytomegalovirus early gene promoter and translational control element and an engineered JE signal

sequence element (Chang et al., *J. Virol.* 74: 4244-4252 (2000)). The JE signal sequence element comprises the JE signal sequence (SEQ ID NO:14).

The cDNA amplicon was subsequently digested with BsmBI and Kas I enzymes and the remaining 1003 bp fragment (nt-466 to 1470) was inserted in to the KasI site of pCBINT to form pCBWN (nucleic acid sequence, SEQ ID NO:15; amino acid sequence, SEQ ID NO:16). Automated DNA sequencing using an ABI prism 377 Sequencer (Applied Biosystems/Perkin Elmer, Foster City, CA) was used to confirm that the recombinant plasmid had a correct prM and E sequence as defined by Lanciotti et al. (*Science* 286: 2333-2337 (1999)).

Plasmid DNA for use in the in vitro transformation of mammalian cells or mouse immunization experiments was purified by anion exchange chromatography as described in Example 1.

Example 10. Immunochemical characterization and evaluation of WNV prM and E proteins expressed by pCBWN. WNV specific gene products encoded by the pCBWN plasmid were expressed in COS-1 cells. Cells were electroporated and transformed with pCBWN plasmid according to Chang et al. (*J. Virol.* 74: 4244-4252 (2000)). Electroporated cells were seeded onto 75 cm^2 culture flasks or a 12-well tissue culture dish containing one sterile coverslip/well. All flasks and 12-well plates were kept at 37°C, 5% CO_2 incubator. Forty hours following electroporation, coverslips containing adherent cells were removed from the wells, washed briefly with PBS, fixed with acetone for 2 minutes at room temperature, and allowed to air dry.

Protein expression was detected using indirect immunofluorescence antibody assay (IFA), as described in Example 2. Flavivirus E-protein specific monoclonal antibody (Mab) 4G2, WNV mouse hyperimmune ascitic fluid (HIAF) and normal mouse serum (NMS) at 1:200 dilution in PBS were used as the primary antibody to detect protein expression (Henchal et al., *Am. J. Trop. Med. Hyg.* 31: 830-836 (1982)).

Tissue culture medium was harvested 40 and 80 hours following electroporation. Antigen-capture (Ag-capture) ELISA was used to detect secreted WN virus antigen in the culture medium of transiently transformed COS-1 cells. The Mab 4G2 and horseradish peroxidase-conjugated Mab 6B6C-1 were used to capture the WN virus antigens and detect captured antigen, respectively (Chang et al., *J. Virol.* 74: 4244-4452 (2000); Henchal et al., *Am. J. Trop. Med. Hyg.* 31: 830-836 (1983); Roehrig et al., *Virology* 128: 118-126 (1983)).

WN virus antigen in the medium was concentrated by precipitation with 10 % polyethylene glycol (PEG)-8000. The precipitant was resuspended in TNE buffer (50 mM Tris, 100 mM NaCl, 10 mM EDTA, pH 7.5), clarified by centrifugation, and stored at 4°C. Alternatively, the precipitant was resuspended in a lyophilization buffer (0.1 M TRIZMA and 0.4% bovine serum albumin in borate saline buffer, pH 9.0), lyophilized and stored at 4°C. Lyophilized preparations were used as antigen for the evaluation in MAC- and indirect IgG ELISAs.

WN virus-specific protein was detected by IFA on the transiently transformed COS-1 cells. E, prM and M proteins expressed in these cells were secreted into the culture medium. WN virus antigen concentrated by PEG precipitation was extracted with 7.0 % ethanol to remove residual PEG (Aizawa et al., *Appl. Enviro. Micro.* 39: 54-57 (1980)). Ethanol extracted antigens and gradient-purified WN virions were analyzed on a NuPAGE, 4-12% gradient Bis-Tris Gel in a Excel Plus Electrophoresis Apparatus (Invitrogen Corp., Carlsbad,CA) and followed by electroblotting onto nitrocellulose membranes using a Excel Plus Blot Unit (Invitrogen Corp.). WN virus-specific proteins produced by the transiently transformed COS-1 cells were detected by WN virus specific mouse HIAF or flavivirus E protein reactive Mab 4G2 in a Western blot analysis, using NMS as a negative serum control. The proteins displayed similar reactivity and identical molecular weights to the corresponding gradient purified virion E, prM and M protein derived from WN virus infected suckling mouse brain (SMB).

In analysis of the NRA as an antigen for diagnostic ELISA, one vial of lyophilized NRA, representing antigen harvested from 40 ml of tissue culture fluid, was reconstituted in 1.0 ml of distilled water and compared with the reconstituted WN virus infected suckling mouse brain (SMB) antigen provided as lyophilized as β-propiolactone-inactivated sucrose-acetone extracts (Clarke et al., *Am. J. Trop. Med. Hyg.* 7: 561-573 (1958)). All recombinant proteins, prM, M and E , had a similar reactivity to that of the gradient-purified virion E, prM and M proteins.

Coded human specimens were tested concurrently with antigens in the same test at the developmental stage. The MAC- and IgG ELISA protocols employed were identical to the published methods (Johnson et al., *J. Clin. Microbiol.* 38: 1827-1831 (2000); Martin et al., *J. Clin. Microbiol.* 38: 1823-1826. (2000)). Human serum specimens were obtained from the serum bank in our facility, which consists of specimens sent to the DVBID for WN virus confirmation testing during the 1999 outbreak In these tests, a screening MAC- and IgG ELISA were performed on a 1:400 specimen dilution. Specimens yielding positive/negative (P/N) OD ratios between 2 and 3 were considered suspect positives. Suspect serum specimens were subject to confirmation as positives by both ELISA end-point titration and plaque-reduction neutralization test (PRNT). All specimens yielding P/N OD ratios greater than 3.0 were considered positives without further confirmatory testing.

An Ag-capture ELISA employing flavivirus-group reactive, anti-E Mab, 4G2 and 6B6C-1, was used to detect NRA secreted into culture fluid of pCBWN transformed COS-1 cells. The antigen could be detected in the medium one day following transformation; and the maximum ELISA titer (1:32-1:64) in the culture fluid without further concentration was observed between day two and day four. NRA was concentrated by PEG precipitation, resuspended in a lyophilization buffer, and lyophilized for preservation. For diagnostic test development, one vial of lyophilized NRA was reconstituted with 1.0 ml distilled water and titrated in the MAC- or indirect IgG ELISA using WN virus positive and negative reference human sera (Johnson et al.,

J. Clin. Microbiol. 38: 1827-1831 (2000); Martin et al., *J. Clin. Microbiol.* 38: 1823-1826 (2000)). Dilutions 1:320 and 1:160 of the NRA were found to be the optimal concentrations for use in MAC- and IgG ELISA, respectively. These dilutions resulted in a P/N OD_{450} ratio of 4.19 and 4.54, respectively, for MAC- and IgG test. The WN virus SMB antigens produced by NY-6480 and Eg101 strains were used at 1:320 and 1:640 dilution for MAC-ELISA, and 1:120 and 1:320 for IgG ELISA, respectively. The negative control antigens, PEG precipitates of the culture medium of normal COS-1 cells and normal SMB antigen, were used at the same dilutions as for the respective NRA and SMB antigen. Human serum specimens, diluted at 1:400, were tested concurrently in triplicate with virus-specific and negative control antigens. For the positive test result to be valid, the OD_{450} for the test serum reacted with viral antigen (P) had to be at least two-fold greater than the corresponding optical density value of the same serum reacted with negative control antigen (N).

The reactivity of NRA and NY-06480, Eg101 and SLE virus SMBs were compared by the MAC- and IgG ELISAs using 21 coded human serum specimens. Of the 21 specimens, 19 had similar results on all three antigens (8 negatives and 11 suspect positives or positives). Eighteen specimens were also tested separately using SLE SMB antigen. Only three of 13 Eg-101-SMB positive specimens were positive in the SLE MAC-ELISA (Table 1). None of WN antigen negative specimens was positive by SLE MAC-ELISA. This result confirmed a previous observation that anti-WN virus IgM did not cross-react significantly with other flaviviruses (Tardei et al., *J. Clin. Microbiol.* 38: 2232-2239 (1940)) and was specific to diagnose acute WN virus infection regardless of whether NRA or SMB antigen was used in the test. All of the specimens were also tested concurrently by indirect IgG ELISA. Ten of 21 specimens were positive using any of the three antigens.

The two discrepant serum specimens (7 and 9) both from the same patient, collected on day-4 and 44 after onset of disease, respectively, were IgM-negative with NRA and SMB NY antigen and IgM-positive using Eg-101 SMB antigen in the initial

test. To investigate these two discordant specimens further, six sequentially collected specimens from this patient were retested by end-point MAC- and IgG ELISAs. A greater than 32-fold serial increase shown in the MAC-ELISA titer between day-3 and day-15 could be demonstrated with all antigens used. Cerebrospinal fluid collected on day-9 after onset of disease also confirmed that this patient indeed was infected by WN shortly prior to taking the sample. The cerebrospinal fluid had IgM P/N reading of 13.71 and 2.04 against Eg-101- and SLE-SMB antigens, respectively. Day-31 and day-44 specimens were negative (<1:400) by using NY-SMB antigen but positive by using NRA and Eg101-SMB. Compatible IgG titers were observed with all three antigen used in the test.

Example 11. Evaluation of the immune response in animals vaccinated with pCBWN. Groups of ten, three-wk-old female ICR mice were used in the study. Mice were injected intramuscularly (i.m.) with a single dose of pCBWN or a green fluorescent protein expressing plasmid (pEGFP) DNA (Clonetech, San Francisco, CA.). The pCBWN plasmid DNA was purified from XL-1 blue cells with EndoFree Plasmid Giga Kits (Qiagen) and resuspended in PBS, pH 7.5, at a concentration of 1.0 μg/μl. Mice that received 100 μg of pEGFP were used as unvaccinated controls. Mice were injected with the pCBWN plasmid at a dose of 100, 10, 1.0, or 0.1 μg in a volume of 100 μl. Groups that received 10, 1.0, or 0.1 μg of pCBWN were vaccinated by the electrotransfer mediated *in vivo* gene delivery protocol using the EMC-830 square wave electroporator (Genetronics Inc. San Diego, CA.). The electrotransfer protocol was based on the method of Mir et al., (*Proc. Natl. Acad. Sci. USA* 96: 4262-4267.(1999)). Immediately following DNA injection, transcutaneous electric pulses were applied by two stainless steel plate electrodes, placed 4.5-5.5 mm apart, at each side of the leg. Electrical contact with the leg skin was ensured by completely wetting the leg with PBS. Two sets of four pulses of 40 volts/mm of 25 msec duration with a 200 msec interval between pulses were applied. The polarity of the electrode was reversed between the set of pulses to enhance electrotransfer efficiency.

Mice were bled every 3 wks following injection. The WN virus specific antibody response was evaluated by Ag-capture ELISA and plaque reduction neutralization test (PRNT). Individual sera were tested by IgG-ELISA, and pooled sera from 10 mice of each group were assayed by PRNT. All the mice vaccinated with pCBWN had IgG ELISA titers ranging from 1:640 to 1:1280 three wks after vaccination. The pooled sera collected at three and six wks had a Nt antibody titer of 1:80. None of the serum specimens from pEGFP control mice displayed any ELISA or Nt titer to WN virus.

To determine if the single i.m. vaccination of pCBWN could protect mice from WN virus infection, mice were challenged with NY-6480 virus either by intraperitoneal injection or by exposure to the bite of virus-infected *Culex* mosquitoes. Half of the mouse groups were challenged intraperitoneally (ip) at 6 wks post vaccination with 1,000 LD_{50} (1,025 PFU/100 µl) of NY99-6480 virus. The remaining mice were each exposed to the bites of three Culex tritaeniorhynchus mosquitoes that has been infected with NY99-6480 virus 7 days prior to the challenge experiment. Mosquitoes were allowed to feed on mice until they were fully engorged. Mice were observed twice daily for three wks after challenge.

It was evident that the presence of Nt antibodies correlated with protective immunity, since all mice immunized with WN virus DNA remained healthy after virus challenge while all control mice developed symptoms of CNS infection 4-6 days following virus challenge and died on an average of 6.9 and 7.4 days after intraperitoneal or infective mosquito challenge, respectively. In the vaccinated group, the pooled sera collected three wks after virus challenge (9-wk post immunization) had Nt antibody titers of 1:640 or 1:320. Pooled vaccinated mouse sera reacted only with E protein in the Western blot analysis.

Groups of ten mice were immunized with 10.0 to 0.1 µg of pCBWN per animal by use of electrotransfer. All groups that received pCBWN were completely protected

from virus challenge. At 6 wks after immunization all groups of electrotransfer mice had Nt titer less than four-fold different than animals receiving 100 μg of pCBWN by conventional i.m. injection without electrotransfer. Both these results evidencing effective immunization suggest that the electrotransfer protocol enhances the immunogenicity and protective efficacy of the DNA vaccine of the invention (when carried out as described in (Mir et al., *Proc. Natl. Acad. Sci. USA.* 96: 4262-4267.(1999)).

Mixed-bred mares and geldings of various ages used in this study were shown to be WN virus and SLE virus antibody-negative by ELISA and PRNT. Four horses were injected i.m. with a single dose (1,000 μg/1,000 μl in PBS, pH 7.5) of pCBWN plasmid. Serum specimens were collected every other day for 38 days prior to virus challenge, and the WN virus specific antibody response was evaluated by MAC- or IgG ELISA and PRNT.

Two days prior to virus challenge, 12 horses (4 vaccinated and 8 control) were relocated into a bio-safety level (BSL)-3 containment building at the Colorado State University. The eight unvaccinated control horses were the subset of a study that was designed to investigate WN virus induced pathogenesis in horses and the potential of horses to serve as amplifying hosts. Horses were each challenged by the bite of 14 or 15 *Aedes albopictus* mosquitoes that had been infected by NY99-6425 or BC787 virus 12 days prior to horse challenge. Mosquitoes were allowed to feed on horses for a period of 10 min. Horses were examined for signs of disease twice daily. Body temperature was recorded, and serum specimens collected twice daily from days 0 (day of infection) to 10, then once daily through day 14. Pulse and respiration were recorded daily after challenge. The collected serum samples were tested by plaque titration for detection of viremia, and by MAC- or IgG ELISA and PRNT for antibody response.

No systemic or local reaction was observed in any vaccinated horse. Individual horse sera were tested by PRNT. Vaccinated horses developed Nt antibody greater than

or equal to 1:5 between days 14 and 31. End point titers for vaccinated horses, #5, #6, #7, and #8, on day-37 (two days prior to mosquito challenge) were 1:40, 1:5, 1:20, and 1:20, respectively. Horses vaccinated with the pCBWN plasmid remained healthy after virus challenge. None of them developed a detectible viremia or fever from days 1 to 14. All unvaccinated control horses became infected with WN virus after exposure to infected mosquito bites. Seven of the eight unvaccinated horses developed viremia that appeared during the first 6 days after virus challenge. Viremic horses developed Nt antibody between day-7 and day-9 after virus challenge. The only horse from the entire study to display clinical signs of disease was horse #11, which became febrile and showed neurologic signs beginning 8 days after infection. This horse progressed to severe clinical disease within 24 hours and was euthanized on day 9. Four representing horses, #9, 10, 14 and 15, presenting viremia for 0, 2, 4, or 6 days, were selected and used as examples in this example. Virus titers ranged from 10^{10} PFU/ml of serum (in horse #10), the lowest level detectable in our assay, to 10^{24}/ml (in horse #9). Horse #14 did not develop a detectible viremia during the test period. However, this horse was infected by the virus, as evidenced by Nt antibody detected after day 12.

Anamnestic Nt antibody response was not observed in vaccinated horses as evidenced by the gradual increase in Nt titer during the experiment. Pre-existing Nt antibody in the vaccinated horse prior to mosquito challenge could suppress initial virus infection and replication. Without virus replication, the challenge virus antigen provided by infected mosquitoes may not contain a sufficient antigen mass to stimulate anamnestic immune response in the vaccinated horse. All vaccinated horses were euthanized at 14 days after virus challenge. Gross pathological and histopathological lesions indicative of WN viral infection were not observed.

Example 12. Preparation of recombinant plasmids containing coding sequences for yellow fever virus (YFV) or St. Louis encephalitis virus (SLEV) prM and E proteins. A strategy similar to constructing the pCDJE2-7 recombinant plasmid was used to prepare YFV and SLEV recombinant plasmids. Genomic RNA was extracted

from 150 µL of YFV strain TRI-788379 or SLE strain 78V-6507 virus seeds using Q1Aamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA.). The viral RNA was used as a template for amplification of YFV or SLEV prM and E gene coding regions. Primers YFDV389 (nucleotide sequence, SEQ ID NO:4; amino acid sequence, SEQ ID NO:5), cYFDV2452 (SEQ ID NO:6), SLEDV410 (nucleotide sequence, SEQ ID NO:7; amino acid sequence, SEQ ID NO: 8) and cSLEDV2449 (SEQ ID NO:9) were used to generate the corresponding recombinant nucleic acids as described above for the preparation of the JEV and WNV recombinant plasmids. RT-PCR amplified cDNA, digested with KpnI and NotI enzymes, was inserted into the KpnI-NotI site of a eukaryotic expression plasmid vector, pCDNA3 (Invitrogen). Both strands of the cDNA were sequenced and verified for identity to sequences from YFV strain TRI-788379 or SLEV strain 78V-6507. Recombinant plasmids pCDYF2 and pCDSLE4-3, which contained the nucleotide sequences of the prM and E coding regions for YFV or SLEV, respectively, were purified using an EndoFree™ Plasmid Maxi Kit (Qiagen), and used for *in vitro* transformation or mouse immunization.

YFV or SLEV specific antigens were expressed in COS-1 cells transformed by pCDYF2 or pCDSLE4-3, respectively. The level of expressed proteins was similar to a YFV- or SLEV-infected COS-1 cell control. As in the JEV model, COS-1 cell lines transformed by vectors bearing genes for the viral antigens were obtained which constitutively express YFV or SLEV antigenic proteins. Epitope mapping by IFA using a panel of YFV or SLEV E-specific Mabs indicated that the authentic E protein was expressed by the pCDYF2- or pCDSLE4-3-transformed COS-1 cells. A preliminary study indicated that 100% of three week-old female, ICR mice seroconverted after intramuscular inoculation with a single dose of 100 µg/100 µL of pCDSLE4-3 plasmid in deionized water.

Example 13. Preparation of recombinant plasmids containing coding sequences for St. Louis encephalitis virus prM and E antigens with JEV signal sequence.

Genomic RNA was extracted from 150 µL of Vero cell culture medium infected with

MSI-7 strain of St. Louis encephalitis virus using the QIAamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA). Extracted RNA was eluted and suspended in 80 µl of nuclease-free water, and used as a template for the amplification of St. Louis encephalitis virus prM and E gene coding sequences. Primer sequences were obtained from the work of Trent et al. (*Virology* 156: 293-304 (1987)). A cDNA fragment containing the genomic nucleotide region was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction site AfeI was engineered at the 5' terminus of the cDNA by using amplimer SLE463 (SEQ ID NO:30). An in-frame translation termination codon , followed by a NotI restriction site was introduced at the 3' terminus of the cDNA by using amplimer cSLE2447 (SEQ ID NO:31). The RT-PCR product was purified by a QIAquick™ PCR Purification Kit (Qiagen).

The double-stranded amplicon, produced by use of the two amplimers above (SEQ ID NO:30 and SEQ ID NO:31), was digested with AfeI and NotI enzymes to generate a 2004 fragment of DNA (463 to 2466nt), and inserted into the AfeI and NotI sites of a pCBJESS-M vector to form pCBSLE (nucleotide sequence, SEQ ID NO:21; amino acid sequence, SEQ ID NO:22). The pCBJESS-M was derived from the pCBamp plasmid, that contained the cytomegalovirus early gene promoter and translational control element and an engineered, modified JE signal sequence element (SEQ ID NO:27). The JE signal sequence element comprises the modified JE signal sequence at –4 (Cys to Gly) and –2 (Gly to Ser) position in the original pCBJESS plasmid.

Automated DNA sequencing using an ABI prism 377 Sequencer (Applied Biosystems/Perkin Elmer, Foster City, CA) was used to confirm that the recombinant plasmid had a correct prM and E sequence as defined by Trent et al. (*Virology* 156: 293-304 (1987)).

Example 14. Preparation of recombinant plasmids containing coding sequences for yellow fever virus (YFV) prM and E proteins with JEV signal sequence. Genomic

RNA was extracted from 150 µL of Vero cell culture medium infected with 17D-213 strain of yellow fever virus using the QIAamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA). Extracted RNA was eluted and suspended in 80 µl of nuclease-free water, and used as a template for the amplification of yellow fever virus prM and E gene coding sequences. Primer sequences were obtained from the work of dos Santos et al. (*Virus Research* 35: 35-41 (1995)). A cDNA fragment containing the genomic nucleotide region was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction site AfeI was engineered at the 5' terminus of the cDNA by using amplimer YF482 (SEQ ID NO:28). An in-frame translation termination codon, followed by a NotI restriction site was introduced at the 3' terminus of the cDNA by using amplimer cYF2433 (SEQ ID NO:29). The RT-PCR product was purified by a QIAquick™ PCR Purification Kit (Qiagen).

The double-stranded amplicon, produced by use of the two amplimers above (SEQ ID NO:28 and SEQ ID NO:29), was digested with AfeI and NotI enzymes to generate a 1971 fragment of DNA (482 to 2452nt), and inserted into the AfeI and NotI sites of a pCBJESS-M vector to form pCBYF (nucleotide sequence, SEQ ID NO:23; amino acid sequence, SEQ ID NO:24). The pCBJESS-M was derived from the pCBamp plasmid, that contained the cytomegalovirus early gene promoter and translational control element and an engineered JE signal sequence element (SEQ ID NO:27). The JE signal sequence element comprises the modified JE signal sequence at –4 (Cys to Gly) and –2 (Gly to Ser) position of JESS in the pCBJESS plasmid.

Automated DNA sequencing using an ABI prism 377 Sequencer (Applied Biosystems/Perkin Elmer, Foster City, CA) was used to confirm that the recombinant plasmid had a correct prM and E sequence as defined by dos Santos et al. (*Virus Research* 35: 35-41 (1995)).

Example 15. Preparation of recombinant plasmids containing coding sequences for Powassan virus prM and E antigens with JEV signal sequence. Genomic RNA was

extracted from 150 µL of Vero cell culture medium infected with LB strain of Powassan virus using the QIAamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA). Extracted RNA was eluted and suspended in 80 µl of nuclease-free water, and used as a template for the amplification of Powassan virus prM and E gene coding sequences. Primer sequences were obtained from the work of Mandl et al. (*Virology* 194: 173-184 (1993)). A cDNA fragment containing the genomic nucleotide region was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction site AfeI was engineered at the 5' terminus of the cDNA by using amplimer POW454 (SEQ ID NO:25). An in-frame translation termination codon, followed by a NotI restriction site was introduced at the 3' terminus of the cDNA by using amplimer cPOW2417 (SEQ ID NO:26). The RT-PCR product was purified by a QIAquick™ PCR Purification Kit (Qiagen).

The double-stranded amplicon, produced by use of the two amplimers above (SEQ ID NO:25 and SEQ ID NO:26), was digested with AfeI and NotI enzymes to generate a 1983 bp fragment of DNA (454 to 2436nt), and inserted into the AfeI and NotI sites of a pCBJESS-M vector to form pCBPOW (nucleotide sequence, SEQ ID NO:19; amino acid sequence, SEQ ID NO:20). The pCBJESS-M was derived from the pCBamp plasmid, that contained the cytomegalovirus early gene promoter and translational control element and an engineered JE signal sequence element (SEQ ID NO:27). The JE signal sequence element comprises the modified JE signal sequence at –4 (Cys to Gly) and –2 (Gly to Ser) position of JESS in the pCBJESS plasmid.

Automated DNA sequencing using an ABI prism 377 Sequencer (Applied Biosystems/Perkin Elmer, Foster City, CA) was used to confirm that the recombinant plasmid had a correct prM and E sequence as defined by Mandl et al. (*Virology* 194:173-184, (1993)).

Example 16. Preparation of plasmids containing coding sequences for dengue serotype 2 structural proteins. Procedures such as those carried out for other

flaviviruses (see Examples 1, 9 and 12-15) are to be followed to prepare vectors including nucleic acid TU's for dengue serotype 2 antigens. According to the examples, the amplimers used for construction of the vectors may be chosen to engineer the normal dengue virus signal sequence or they may be chosen so as to engineer a signal sequence from another flavivirus, such as a modified Japanese encephalitis virus signal sequence.

A plasmid containing the dengue serotype 2 gene region from prM to E is to be constructed. The dengue serotype 2 prM and E genes (Deubel et al., *Virology* 155:365-377 (1986); Gruenberg et al., *J. Gen. Virol.* 69: 1301-1398 (1988); Hahn et al., *Virology* 162:167-180 (1988)) are to be ligated into a plasmid such as pCDNA3, and then excised and cloned into vectors such as pCBamp, pCEP4, pREP4, or pRc/RSV (supplied by Invitrogen, Carlsbad, CA) to enable expression. If necessary, a dengue serotype 2 virus-specific sequence encoded in a cDNA sequence may be amplified using a procedure such as the polymerase chain reaction (PCR). Alternatively, if the viral RNA is the source of the gene region, a DNA sequence may be amplified by a RT-PCR procedure. A DNA fragment including an initiation codon at the 5' end, and a termination codon at the 3' end is to be cloned into an expression vector at an appropriate restriction nuclease-specific site, in such a way that the cytomegalovirus (CMV) immediate early (IE) promoter, an initiation codon, and a terminator, are operably linked to the dengue serotype 2 virus sequence.

Example 17. Vaccination of mice using a dengue serotype 2 DNA vaccine. The dengue serotype 2 nucleic TU vaccine encoding the gene region from prM to E prepared in Example 16 is to be suspended in a suitable pharmaceutical carrier, such as water for injection or buffered physiological saline, and injected intramuscularly into groups of weanling mice. Control groups receive a comparable plasmid preparation lacking the dengue serotype 2 specific genes. The generation of dengue serotype 2-specific antibodies, and/or of dengue serotype 2-specific immune system cytotoxic cells, is to be assessed at fixed intervals thereafter, for example at weekly intervals. At

about two to four months after administration of the nucleic acid TU vaccine, mice are to be challenged with dengue serotype 2 virus. Levels of viremia are to be assessed at appropriate intervals thereafter, such as every second day. Passive protection by maternal antibody is to be assessed as indicated in Example 8.

Table 1. Transient expression of JE prM and E proteins by various recombinant plasmids in two transferred cell lines.

Vector					Recombinant Plasmid	IFA intensity/percentage of antigen-positive cells*	
	Promotor	Intron	Poly (A)	ORI		COS-1	COS-7
pCDNA3	CMV	No	BGH	SV40	pCDJE2-7	3+/40	3+/35
pCBamp	CMV	No	BGH	No	pCBJE1-14	3+/45	nd
pC1Bamp	CMV	Ycs	BGH	No	pC1BJES14	3+/39	nd
pCEP4	CMV	No	SV40	OriP	pCEJE	2+/4	2+/3
pREP4	RSV	No	SV40	OriP	pREJE	1+/3	1+/2
pRe/RSV	RSV	No	BGH	SV40	pRCJE	1+/3	1+/3
pCDNA3	CMV	No	BGH	SV40	pCDNA3/CAT	-	-

*Various cell lines were transformed with pCDNA3/CAT (negative control), pCDJE2-7, pCBJE1-14, pC1BJES14, pCEJEm pREJE, or pRCJE, Cells ere trypsinized 48 hours later and tested by an indirect immunofluorescent antibody assay (IFA) with JE virus-specific HIAF. Data are presented as the intensity (scale of 1+ to 4+) and the percentage of IFA positive cells. The pCDNA3/CAT transformed cells were used as the negative control.

Table 2. Characterization of proteins expressed by a pCDJE2-7 stably transformed clone (JE-4B) of COS-1 cells with JE virus-reactive antibodies.

Mab or antiserum	Biological Activity of Mab		Immunofluorescent intensity of cells	
	Specificity	Biological Function	JEV infected	4B
Mab:				
MC3	JEV Specific		2+	2+
2F2	JEV Specific	HI, N	4+	4+
112	JEV Specific		4+	4+
503	JEV Specific	N	4+	3+
109	Subgroup	HI	2+	1+
N.04	Subgroup	HI, N	3+	4+
201	Subgroup		1+	1+
203	Subgroup		4+	3+
204	Subgroup		2+	2+
301	Subgroup	HI	2+	2+
504	Flavivirus		4+	4+
6B6C-1	Flavivirus		2+	2÷
3B4C-4	VEE		-	-
H1AF:				
Anti-JEV			4+	3÷
Anti-WEE			-	-
PBS			-	-

Table 3. Persistence of the immune response in mice immunized with pCDJE2-7 or JE-VEX vaccine.

	ELISA Titer (log_{10})						$PRNT_{90\%}$ Titer		
	3 wks	6 wks	9 wks	23 wks	40 wks	60 wks*	3 wks	6 wks	9 wks
1x pCDJE2-7	2.6-3.2	3.8-5.0	3.8-4.4	>3.2	>3.2	2.4, 2.4, 3.8, 4.4	<20	20	40-160
2x pCDJE2-7	2.6-3.8	4.4	3.8-4.4	>3.2	>3.2	2.6, 3.8, 3.8	<20	20-40	40-160
2x JE-VAX	2.6-3.8	4.4-5.0	3.8-5.6	>3.2	>3.2	<2,<2,<2,4.4	<20	20-40	20-160
2x pCDNA3/CAT	<2	<2	<2	ND	ND	<2	<20	<20	<20

Mice were inoculated with 1 or 2, 100 µg/dose plasmid DNA, or 1/5 human dose of JE-VAX vaccine. Sera were collected for testing prior to the second immunization.

* Individual serum titers.

Table 4. The age-dependent percent seropositive rate in mice following vaccination with various JEV vaccines.

	3-day old		3-week old	
	3 weeks PV	7 weeks PV	3 weeks PV	7 weeks PV
JE-VAX	0	0	100	100
pCDNA3/CAT	0	0	0	0
pCDJE2-7	40	60	90	90
pC1BJES14	10	60	80	100
pCBJE1-14	80	100	100	100

Table 5. Protection from JEV challenge in 8 week old mice following vaccination at 3 days old with various JEV vaccines.

Vaccine	Pre-challenge JEV seroconversion	Days post-challenge survival rate (%)				
		6	7	8	9	21
JE-VAX	0	100	100	60	40	40
pCDNA3/CAT	0	100	80	30	30	30
pCDJE2-7	60	100	100	100	100	100
pC1BJES14	60	100	100	100	100	100
pCBJE1-14	100	100	100	100	100	100

Table 6. Evaluation of the ability of maternal antibody from JEV-nucleic acid-vaccinated female mice to protect their pups from fatal JEV encephalitis.

Vaccinated mother		JEV challenged pups		
Vaccine	$PRNT_{90\%}$	Challenge age (days)	No. survival[1]	ELISA[2]
1 x pCDJE2-7	40	4	0/11	
2 x pCDJE2-7	80	4	12/12	12/12
2 x JE-VAX	20	3	0/16	
2 x pCDNA-3/CAT	<10	5	0/14	
1 x pCDJE2-7	20	15	5/11	5/5
2 x pCDJE2-7	40	14	8/12	7/8
2 x JE-VAX	80	13	5/5	5/5
2 x pCDNA-3/CAT	<10	14	0/14	

Mice were inoculated intramuscularly with 1 or 2, 100 μg dose of plasmid DNA, or subcutaneously with two, 1/5 human dose of JE-VAX vaccine. Sera were collected 9 weeks post-vaccination for PRNT testing prior to mating with non-immune male.

[1]: No Survivors/total for each litter.

[2]: Number of JEV ELISA-antibody-positive animals (titer ≥ 1:400)/No. of survivors; sera were collected for testing 12 weeks after challenge.

What is claimed is:

1. An isolated nucleic acid comprising a transcriptional unit encoding a signal sequence of a structural protein of a first flavivirus and an immunogenic flavivirus antigen of a second flavivirus, wherein the transcriptional unit directs the synthesis of the antigen.

2. The nucleic acid of claim 1, wherein the signal sequence is a Japanese encephalitis virus signal sequence.

3. The nucleic acid of claim 1, wherein the immunogenic flavivirus antigen is of a flavivirus selected from the group consisting of yellow fever virus, dengue serotype 1 virus, dengue serotype 2 virus, dengue serotype 3 virus, dengue serotype 4 virus, Japanese encephalitis virus, Powassan virus and West Nile virus.

4. The nucleic acid of claim 1, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus and an M protein and an E protein of West Nile virus.

5. The nucleic acid of claim 1, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus and an M protein and an E protein of yellow fever virus.

6. The nucleic acid of claim 1, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus and an M protein and an E protein of St. Louis encephalitis virus.

7. The nucleic acid of claim 1, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus and an M protein and an E protein of Powassan virus.

8. The nucleic acid of claim 1, wherein the antigen is selected from the group consisting of an M protein of a flavivirus, an E protein of a flavivirus, both an M protein and an E protein of a flavivirus, a portion of an M protein of a flavivirus, a portion of an E protein of a flavivirus and both a portion of an M protein of a flavivirus and a portion of an E protein of a flavivirus or any combination thereof.

9. The nucleic acid of claim 8, wherein the antigen is both the M protein and the E protein of a flavivirus.

10. The nucleic acid of claim 1, wherein the nucleic acid is DNA.

11. The nucleic acid of claim 10, comprising a nucleotide sequence selected from the group consisting of SEQ ID NO:15, SEQ ID NO:19, SEQ ID NO:21 and SEQ ID NO:23.

12. The nuclcic acid of claim 1, wherein the transcriptional unit comprises a control sequence disposed appropriately such that it operably controls the synthesis of the antigen.

13. The nucleic acid of claim 12, wherein thc control sequence is the cytomegalovirus immediate early promoter.

14. The nucleic acid of claim 1, comprising a Kozak consensus sequence located at a translational start site for a polypeptide comprising the antigen encoded by the TU.

15. The nucleic acid of claim 1 wherein the transcriptional unit comprises a poly-A terminator.

16. A cell comprising the nucleic acid of claim 1.

17. A composition comprising the nucleic acid of claim 1 and a pharmaceutically acceptable carrier.

18. A method of immunizing a subject against infection by a flavivirus, comprising administering to the subject an effective amount of the composition of claim 17.

19. The method of claim 18, wherein the flavivirus antigen is of a flavivirus selected from the group consisting of yellow fever virus, dengue serotype 1 virus, dengue serotype 2 virus, dengue serotype 3 virus, dengue serotype 4 virus, Japanese encephalitis virus, Powassan virus and West Nile virus.

20. The method of claim 18, wherein the antigen is selected from the group consisting of an M protein of a flavivirus, an E protein of a flavivirus, both an M protein and an E protein of a flavivirus, a portion of an M protein of a flavivirus, a portion of an E protein of a flavivirus and both a portion of an M protein of a flavivirus and a portion of an E protein of a flavivirus or any combination thereof.

21. The method of claim 20, wherein the antigen is both the M protein and the E protein of a flavivirus, and wherein a cell within the body of the subject, after incorporating the nucleic acid within it, secretes subviral particles comprising the M protein and the E protein.

22. The method of claim 18, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus, and an M protein and an E protein of West Nile virus.

23. The method of claim 18, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus, and an M protein and an E protein of yellow fever virus.

24. The method of claim 18, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus, and an M protein and an E protein of St. Louis encephalitis virus.

25. The method of claim 18, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus, and an M protein and an E protein of Powassan virus.

26. The method of claim 18, comprising administering the composition to the subject in a single dose.

27. The method of claim 18, wherein the composition is administered via a parenteral route.

28. The nucleic acid of claim 1, wherein the antigen is a St. Louis encephalitis virus antigen.

29. The method of claim 18, wherein the antigen is a St. Louis encephalitis virus antigen.

30. The nucleic acid of claim 1, wherein the antigen is a Japanese encephalitis virus antigen.

31. The method of claim 18, wherein the antigen is a Japanese encephalitis virus antigen.

32. The nucleic acid of claim 1, wherein the antigen is a yellow fever virus antigen.

33. The method of claim 18, wherein the antigen is a yellow fever virus antigen.

34. The nucleic acid of claim 1, wherein the antigen is a dengue virus antigen.

35. The method of claim 18, wherein the antigen is a dengue virus antigen.

36. The nucleic acid of claim 1, wherein the antigen is a West Nile virus antigen.

37. The method of claim 18, wherein the antigen is a West Nile virus antigen.

38. An antigen produced from the nucleic acid of claim 1.

39. A method of detecting a flavivirus antibody in a sample, comprising:

(a) contacting the sample with the antigen of claim 38 under conditions whereby an antigen/antibody complex can form; and

(b) detecting antigen/antibody complex formation, thereby detecting a flavivirus antibody in the sample.

40. An antibody produced in response to immunization by the antigen of claim 38.

41. A method of detecting a flavivirus antigen in a sample, comprising:

(a) contacting the sample with the antibody of claim 40 under conditions whereby an antigen/antibody complex can form; and

(b) detecting antigen/antibody complex formation, thereby detecting a flavivirus antigen in a sample.

42. A method of diagnosing a flavivirus infection in a subject, comprising:

(a) contacting a sample from the subject with the antigen of claim 38 under conditions whereby an antigen/antibody complex can form; and

(b) detecting antigen/antibody complex formation, thereby diagnosing a flavivirus infection in a subject.

43. A method of diagnosing a flavivirus infection in a subject, comprising:

(a) contacting a sample from the subject with the antibody of claim 40 under conditions whereby an antigen/antibody complex can form; and

(b) detecting antigen/antibody complex formation, thereby diagnosing a flavivirus infection in a subject.

Abstract

The present invention encompasses isolated nucleic acids containing transcriptional units which encode a signal sequence of one flavivirus and an immunogenic flavivirus antigen of a second flavivirus. The invention further encompasses a nucleic acid and protein vaccine and the use of the vaccine to immunize a subject against flavivirus infection. The invention also provides antigens encoded by nucleic acids of the invention, antibodies elicited in response to the antigens and use of the antigens and/or antibodies in detecting flavivirus or diagnosing flavivirus infection.

DECLARATION
FOR PATENT APPLICATION

(X) Original () Supplemental () Substitute () PCT

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am an original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled **"NUCLEIC ACID VACCINES FOR PREVENTION OF FLAVIVIRUS INFECTION,"** which is described and claimed in the specification

(check one) [X] which is attached hereto, or
[] which was filed on , as Application Serial No. and with amendments through (if applicable), or
[] in International Application No. PCT/, filed , and as amended on (if applicable).

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information known by me to be material to the patentability of the claims of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code §119 (a)-(d) or §365(b) of any foreign application(s) for patent or inventor's certificate, or §365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or of any PCT international application having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN APPLICATIONS: (ENTER BELOW IF APPLICABLE)			PRIORITY CLAIMED (MARK APPROPRIATE BOX BELOW)	
APP. NUMBER	COUNTRY	DAY/MONTH/YEAR FILED	YES	NO
PCT/US99/12298	PCT	3 June 1999	X	

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below.

APPLICATION NUMBER	FILING DATE
60/087,908	4 June 1998

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose all information known by me to be material to the patentability of the claims of this application as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

APPLICATION SERIAL NO.	FILING DATE	STATUS (MARK APPROPRIATE COLUMN BELOW)		
		PATENTED	PENDING	ABANDONED
09/701,536	11/29/00		X	

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of first inventor: GWONG-JEN J. CHANG

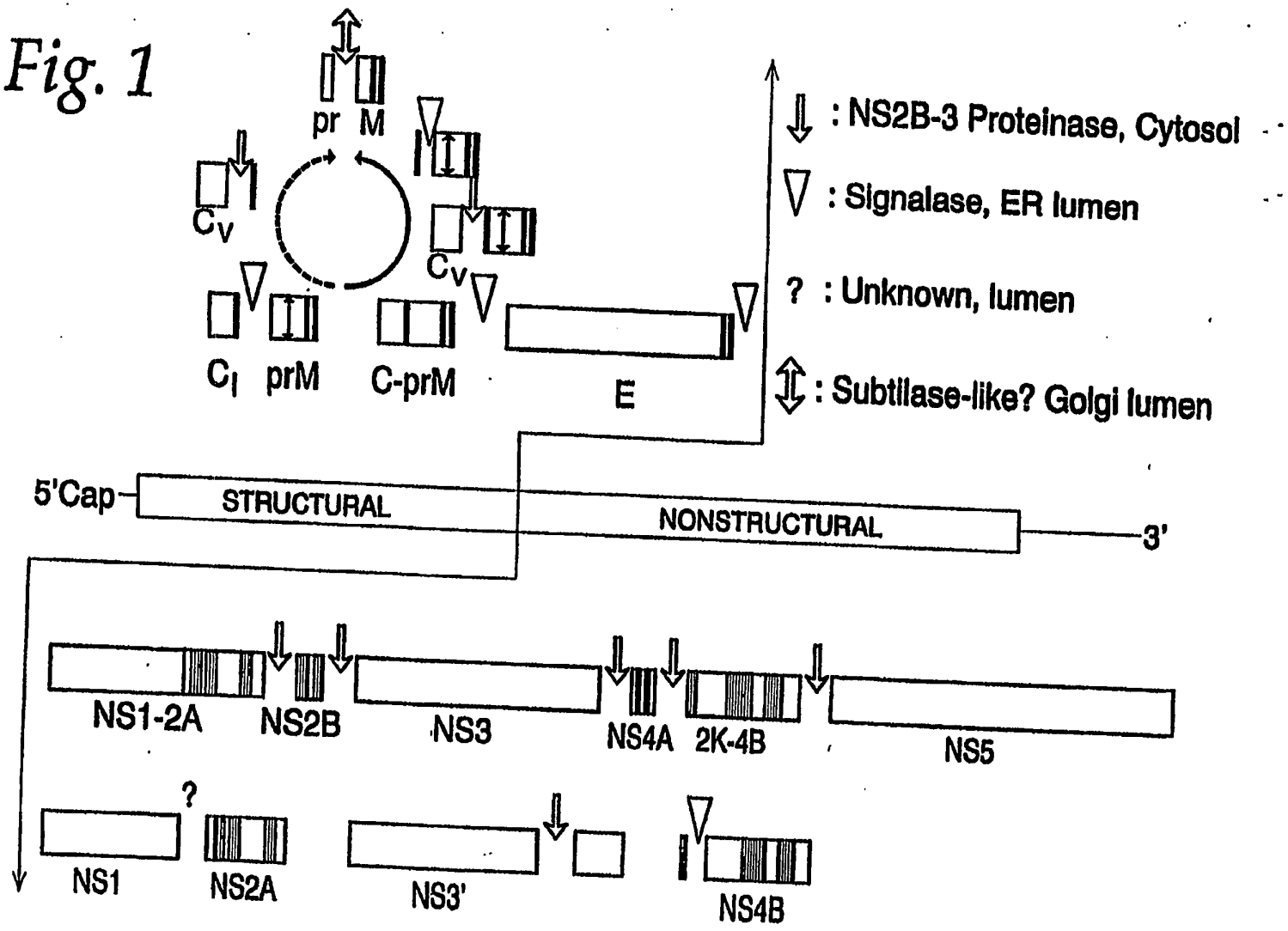
Inventor's signature: Gwong-Jen Chang Date: April 2, 2001

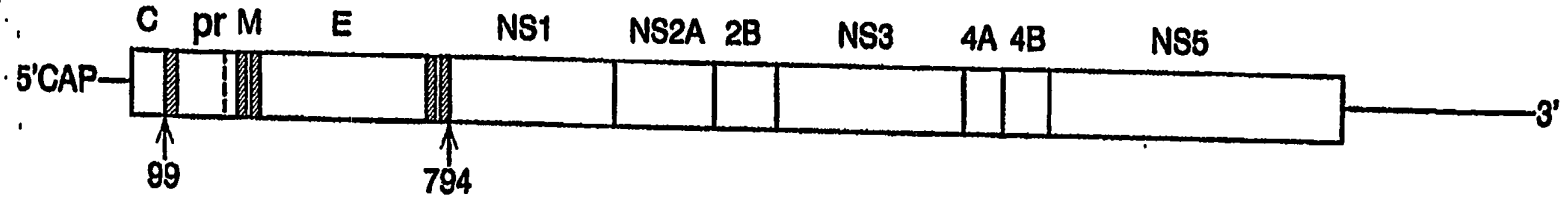
Residence: 4237 Beaver Creek Drive, Fort Collins, CO 80526

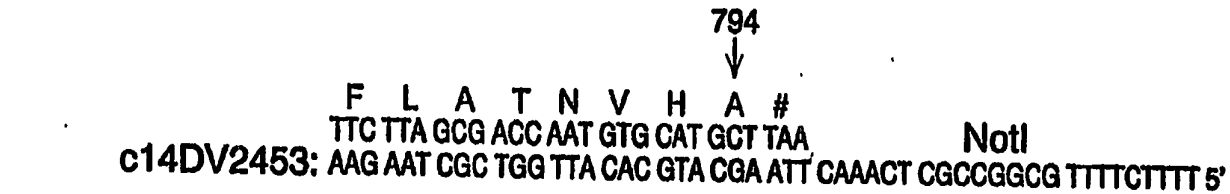
Post Office Address: 4237 Beaver Creek Drive, Fort Collins, CO 80526

Citizenship: USA

Fig. 1


pr
M
Cv
Cv
Cl
prM
C-prM
E
: NS2B-3 Proteinase, Cytosol
: Signalase, ER lumen
? : Unknown, lumen
: Subtilase-like? Golgi lumen
5'Cap
STRUCTURAL
NONSTRUCTURAL
3'
NS1-2A
NS2B
NS3
NS4A
2K-4B
NS5
?
NS1
NS2A
NS3'
NS4B





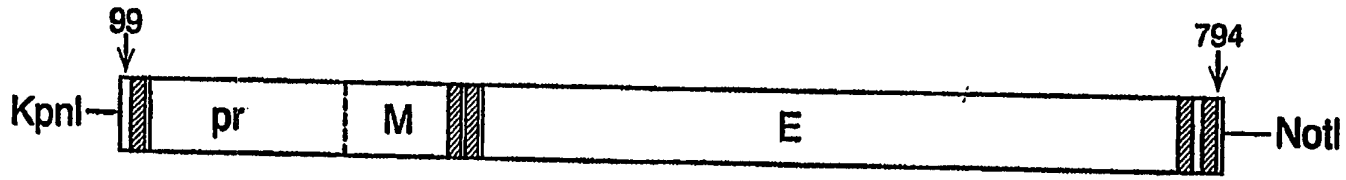


Fig. 2

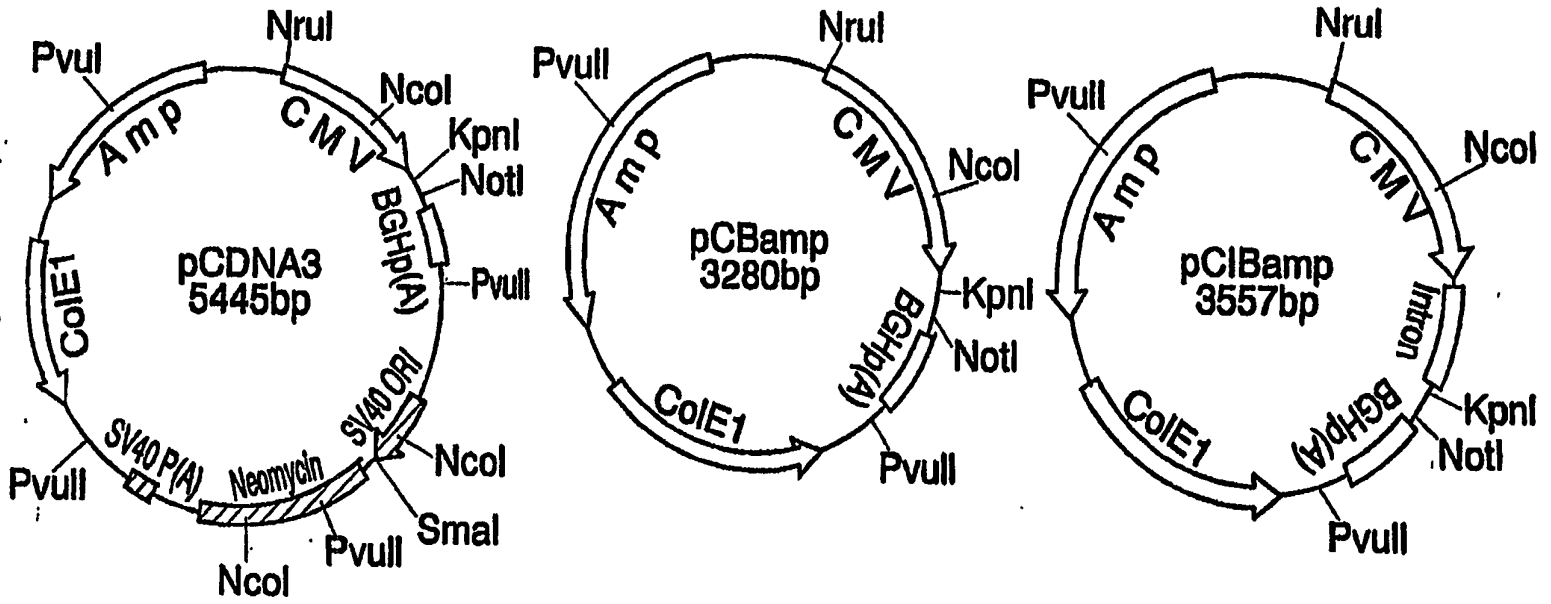

pCDNA3
5445bp
Amp
CMV
ColE1
BGHp(A)
SV40 ORI
SV40 P(A)
Neomycin
PvuI
NruI
NcoI
KpnI
NotI
PvuII
SmaI
pCBamp
3280bp
pCIBamp
3557bp
Intron

Fig. 3

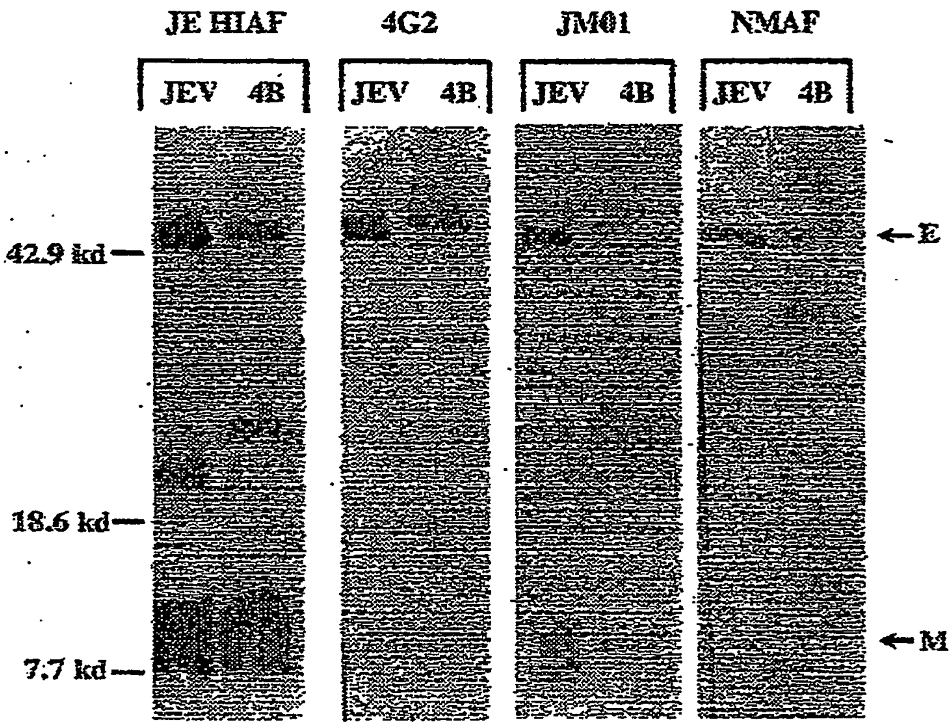

JE HIAF
4G2
JM01
NMAF
JEV
4B
JEV
4B
JEV
4B
JEV
4B
42.9 kd
18.6 kd
7.7 kd
E
M

Fig. 4

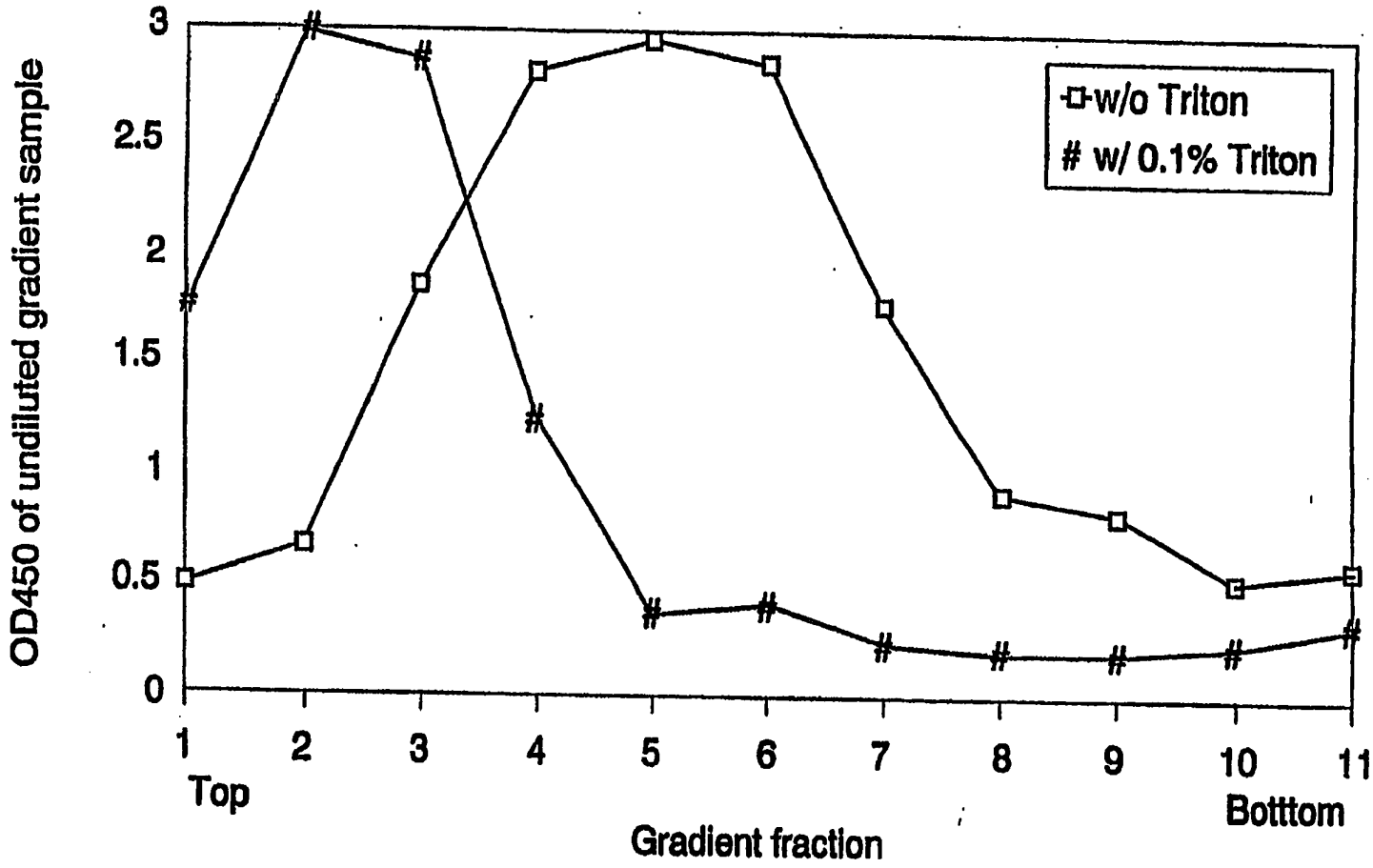


Fig. 5

SEQUENCE LISTING

<110> Chang, Gwong-Jen J

<120> Nucleic Acid Vaccines for Prevention of
Flavivirus Infection

<130> 14114.0332U3

<150> PCT/US99/12298
<151> 1999-06-03

<150> 09/701,536
<151> 2000-11-29

<150> 60/087,908
<151> 1998-06-04

<160> 31

<170> FastSEQ for Windows Version 4.0

<210> 1
<211> 48
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (25)...(48)

<221> misc_feature
<222> 1-48
<223> Amplimer 14DV389

<400> 1
cttggtacct ctagagccgc cgcc atg ggc aga aag caa aac aaa aga 48
Met Gly Arg Lys Gln Asn Lys Arg
1 5

<210> 2
<211> 8
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<400> 2
Met Gly Arg Lys Gln Asn Lys Arg
1 5

<210> 3
<211> 50
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-50
<223> Amplimer c14DV2453

<400> 3
ttttcttttg cggccgctca aacttaagca tgcacattgg tcgctaagaa 50

<210> 4
<211> 48
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (25)...(48)

<221> misc_feature
<222> (1)...(48)
<223> Amplimer YFDV389

<400> 4
cttggtacct ctagagccgc cgcc atg cgt tcc cat gat gtt ctg act 48
Met Arg Ser His Asp Val Leu Thr
1 5

<210> 5
<211> 8
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<400> 5
Met Arg Ser His Asp Val Leu Thr
1 5

<210> 6
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-41
<223> Amplimer cYFDV2452

<400> 6
ttttcttttg cggccgctca cgccccaact cctagagaaa c 41

<210> 7
<211> 51
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (25)...(54)

<221> misc_feature
<222> 1-54
<223> Amplimer SLEDV410

<400> 7
cttggtacct ctagagccgc cgcc atg tct aaa aaa aga gga ggg acc aga 51
Met Ser Lys Lys Arg Gly Gly Thr Arg
1 5

<210> 8
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<400> 8
Met Ser Lys Lys Arg Gly Gly Thr Arg
1 5

<210> 9
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-38
<223> Amplimer cSLEDV2449

<400> 9
ttttcttttg cggccgctta ggcttgcacg ctggttgc 38

<210> 10
<211> 7500
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (916)...(3009)

<221> misc_feature
<222> 1-7500
<223> pCDJE 2-7

<400> 10
gacggatcgg gagatctccc gatcccctat ggtcgactct cagtacaatc tgctctgatg 60
ccgcatagtt aagccagtat ctgctccctg cttgtgtgtt ggaggtcgct gagtagtgcg 120
cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc 180
ttagggttag gcgttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt 240
gattattgac tagttattaa tagtaatcaa ttacggggtc attagttcat agcccatata 300
tggagttccg cgttacataa cttacggtaa atggcccgcc tggctgaccg cccaacgacc 360
cccgcccatt gacgtcaata atgacgtatg ttcccatagt aacgccaata gggactttcc 420
attgacgtca atgggtggac tatttacggt aaactgccca cttggcagta catcaagtgt 480
atcatatgcc aagtacgccc cctattgacg tcaatgacgg taaatggccc gcctggcatt 540
atgcccagta catgacctta tgggactttc ctacttggca gtacatctac gtattagtca 600
tcgctattac catggtgatg cggttttggc agtacatcaa tgggcgtgga tagcggtttg 660
actcacgggg atttccaagt ctccacccca ttgacgtcaa tgggagtttg ttttggcacc 720
aaaatcaacg ggactttcca aaatgtcgta acaactccgc cccattgacg caaatgggcg 780
gtaggcgtgt acggtgggag gtctatataa gcagagctct ctggctaact agagaaccca 840
ctgcttactg gcttatcgaa attaatacga ctcactatag ggagacccaa gcttggtacc 900

```
gagctcgccg ccgcc atg ggc aga aag caa aac aaa aga gga gga aat gaa      951
                Met Gly Arg Lys Gln Asn Lys Arg Gly Gly Asn Glu
                 1               5                  10

ggc tca atc atg tgg ctc gcg agc ttg gca gtt gtc ata gct tgt gcg      999
Gly Ser Ile Met Trp Leu Ala Ser Leu Ala Val Val Ile Ala Cys Ala
         15                  20                  25

gga gcc atg aag ttg tcg aat ttc cag ggg aag ctt ttg atg acc atc     1047
Gly Ala Met Lys Leu Ser Asn Phe Gln Gly Lys Leu Leu Met Thr Ile
     30                  35                  40

aac aac acg gac att gca gac gtt atc gtg att ccc acc tca aaa gga     1095
Asn Asn Thr Asp Ile Ala Asp Val Ile Val Ile Pro Thr Ser Lys Gly
 45                  50                  55                  60

gag aac aga tgc tgg gtc cgg gca atc gac gtc ggc tac atg tgt gag     1143
Glu Asn Arg Cys Trp Val Arg Ala Ile Asp Val Gly Tyr Met Cys Glu
                 65                  70                  75

gac act atc acg tac gaa tgt cct aag ctt acc atg ggc aat gat cca     1191
Asp Thr Ile Thr Tyr Glu Cys Pro Lys Leu Thr Met Gly Asn Asp Pro
             80                  85                  90

gag gat gtg gat tgc tgg tgt gac aac caa gaa gtc tac gtc caa tat     1239
Glu Asp Val Asp Cys Trp Cys Asp Asn Gln Glu Val Tyr Val Gln Tyr
         95                  100                 105

gga cgg tgc acg cgg acc agg cat tcc aag cga agc agg aga tcc gtg     1287
Gly Arg Cys Thr Arg Thr Arg His Ser Lys Arg Ser Arg Arg Ser Val
    110                 115                 120

tcg gtc caa aca cat ggg gag agt tca cta gtg aat aaa aaa gag gct     1335
Ser Val Gln Thr His Gly Glu Ser Ser Leu Val Asn Lys Lys Glu Ala
125                 130                 135                 140

tgg ctg gat tca acg aaa gcc aca cga tat ctc atg aaa act gag aac     1383
Trp Leu Asp Ser Thr Lys Ala Thr Arg Tyr Leu Met Lys Thr Glu Asn
                145                 150                 155

tgg atc ata agg aat cct ggc tat gct ttc ctg gcg gcg gta ctt ggc     1431
Trp Ile Ile Arg Asn Pro Gly Tyr Ala Phe Leu Ala Ala Val Leu Gly
            160                 165                 170

tgg atg ctt ggc agt aac aac ggt caa cgc gtg gta ttt acc atc ctc     1479
Trp Met Leu Gly Ser Asn Asn Gly Gln Arg Val Val Phe Thr Ile Leu
        175                 180                 185

ctg ctg ttg gtc gct ccg gct tac agt ttt aat tgt ctg gga atg ggc     1527
Leu Leu Leu Val Ala Pro Ala Tyr Ser Phe Asn Cys Leu Gly Met Gly
    190                 195                 200
```

```
aat cgt gac ttc ata gaa gga gcc agt gga gcc act tgg gtg gac ttg     1575
Asn Arg Asp Phe Ile Glu Gly Ala Ser Gly Ala Thr Trp Val Asp Leu
205                 210                 215                 220

gtg ctg gaa gga gat agc tgc ttg aca atc atg gca aac gac aaa cca     1623
Val Leu Glu Gly Asp Ser Cys Leu Thr Ile Met Ala Asn Asp Lys Pro
                225                 230                 235

aca ttg gac gtc cgc atg att aac atc gaa gct agc caa ctt gct gag     1671
Thr Leu Asp Val Arg Met Ile Asn Ile Glu Ala Ser Gln Leu Ala Glu
            240                 245                 250

gtc aga agt tac tgc tat cat gct tca gtc act gac atc tcg acg gtg     1719
Val Arg Ser Tyr Cys Tyr His Ala Ser Val Thr Asp Ile Ser Thr Val
        255                 260                 265

gct cgg tgc ccc acg act gga gaa gcc cac aac gag aag cga gct gat     1767
Ala Arg Cys Pro Thr Thr Gly Glu Ala His Asn Glu Lys Arg Ala Asp
    270                 275                 280

agt agc tat gtg tgc aaa caa ggc ttc act gac cgt ggg tgg ggc aac     1815
Ser Ser Tyr Val Cys Lys Gln Gly Phe Thr Asp Arg Gly Trp Gly Asn
285                 290                 295                 300

gga tgt gga ctt ttc ggg aag gga agc att gac aca tgt gca aaa ttc     1863
Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile Asp Thr Cys Ala Lys Phe
                305                 310                 315

tcc tgc acc agt aaa gcg att ggg aga aca atc cag cca gaa aac atc     1911
Ser Cys Thr Ser Lys Ala Ile Gly Arg Thr Ile Gln Pro Glu Asn Ile
            320                 325                 330

aaa tac gaa gtt ggc att ttt gtg cat gga acc acc act tcg gaa aac     1959
Lys Tyr Glu Val Gly Ile Phe Val His Gly Thr Thr Thr Ser Glu Asn
        335                 340                 345

cat ggg aat tat tca gcg caa gtt ggg gcg tcc cag gcg gca aag ttt     2007
His Gly Asn Tyr Ser Ala Gln Val Gly Ala Ser Gln Ala Ala Lys Phe
    350                 355                 360

aca gta aca ccc aat gct cct tcg ata acc ctc aaa ctt ggt gac tac     2055
Thr Val Thr Pro Asn Ala Pro Ser Ile Thr Leu Lys Leu Gly Asp Tyr
365                 370                 375                 380

gga gaa gtc aca ctg gac tgt gag cca agg agt gga ctg aac act gaa     2103
Gly Glu Val Thr Leu Asp Cys Glu Pro Arg Ser Gly Leu Asn Thr Glu
                385                 390                 395

gcg ttt tac gtc atg acc gtg ggg tca aag tca ttt ctg gtc cat agg     2151
Ala Phe Tyr Val Met Thr Val Gly Ser Lys Ser Phe Leu Val His Arg
            400                 405                 410

gag tgg ttt cat gac ctc gct ctc ccc tgg acg tcc cct tcg agc aca     2199
```

```
Glu Trp Phe His Asp Leu Ala Leu Pro Trp Thr Ser Pro Ser Ser Thr
        415                 420                 425

gcg tgg aga aac aga gaa ctc ctc atg gaa ttt gaa gag gcg cac gcc     2247
Ala Trp Arg Asn Arg Glu Leu Leu Met Glu Phe Glu Glu Ala His Ala
    430                 435                 440

aca aaa cag tcc gtt gtt gct ctt ggg tca cag gaa gga ggc ctc cat     2295
Thr Lys Gln Ser Val Val Ala Leu Gly Ser Gln Glu Gly Gly Leu His
445                 450                 455                 460

cag gcg ttg gca gga gcc atc gtg gtg gag tac tca agc tca gtg aag     2343
Gln Ala Leu Ala Gly Ala Ile Val Val Glu Tyr Ser Ser Ser Val Lys
                465                 470                 475

tta aca tca ggc cac ctg aaa tgt agg ctg aaa atg gac aaa ctg gct     2391
Leu Thr Ser Gly His Leu Lys Cys Arg Leu Lys Met Asp Lys Leu Ala
            480                 485                 490

ctg aaa ggc aca acc tat ggc atg tgt aca gaa aaa ttc tcg ttc gcg     2439
Leu Lys Gly Thr Thr Tyr Gly Met Cys Thr Glu Lys Phe Ser Phe Ala
        495                 500                 505

aaa aat ccg gcg gac act ggt cac gga aca gtt gtc att gaa ctc tcc     2487
Lys Asn Pro Ala Asp Thr Gly His Gly Thr Val Val Ile Glu Leu Ser
    510                 515                 520

tac tct ggg agt gat ggc ccc tgc aaa att ccg att gct tcc gtt gcg     2535
Tyr Ser Gly Ser Asp Gly Pro Cys Lys Ile Pro Ile Ala Ser Val Ala
525                 530                 535                 540

agc ctc aat gac atg acc ccc gtt ggg cgg ctg gtg aca gtg aac ccc     2583
Ser Leu Asn Asp Met Thr Pro Val Gly Arg Leu Val Thr Val Asn Pro
                545                 550                 555

ttc gtc gcg act tcc agt gcc agc tca aag gtg ctg gtc gag atg gaa     2631
Phe Val Ala Thr Ser Ser Ala Ser Ser Lys Val Leu Val Glu Met Glu
            560                 565                 570

ccc ccc ttc gga gac tcc tac atc gta gtt gga agg gga gac aag cag     2679
Pro Pro Phe Gly Asp Ser Tyr Ile Val Val Gly Arg Gly Asp Lys Gln
        575                 580                 585

atc aac cac cat tgg cac aaa gct gga agc acg ctg ggc aag gcc ttt     2727
Ile Asn His His Trp His Lys Ala Gly Ser Thr Leu Gly Lys Ala Phe
    590                 595                 600

tca aca act ttg aag gga gct caa aga ctg gca gcg ttg ggc gac aca     2775
Ser Thr Thr Leu Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr
605                 610                 615                 620

gcc tgg gac ttt ggc tct att gga ggg gtc ttc aac tcc ata gga aaa     2823
Ala Trp Asp Phe Gly Ser Ile Gly Gly Val Phe Asn Ser Ile Gly Lys
                625                 630                 635
```

```
gcc gtt cac caa gtg ttt ggt ggt gcc ttc aga aca ctc ttt ggg gga     2871
Ala Val His Gln Val Phe Gly Gly Ala Phe Arg Thr Leu Phe Gly Gly
            640                 645                 650

atg tct tgg atc aca caa ggg cta atg ggt gcc cta ctg ctc tgg atg     2919
Met Ser Trp Ile Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met
        655                 660                 665

ggc gtc aac gca cga gac cga tca att gct ttg gcc ttc tta gcc aca     2967
Gly Val Asn Ala Arg Asp Arg Ser Ile Ala Leu Ala Phe Leu Ala Thr
    670                 675                 680

ggg ggt gtg ctc gtg ttc tta gcg acc aat gtg cat gct taa             3009
Gly Gly Val Leu Val Phe Leu Ala Thr Asn Val His Ala  *
685                 690                 695

ttagtttgag cggccgctcg agcatgcatc tagagggccc tattctatag tgtcacctaa   3069
atgctagagc tcgctgatca gcctcgactg tgccttctag ttgccagcca tctgttgttt   3129
gcccctcccc cgtgccttcc ttgaccctgg aaggtgccac tcccactgtc ctttcctaat   3189
aaaatgagga aattgcatcg cattgtctga gtaggtgtca ttctattctg gggggtgggg   3249
tggggcagga cagcaagggg gaggattggg aagacaatag caggcatgct ggggatgcgg   3309
tgggctctat ggcttctgag gcggaaagaa ccagctgggg ctctaggggg tatccccacg   3369
cgccctgtag cggcgcatta agcgcggcgg gtgtggtggt tacgcgcagc gtgaccgcta   3429
cacttgccag cgccctagcg cccgctcctt tcgctttctt cccttccttt ctcgccacgt   3489
tcgccggctt tccccgtcaa gctctaaatc ggggcatccc tttagggttc cgatttagtg   3549
ctttacggca cctcgacccc aaaaaacttg attagggtga tggttcacgt agtgggccat   3609
cgccctgata gacggttttt cgccctttga cgttggagtc cacgttcttt aatagtggac   3669
tcttgttcca aactggaaca acactcaacc ctatctcggt ctattctttt gatttataag   3729
ggattttggg gatttcggcc tattggttaa aaaatgagct gatttaacaa aaatttaacg   3789
cgaattaatt ctgtggaatg tgtgtcagtt agggtgtgga aagtccccag gctccccagg   3849
caggcagaag tatgcaaagc atgcatctca attagtcagc aaccaggtgt ggaaagtccc   3909
caggctcccc agcaggcaga agtatgcaaa gcatgcatct caattagtca gcaaccatag   3969
tcccgccccc aactccgccc atcccgcccc taactccgcc cagttccgcc cattctccgc   4029
cccatggctg actaattttt tttatttatg cagaggccga ggccgcctct gcctctgagc   4089
tattccagaa gtagtgagga ggcttttttg gaggcctagg cttttgcaaa aagctcccgg   4149
gagcttgtat atccatttcc ggatctgatc aagagacagg atgaggatcg tttcgcatga   4209
ttgaacaaga tggattgcac gcaggttctc cggccgcttg ggtggagagg ctattcggct   4269
atgactgggc acaacagaca atcggctgct ctgatgccgc cgtgttccgg ctgtcagcgc   4329
aggggcgccc ggttcttttt gtcaagaccg acctgtccgg tgccctgaat gaactgcagg   4389
acgaggcagc gcggctatcg tggctggcca cgacgggcgt tccttgcgca gctgtgctcg   4449
acgttgtcac tgaagcggga agggactggc tgctattggg cgaagtgccg gggcaggatc   4509
tcctgtcatc tcaccttgct cctgccgaga aagtatccat catggctgat gcaatgcggc   4569
ggctgcatac gcttgatccg gctacctgcc cattcgacca ccaagcgaaa catcgcatcg   4629
agcgagcacg tactcggatg gaagccggtc ttgtcgatca ggatgatctg gacgaagagc   4689
atcaggggct cgcgccagcc gaactgttcg ccaggctcaa ggcgcgcatg cccgacggcg   4749
aggatctcgt cgtgacccat ggcgatgcct gcttgccgaa tatcatggtg gaaaatggcc   4809
gcttttctgg attcatcgac tgtggccggc tgggtgtggc ggaccgctat caggacatag   4869
cgttggctac ccgtgatatt gctgaagagc ttggcggcga atgggctgac cgcttcctcg   4929
tgctttacgg tatcgccgct cccgattcgc agcgcatcgc cttctatcgc cttcttgacg   4989
agttcttctg agcgggactc tggggttcga aatgaccgac caagcgacgc ccaacctgcc   5049
atcacgagat ttcgattcca ccgccgcctt ctatgaaagg ttgggcttcg gaatcgtttt   5109
ccgggacgcc ggctggatga tcctccagcg cggggatctc atgctggagt tcttcgccca   5169
ccccaacttg tttattgcag cttataatgg ttacaaataa agcaatagca tcacaaattt   5229
```

cacaaataaa gcatttttt cactgcattc tagttgtggt ttgtccaaac tcatcaatgt 5289
atcttatcat gtctgtatac cgtcgacctc tagctagagc ttggcgtaat catggtcata 5349
gctgtttcct gtgtgaaatt gttatccgct cacaattcca cacaacatac gagccggaag 5409
cataaagtgt aaagcctggg gtgcctaatg agtgagctaa ctcacattaa ttgcgttgcg 5469
ctcactgccc gctttccagt cgggaaacct gtcgtgccag ctgcattaat gaatcggcca 5529
acgcgcgggg agaggcggtt tgcgtattgg gcgctcttcc gcttcctcgc tcactgactc 5589
gctgcgctcg gtcgttcggc tgcggcgagc ggtatcagct cactcaaagg cggtaatacg 5649
gttatccaca gaatcagggg ataacgcagg aaagaacatg tgagcaaaag gccagcaaaa 5709
ggccaggaac cgtaaaaagg ccgcgttgct ggcgtttttc cataggctcc gccccctga 5769
cgagcatcac aaaaatcgac gctcaagtca gaggtggcga aacccgacag gactataaag 5829
ataccaggcg tttccccctg gaagctccct cgtgcgctct cctgttccga ccctgccgct 5889
taccggatac ctgtccgcct ttctcccttc gggaagcgtg gcgctttctc aatgctcacg 5949
ctgtaggtat ctcagttcgg tgtaggtcgt tcgctccaag ctgggctgtg tgcacgaacc 6009
ccccgttcag cccgaccgct gcgccttatc cggtaactat cgtcttgagt ccaacccggt 6069
aagacacgac ttatcgccac tggcagcagc cactggtaac aggattagca gagcgaggta 6129
tgtaggcggt gctacagagt tcttgaagtg gtggcctaac tacggctaca ctagaaggac 6189
agtatttggt atctgcgctc tgctgaagcc agttaccttc ggaaaaagag ttggtagctc 6249
ttgatccggc aaacaaacca ccgctggtag cggtggtttt tttgtttgca agcagcagat 6309
tacgcgcaga aaaaaaggat ctcaagaaga tcctttgatc ttttctacgg ggtctgacgc 6369
tcagtggaac gaaaactcac gttaagggat tttggtcatg agattatcaa aaaggatctt 6429
cacctagatc cttttaaatt aaaaatgaag ttttaaatca atctaaagta tatatgagta 6489
aacttggtct gacagttacc aatgcttaat cagtgaggca cctatctcag cgatctgtct 6549
atttcgttca tccatagttg cctgactccc cgtcgtgtag ataactacga tacgggaggg 6609
cttaccatct ggccccagtg ctgcaatgat accgcgagac ccacgctcac cggctccaga 6669
tttatcagca ataaaccagc cagccggaag ggccgagcgc agaagtggtc ctgcaacttt 6729
atccgcctcc atccagtcta ttaattgttg ccgggaagct agagtaagta gttcgccagt 6789
taatagtttg cgcaacgttg ttgccattgc tacaggcatc gtggtgtcac gctcgtcgtt 6849
tggtatggct tcattcagct ccggttccca acgatcaagg cgagttacat gatcccccat 6909
gttgtgcaaa aaagcggtta gctccttcgg tcctccgatc gttgtcagaa gtaagttggc 6969
cgcagtgtta tcactcatgg ttatggcagc actgcataat tctcttactg tcatgccatc 7029
cgtaagatgc ttttctgtga ctggtgagta ctcaaccaag tcattctgag aatagtgtat 7089
gcggcgaccg agttgctctt gcccggcgtc aatacgggat aataccgcgc cacatagcag 7149
aactttaaaa gtgctcatca ttggaaaacg ttcttcgggg cgaaaactct caaggatctt 7209
accgctgttg agatccagtt cgatgtaacc cactcgtgca cccaactgat cttcagcatc 7269
ttttactttc accagcgttt ctgggtgagc aaaaacagga aggcaaaatg ccgcaaaaaa 7329
gggaataagg gcgacacgga aatgttgaat actcatactc ttcctttttc aatattattg 7389
aagcatttat cagggttatt gtctcatgag cggatacata tttgaatgta tttagaaaaa 7449
taaacaaata ggggttccgc gcacatttcc ccgaaaagtg ccacctgacg t 7500

<210> 11
<211> 697
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<223> pCDJE 2-7

<400> 11

```
Met Gly Arg Lys Gln Asn Lys Arg Gly Gly Asn Glu Gly Ser Ile Met
 1               5                  10                  15
Trp Leu Ala Ser Leu Ala Val Val Ile Ala Cys Ala Gly Ala Met Lys
            20                  25                  30
Leu Ser Asn Phe Gln Gly Lys Leu Leu Met Thr Ile Asn Asn Thr Asp
        35                  40                  45
Ile Ala Asp Val Ile Val Ile Pro Thr Ser Lys Gly Glu Asn Arg Cys
    50                  55                  60
Trp Val Arg Ala Ile Asp Val Gly Tyr Met Cys Glu Asp Thr Ile Thr
65                  70                  75                  80
Tyr Glu Cys Pro Lys Leu Thr Met Gly Asn Asp Pro Glu Asp Val Asp
                85                  90                  95
Cys Trp Cys Asp Asn Gln Glu Val Tyr Val Gln Tyr Gly Arg Cys Thr
            100                 105                 110
Arg Thr Arg His Ser Lys Arg Ser Arg Arg Ser Val Ser Val Gln Thr
        115                 120                 125
His Gly Glu Ser Ser Leu Val Asn Lys Lys Glu Ala Trp Leu Asp Ser
    130                 135                 140
Thr Lys Ala Thr Arg Tyr Leu Met Lys Thr Glu Asn Trp Ile Ile Arg
145                 150                 155                 160
Asn Pro Gly Tyr Ala Phe Leu Ala Ala Val Leu Gly Trp Met Leu Gly
                165                 170                 175
Ser Asn Asn Gly Gln Arg Val Val Phe Thr Ile Leu Leu Leu Leu Val
            180                 185                 190
Ala Pro Ala Tyr Ser Phe Asn Cys Leu Gly Met Gly Asn Arg Asp Phe
        195                 200                 205
Ile Glu Gly Ala Ser Gly Ala Thr Trp Val Asp Leu Val Leu Glu Gly
    210                 215                 220
Asp Ser Cys Leu Thr Ile Met Ala Asn Asp Lys Pro Thr Leu Asp Val
225                 230                 235                 240
Arg Met Ile Asn Ile Glu Ala Ser Gln Leu Ala Glu Val Arg Ser Tyr
                245                 250                 255
Cys Tyr His Ala Ser Val Thr Asp Ile Ser Thr Val Ala Arg Cys Pro
            260                 265                 270
Thr Thr Gly Glu Ala His Asn Glu Lys Arg Ala Asp Ser Ser Tyr Val
        275                 280                 285
Cys Lys Gln Gly Phe Thr Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu
    290                 295                 300
Phe Gly Lys Gly Ser Ile Asp Thr Cys Ala Lys Phe Ser Cys Thr Ser
305                 310                 315                 320
Lys Ala Ile Gly Arg Thr Ile Gln Pro Glu Asn Ile Lys Tyr Glu Val
                325                 330                 335
Gly Ile Phe Val His Gly Thr Thr Thr Ser Glu Asn His Gly Asn Tyr
            340                 345                 350
Ser Ala Gln Val Gly Ala Ser Gln Ala Ala Lys Phe Thr Val Thr Pro
        355                 360                 365
Asn Ala Pro Ser Ile Thr Leu Lys Leu Gly Asp Tyr Gly Glu Val Thr
    370                 375                 380
Leu Asp Cys Glu Pro Arg Ser Gly Leu Asn Thr Glu Ala Phe Tyr Val
385                 390                 395                 400
Met Thr Val Gly Ser Lys Ser Phe Leu Val His Arg Glu Trp Phe His
                405                 410                 415
```

```
Asp Leu Ala Leu Pro Trp Thr Ser Pro Ser Ser Thr Ala Trp Arg Asn
            420                 425                 430
Arg Glu Leu Leu Met Glu Phe Glu Glu Ala His Ala Thr Lys Gln Ser
        435                 440                 445
Val Val Ala Leu Gly Ser Gln Glu Gly Gly Leu His Gln Ala Leu Ala
    450                 455                 460
Gly Ala Ile Val Val Glu Tyr Ser Ser Ser Val Lys Leu Thr Ser Gly
465                 470                 475                 480
His Leu Lys Cys Arg Leu Lys Met Asp Lys Leu Ala Leu Lys Gly Thr
                485                 490                 495
Thr Tyr Gly Met Cys Thr Glu Lys Phe Ser Phe Ala Lys Asn Pro Ala
            500                 505                 510
Asp Thr Gly His Gly Thr Val Val Ile Glu Leu Ser Tyr Ser Gly Ser
        515                 520                 525
Asp Gly Pro Cys Lys Ile Pro Ile Ala Ser Val Ala Ser Leu Asn Asp
    530                 535                 540
Met Thr Pro Val Gly Arg Leu Val Thr Val Asn Pro Phe Val Ala Thr
545                 550                 555                 560
Ser Ser Ala Ser Ser Lys Val Leu Val Glu Met Glu Pro Pro Phe Gly
                565                 570                 575
Asp Ser Tyr Ile Val Val Gly Arg Gly Asp Lys Gln Ile Asn His His
            580                 585                 590
Trp His Lys Ala Gly Ser Thr Leu Gly Lys Ala Phe Ser Thr Thr Leu
        595                 600                 605
Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr Ala Trp Asp Phe
    610                 615                 620
Gly Ser Ile Gly Gly Val Phe Asn Ser Ile Gly Lys Ala Val His Gln
625                 630                 635                 640
Val Phe Gly Gly Ala Phe Arg Thr Leu Phe Gly Gly Met Ser Trp Ile
                645                 650                 655
Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met Gly Val Asn Ala
            660                 665                 670
Arg Asp Arg Ser Ile Ala Leu Ala Phe Leu Ala Thr Gly Gly Val Leu
        675                 680                 685
Val Phe Leu Ala Thr Asn Val His Ala
    690                 695
```

<210> 12
<211> 46
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-46
<223> WN 466

<400> 12
cttggtaccc gtctcggcgc cgtgaccctc tcgaacttcc agggca 46

<210> 13
<211> 43
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-43
<223> CWN2444

<400> 13
agaggcactt gcacgtgcgg acttccgccg gcgaaaaaga aaa 43

<210> 14
<211> 24
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<223> JE Signal

<400> 14
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser Leu Ala
1 5 10 15
Val Val Ile Ala Cys Ala Gly Ala
20

<210> 15
<211> 5308
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (911)...(2987)

<221> misc_feature
<222> (1)...(5308)
<223> pCBWN

<400> 15
gacggatcgg gagatctccc gatcccctat ggtgcactct cagtacaatc tgctctgatg 60
ccgcatagtt aagccagtat ctgctccctg cttgtgtgtt ggaggtcgct gagtagtgcg 120
cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc 180

```
ttagggttag gcgttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt    240
gattattgac tagttattaa tagtaatcaa ttacggggtc attagttcat agcccatata    300
tggagttccg cgttacataa cttacggtaa atggcccgcc tggctgaccg cccaacgacc    360
cccgcccatt gacgtcaata atgacgtatg ttcccatagt aacgccaata gggactttcc    420
attgacgtca atgggtggag tatttacggt aaactgccca cttggcagta catcaagtgt    480
atcatatgcc aagtacgccc cctattgacg tcaatgacgg taaatggccc gcctggcatt    540
atgcccagta catgacctta tgggactttc ctacttggca gtacatctac gtattagtca    600
tcgctattac catggtgatg cggtttttgg cagtacatca atgggcgtgg atagcggttt    660
gactcacggg gatttccaag tctccacccc attgacgtca atgggagttt gttttggcac    720
caaaatcaac gggactttcc aaaatgtcgt aacaactccg ccccattgac gcaaatgggc    780
ggtaggcgtg tacggtggga ggtctatata agcagagctc tctggctaac tagagaaccc    840
actgcttact ggcttatcga attaatacg actcactata gggagaccca agcttggtac    900
cgccgccgcc atg ggc aag agg tcc gcc ggc tca atc atg tgg ctc gcg        949
            Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala
             1               5                  10

agc ttg gca gtt gtc ata gct tgt gca ggc gcc gtg acc ctc tcg aac      997
Ser Leu Ala Val Val Ile Ala Cys Ala Gly Ala Val Thr Leu Ser Asn
     15                  20                  25

ttc cag ggc aag gtg atg atg acg gta aat gct act gac gtc aca gat     1045
Phe Gln Gly Lys Val Met Met Thr Val Asn Ala Thr Asp Val Thr Asp
 30                  35                  40                  45

gtc atc acg att cca aca gct gct gga aag aac cta tgc att gtc aga     1093
Val Ile Thr Ile Pro Thr Ala Ala Gly Lys Asn Leu Cys Ile Val Arg
                 50                  55                  60

gca atg gat gtg gga tac atg tgc gat gat act atc act tat gaa tgc     1141
Ala Met Asp Val Gly Tyr Met Cys Asp Asp Thr Ile Thr Tyr Glu Cys
             65                  70                  75

cca gtg ctg tcg gct ggt aat gat cca gaa gac atc gac tgt tgg tgc     1189
Pro Val Leu Ser Ala Gly Asn Asp Pro Glu Asp Ile Asp Cys Trp Cys
         80                  85                  90

aca aag tca gca gtc tac gtc agg tat gga aga tgc acc aag aca cgc     1237
Thr Lys Ser Ala Val Tyr Val Arg Tyr Gly Arg Cys Thr Lys Thr Arg
     95                  100                 105

cac tca aga cgc agt cgg agg tca ctg aca gtg cag aca cac gga gaa     1285
His Ser Arg Arg Ser Arg Arg Ser Leu Thr Val Gln Thr His Gly Glu
110                 115                 120                 125

agc act cta gcg aac aag aag ggg gct tgg atg gac agc acc aag gcc     1333
Ser Thr Leu Ala Asn Lys Lys Gly Ala Trp Met Asp Ser Thr Lys Ala
                130                 135                 140

aca agg tat ttg gta aaa aca gaa tca tgg atc ttg agg aac cct gga     1381
Thr Arg Tyr Leu Val Lys Thr Glu Ser Trp Ile Leu Arg Asn Pro Gly
            145                 150                 155
```

```
tat gcc ctg gtg gca gcc gtc att ggt tgg atg ctt ggg agc aac acc     1429
Tyr Ala Leu Val Ala Ala Val Ile Gly Trp Met Leu Gly Ser Asn Thr
        160                 165                 170

atg cag aga gtt gtg ttt gtc gtg cta ttg ctt ttg gtg gcc cca gct     1477
Met Gln Arg Val Val Phe Val Val Leu Leu Leu Leu Val Ala Pro Ala
    175                 180                 185

tac agc ttc aac tgc ctt gga atg agc aac aga gac ttc ttg gaa gga     1525
Tyr Ser Phe Asn Cys Leu Gly Met Ser Asn Arg Asp Phe Leu Glu Gly
190                 195                 200                 205

gtg tct gga gca aca tgg gtg gat ttg gtt ctc gaa ggc gac agc tgc     1573
Val Ser Gly Ala Thr Trp Val Asp Leu Val Leu Glu Gly Asp Ser Cys
                210                 215                 220

gtg act atc atg tct aag gac aag cct acc atc gat gtg aag atg atg     1621
Val Thr Ile Met Ser Lys Asp Lys Pro Thr Ile Asp Val Lys Met Met
            225                 230                 235

aat atg gag gcg gcc aac ctg gca gag gtc cgc agt tat tgc tat ttg     1669
Asn Met Glu Ala Ala Asn Leu Ala Glu Val Arg Ser Tyr Cys Tyr Leu
        240                 245                 250

gct acc gtc agc gat ctc tcc acc aaa gct gcg tgc ccg acc atg gga     1717
Ala Thr Val Ser Asp Leu Ser Thr Lys Ala Ala Cys Pro Thr Met Gly
    255                 260                 265

gaa gct cac aat gac aaa cgt gct gac cca gct ttt gtg tgc aga caa     1765
Glu Ala His Asn Asp Lys Arg Ala Asp Pro Ala Phe Val Cys Arg Gln
270                 275                 280                 285

gga gtg gtg gac agg ggc tgg ggc aac ggc tgc gga cta ttt ggc aaa     1813
Gly Val Val Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys
                290                 295                 300

gga agc att gac aca tgc gcc aaa ttt gcc tgc tct acc aag gca ata     1861
Gly Ser Ile Asp Thr Cys Ala Lys Phe Ala Cys Ser Thr Lys Ala Ile
            305                 310                 315

gga aga acc atc ttg aaa gag aat atc aag tac gaa gtg gcc att ttt     1909
Gly Arg Thr Ile Leu Lys Glu Asn Ile Lys Tyr Glu Val Ala Ile Phe
        320                 325                 330

gtc cat gga cca act act gtg gag tcg cac gga aac tac tcc aca cag     1957
Val His Gly Pro Thr Thr Val Glu Ser His Gly Asn Tyr Ser Thr Gln
    335                 340                 345

gtt gga gcc act cag gca ggg aga ttc agc atc act cct gcg gcg cct     2005
Val Gly Ala Thr Gln Ala Gly Arg Phe Ser Ile Thr Pro Ala Ala Pro
350                 355                 360                 365

tca tac aca cta aag ctt gga gaa tat gga gag gtg aca gtg gac tgt     2053
```

```
Ser Tyr Thr Leu Lys Leu Gly Glu Tyr Gly Glu Val Thr Val Asp Cys
                370                 375                 380

gaa cca cgg tca ggg att gac acc aat gca tac tac gtg atg act gtt     2101
Glu Pro Arg Ser Gly Ile Asp Thr Asn Ala Tyr Tyr Val Met Thr Val
            385                 390                 395

gga aca aag acg ttc ttg gtc cat cgt gag tgg ttc atg gac ctc aac     2149
Gly Thr Lys Thr Phe Leu Val His Arg Glu Trp Phe Met Asp Leu Asn
        400                 405                 410

ctc cct tgg agc agt gct gga agt act gtg tgg agg aac aga gag acg     2197
Leu Pro Trp Ser Ser Ala Gly Ser Thr Val Trp Arg Asn Arg Glu Thr
    415                 420                 425

tta atg gag ttt gag gaa cca cac gcc acg aag cag tct gtg ata gca     2245
Leu Met Glu Phe Glu Glu Pro His Ala Thr Lys Gln Ser Val Ile Ala
430                 435                 440                 445

ttg ggc tca caa gag gga gct ctg cat caa gct ttg gct gga gcc att     2293
Leu Gly Ser Gln Glu Gly Ala Leu His Gln Ala Leu Ala Gly Ala Ile
                450                 455                 460

cct gtg gaa ttt tca agc aac act gtc aag ttg acg tcg ggt cat ttg     2341
Pro Val Glu Phe Ser Ser Asn Thr Val Lys Leu Thr Ser Gly His Leu
            465                 470                 475

aag tgt aga gtg aag atg gaa aaa ttg cag ttg aag gga aca acc tat     2389
Lys Cys Arg Val Lys Met Glu Lys Leu Gln Leu Lys Gly Thr Thr Tyr
        480                 485                 490

ggc gtc tgt tca aag gct ttc aag ttt ctt ggg act ccc gcg gac aca     2437
Gly Val Cys Ser Lys Ala Phe Lys Phe Leu Gly Thr Pro Ala Asp Thr
    495                 500                 505

ggt cac ggc act gtg gtg ttg gaa ttg cag tac act ggc acg gat gga     2485
Gly His Gly Thr Val Val Leu Glu Leu Gln Tyr Thr Gly Thr Asp Gly
510                 515                 520                 525

cct tgc aaa gtt cct atc tcg tca gtg gct tca ttg aac gac cta acg     2533
Pro Cys Lys Val Pro Ile Ser Ser Val Ala Ser Leu Asn Asp Leu Thr
                530                 535                 540

cca gtg ggc aga ttg gtc act gtc aac cct ttt gtt tca gtg gcc acg     2581
Pro Val Gly Arg Leu Val Thr Val Asn Pro Phe Val Ser Val Ala Thr
            545                 550                 555

gcc aac gct aag gtc ctg att gaa ttg gaa cca ccc ttt gga gac tca     2629
Ala Asn Ala Lys Val Leu Ile Glu Leu Glu Pro Pro Phe Gly Asp Ser
        560                 565                 570

tac ata gtg gtg ggc aga gga gaa caa cag atc aat cac cat tgg cac     2677
Tyr Ile Val Val Gly Arg Gly Glu Gln Gln Ile Asn His His Trp His
    575                 580                 585
```

```
aag tct gga agc agc att ggc aaa gcc ttt aca acc acc ctc aaa gga        2725
Lys Ser Gly Ser Ser Ile Gly Lys Ala Phe Thr Thr Thr Leu Lys Gly
590                 595                 600                 605

gcg cag aga cta gcc gct cta gga gac aca gct tgg gac ttt gga tca        2773
Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser
                610                 615                 620

gtt gga ggg gtg ttc acc tca gtt ggg aag gct gtc cat caa gtg ttc        2821
Val Gly Gly Val Phe Thr Ser Val Gly Lys Ala Val His Gln Val Phe
            625                 630                 635

gga gga gca ttc cgc tca ctg ttc gga ggc atg tcc tgg ata acg caa        2869
Gly Gly Ala Phe Arg Ser Leu Phe Gly Gly Met Ser Trp Ile Thr Gln
        640                 645                 650

gga ttg ctg ggg gct ctc ctg ttg tgg atg ggc atc aat gct cgt gat        2917
Gly Leu Leu Gly Ala Leu Leu Leu Trp Met Gly Ile Asn Ala Arg Asp
    655                 660                 665

agg tcc ata gct ctc acg ttt ctc gca gtt gga gga gtt ctg ctc ttc        2965
Arg Ser Ile Ala Leu Thr Phe Leu Ala Val Gly Gly Val Leu Leu Phe
670                 675                 680                 685

ctc tcc gtg aac gtg cac gcc t gaaggcggcc gctcgagcat gcatctagag         3017
Leu Ser Val Asn Val His Ala
                690

ggccctattc tatagtgtca cctaaatgct agagctcgct gatcagcctc gactgtgcct      3077
tctagttgcc agccatctgt tgtttgcccc tcccccgtgc cttccttgac cctggaaggt      3137
gccactccca ctgtcctttc ctaataaaat gaggaaattg catcgcattg tctgagtagg      3197
tgtcattcta ttctgggggg tggggtgggg caggacagca agggggagga ttgggaagac      3257
aatagcaggc atgctgggga tgcggtgggc tctatggctt ctgaggcgga aagaaccagc      3317
tgcattaatg aatcggccaa cgcgcgggga gaggcggttt gcgtattggg cgctcttccg      3377
cttcctcgct cactgactcg ctgcgctcgg tcgttcggct gcggcgagcg gtatcagctc      3437
actcaaaggc ggtaatacgg ttatccacag aatcagggga taacgcagga aagaacatgt      3497
gagcaaaagg ccagcaaaag gccaggaacc gtaaaaaggc cgcgttgctg gcgtttttcc      3557
ataggctccg cccccctgac gagcatcaca aaaatcgacg ctcaagtcag aggtggcgaa      3617
acccgacagg actataaaga taccaggcgt ttccccctgg aagctccctc gtgcgctctc      3677
ctgttccgac cctgccgctt accggatacc tgtccgcctt tctcccttcg ggaagcgtgg      3737
cgctttctca tagctcacgc tgtaggtatc tcagttcggt gtaggtcgtt cgctccaagc      3797
tgggctgtgt gcacgaaccc cccgttcagc ccgaccgctg cgccttatcc ggtaactatc      3857
gtcttgagtc caacccggta agacacgact tatcgccact ggcagcagcc actggtaaca      3917
ggattagcag agcgaggtat gtaggcggtg ctacagagtt cttgaagtgg tggcctaact      3977
acggctacac tagaagaaca gtatttggta tctgcgctct gctgaagcca gttaccttcg      4037
gaaaaagagt tggtagctct tgatccggca aacaaaccac cgctggtagc ggtggttttt      4097
ttgtttgcaa gcagcagatt acgcgcagaa aaaaaggatc tcaagaagat cctttgatct      4157
tttctacggg gtctgacgct cagtggaacg aaaactcacg ttaagggatt ttggtcatga      4217
gattatcaaa aaggatcttc acctagatcc ttttaaatta aaaatgaagt tttaaatcaa      4277
tctaaagtat atatgagtaa acttggtctg acagttacca atgcttaatc agtgaggcac      4337
ctatctcagc gatctgtcta tttcgttcat ccatagttgc ctgactcccc gtcgtgtaga      4397
taactacgat acgggagggc ttaccatctg gccccagtgc tgcaatgata ccgcgagacc      4457
cacgctcacc ggctccagat ttatcagcaa taaaccagcc agccggaagg gccgagcgca      4517
```

```
gaagtggtcc tgcaacttta tccgcctcca tccagtctat taattgttgc cgggaagcta   4577
gagtaagtag ttcgccagtt aatagtttgc gcaacgttgt tgccattgct acaggcatcg   4637
tggtgtcacg ctcgtcgttt ggtatggctt cattcagctc cggttcccaa cgatcaaggc   4697
gagttacatg atcccccatg ttgtgcaaaa aagcggttag ctccttcggt cctccgatcg   4757
ttgtcagaag taagttggcc gcagtgttat cactcatggt tatggcagca ctgcataatt   4817
ctcttactgt catgccatcc gtaagatgct tttctgtgac tggtgagtac tcaaccaagt   4877
cattctgaga atagtgtatg cggcgaccga gttgctcttg cccggcgtca atacgggata   4937
ataccgcgcc acatagcaga actttaaaag tgctcatcat tggaaaacgt tcttcggggc   4997
gaaaactctc aaggatctta ccgctgttga gatccagttc gatgtaaccc actcgtgcac   5057
ccaactgatc ttcagcatct tttactttca ccagcgtttc tgggtgagca aaaacaggaa   5117
ggcaaaatgc cgcaaaaaag ggaataaggg cgacacggaa atgttgaata ctcatactct   5177
tccttttca atattattga agcatttatc agggttattg tctcatgagc ggatacatat   5237
ttgaatgtat ttagaaaaat aaacaaatag gggttccgcg cacatttccc cgaaaagtgc   5297
cacctgacgt c                                                       5308

<210> 16
<211> 692
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
      synthetic construct

<223> pCBWN

<400> 16
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser Leu Ala
 1               5                  10                  15
Val Val Ile Ala Cys Ala Gly Ala Val Thr Leu Ser Asn Phe Gln Gly
            20                  25                  30
Lys Val Met Met Thr Val Asn Ala Thr Asp Val Thr Asp Val Ile Thr
        35                  40                  45
Ile Pro Thr Ala Ala Gly Lys Asn Leu Cys Ile Val Arg Ala Met Asp
    50                  55                  60
Val Gly Tyr Met Cys Asp Asp Thr Ile Thr Tyr Glu Cys Pro Val Leu
65                  70                  75                  80
Ser Ala Gly Asn Asp Pro Glu Asp Ile Asp Cys Trp Cys Thr Lys Ser
                85                  90                  95
Ala Val Tyr Val Arg Tyr Gly Arg Cys Thr Lys Thr Arg His Ser Arg
            100                 105                 110
Arg Ser Arg Arg Ser Leu Thr Val Gln Thr His Gly Glu Ser Thr Leu
        115                 120                 125
Ala Asn Lys Lys Gly Ala Trp Met Asp Ser Thr Lys Ala Thr Arg Tyr
    130                 135                 140
Leu Val Lys Thr Glu Ser Trp Ile Leu Arg Asn Pro Gly Tyr Ala Leu
145                 150                 155                 160
Val Ala Ala Val Ile Gly Trp Met Leu Gly Ser Asn Thr Met Gln Arg
                165                 170                 175
Val Val Phe Val Val Leu Leu Leu Leu Val Ala Pro Ala Tyr Ser Phe
            180                 185                 190
Asn Cys Leu Gly Met Ser Asn Arg Asp Phe Leu Glu Gly Val Ser Gly
        195                 200                 205
```

Ala Thr Trp Val Asp Leu Val Leu Glu Gly Asp Ser Cys Val Thr Ile
210 215 220
Met Ser Lys Asp Lys Pro Thr Ile Asp Val Lys Met Met Asn Met Glu
225 230 235 240
Ala Ala Asn Leu Ala Glu Val Arg Ser Tyr Cys Tyr Leu Ala Thr Val
245 250 255
Ser Asp Leu Ser Thr Lys Ala Ala Cys Pro Thr Met Gly Glu Ala His
260 265 270
Asn Asp Lys Arg Ala Asp Pro Ala Phe Val Cys Arg Gln Gly Val Val
275 280 285
Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile
290 295 300
Asp Thr Cys Ala Lys Phe Ala Cys Ser Thr Lys Ala Ile Gly Arg Thr
305 310 315 320
Ile Leu Lys Glu Asn Ile Lys Tyr Glu Val Ala Ile Phe Val His Gly
325 330 335
Pro Thr Thr Val Glu Ser His Gly Asn Tyr Ser Thr Gln Val Gly Ala
340 345 350
Thr Gln Ala Gly Arg Phe Ser Ile Thr Pro Ala Ala Pro Ser Tyr Thr
355 360 365
Leu Lys Leu Gly Glu Tyr Gly Glu Val Thr Val Asp Cys Glu Pro Arg
370 375 380
Ser Gly Ile Asp Thr Asn Ala Tyr Tyr Val Met Thr Val Gly Thr Lys
385 390 395 400
Thr Phe Leu Val His Arg Glu Trp Phe Met Asp Leu Asn Leu Pro Trp
405 410 415
Ser Ser Ala Gly Ser Thr Val Trp Arg Asn Arg Glu Thr Leu Met Glu
420 425 430
Phe Glu Glu Pro His Ala Thr Lys Gln Ser Val Ile Ala Leu Gly Ser
435 440 445
Gln Glu Gly Ala Leu His Gln Ala Leu Ala Gly Ala Ile Pro Val Glu
450 455 460
Phe Ser Ser Asn Thr Val Lys Leu Thr Ser Gly His Leu Lys Cys Arg
465 470 475 480
Val Lys Met Glu Lys Leu Gln Leu Lys Gly Thr Thr Tyr Gly Val Cys
485 490 495
Ser Lys Ala Phe Lys Phe Leu Gly Thr Pro Ala Asp Thr Gly His Gly
500 505 510
Thr Val Val Leu Glu Leu Gln Tyr Thr Gly Thr Asp Gly Pro Cys Lys
515 520 525
Val Pro Ile Ser Ser Val Ala Ser Leu Asn Asp Leu Thr Pro Val Gly
530 535 540
Arg Leu Val Thr Val Asn Pro Phe Val Ser Val Ala Thr Ala Asn Ala
545 550 555 560
Lys Val Leu Ile Glu Leu Glu Pro Pro Phe Gly Asp Ser Tyr Ile Val
565 570 575
Val Gly Arg Gly Glu Gln Gln Ile Asn His His Trp His Lys Ser Gly
580 585 590
Ser Ser Ile Gly Lys Ala Phe Thr Thr Thr Leu Lys Gly Ala Gln Arg
595 600 605
Leu Ala Ala Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser Val Gly Gly
610 615 620
Val Phe Thr Ser Val Gly Lys Ala Val His Gln Val Phe Gly Gly Ala
625 630 635 640

```
Phe Arg Ser Leu Phe Gly Gly Met Ser Trp Ile Thr Gln Gly Leu Leu
                645                 650                 655
Gly Ala Leu Leu Leu Trp Met Gly Ile Asn Ala Arg Asp Arg Ser Ile
            660                 665                 670
Ala Leu Thr Phe Leu Ala Val Gly Gly Val Leu Leu Phe Leu Ser Val
        675                 680                 685
Asn Val His Ala
    690
```

<210> 17
<211> 5334
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (916)...(3007)

<221> misc_feature
<222> (1)...(5334)
<223> pCBJE 1-14

<400> 17

```
gacggatcgg gagatctccc gatcccctat ggtgcactct cagtacaatc tgctctgatg     60
ccgcatagtt aagccagtat ctgctccctg cttgtgtgtt ggaggtcgct gagtagtgcg    120
cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc    180
ttagggttag gcgttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt    240
gattattgac tagttattaa tagtaatcaa ttacggggtc attagttcat agcccatata    300
tggagttccg cgttacataa cttacggtaa atggcccgcc tggctgaccg cccaacgacc    360
cccgcccatt gacgtcaata atgacgtatg ttcccatagt aacgccaata gggactttcc    420
attgacgtca atgggtggag tatttacggt aaactgccca cttggcagta catcaagtgt    480
atcatatgcc aagtacgccc cctattgacg tcaatgacgg taaatggccc gcctggcatt    540
atgcccagta catgacctta tgggactttc ctacttggca gtacatctac gtattagtca    600
tcgctattac catggtgatg cggttttggc agtacatcaa tgggcgtgga tagcggtttg    660
actcacgggg atttccaagt ctccacccca ttgacgtcaa tgggagtttg ttttggcacc    720
aaaatcaacg ggactttcca aaatgtcgta acaactccgc cccattgacg caaatgggcg    780
gtaggcgtgt acggtgggag gtctatataa gcagagctct ctggctaact agagaaccca    840
ctgcttactg gcttatcgaa attaatacga ctcactatag ggagacccaa gcttggtacc    900
tctagaggcg ccgcc atg ggc aga aag caa aac aaa aga gga gga aat gaa      951
                  Met Gly Arg Lys Gln Asn Lys Arg Gly Gly Asn Glu
                   1               5                  10

ggc tca atc atg tgg ctc gcg agc ttg gca gtt gtc ata gct tgt gcg      999
Gly Ser Ile Met Trp Leu Ala Ser Leu Ala Val Val Ile Ala Cys Ala
          15                  20                  25

gga gcc atg aag ttg tcg aat ttc cag ggg aag ctt ttg atg acc atc     1047
Gly Ala Met Lys Leu Ser Asn Phe Gln Gly Lys Leu Leu Met Thr Ile
      30                  35                  40
```

```
aac aac acg gac att gca gac gtt atc gtg att ccc acc tca aaa gga      1095
Asn Asn Thr Asp Ile Ala Asp Val Ile Val Ile Pro Thr Ser Lys Gly
 45                  50                  55                  60

gag aac aga tgc tgg gtc cgg gca atc gac gtc ggc tac atg tgt gag      1143
Glu Asn Arg Cys Trp Val Arg Ala Ile Asp Val Gly Tyr Met Cys Glu
                 65                  70                  75

gac act atc acg tac gaa tgt cct aag ctt acc atg ggc aat gat cca      1191
Asp Thr Ile Thr Tyr Glu Cys Pro Lys Leu Thr Met Gly Asn Asp Pro
             80                  85                  90

gag gat gtg gat tgc tgg tgt gac aac caa gaa gtc tac gtc caa tat      1239
Glu Asp Val Asp Cys Trp Cys Asp Asn Gln Glu Val Tyr Val Gln Tyr
         95                  100                 105

gga cgg tgc acg cgg acc agg cat tcc aag cga agc agg aga tcc gtg      1287
Gly Arg Cys Thr Arg Thr Arg His Ser Lys Arg Ser Arg Arg Ser Val
    110                 115                 120

tcg gtc caa aca cat ggg gag agt tca cta gtg aat aaa aaa gag gct      1335
Ser Val Gln Thr His Gly Glu Ser Ser Leu Val Asn Lys Lys Glu Ala
125                 130                 135                 140

tgg ctg gat tca acg aaa gcc aca cga tat ctc atg aaa act gag aac      1383
Trp Leu Asp Ser Thr Lys Ala Thr Arg Tyr Leu Met Lys Thr Glu Asn
                145                 150                 155

tgg atc ata agg aat cct ggc tat gct ttc ctg gcg gcg gta ctt ggc      1431
Trp Ile Ile Arg Asn Pro Gly Tyr Ala Phe Leu Ala Ala Val Leu Gly
            160                 165                 170

tgg atg ctt ggc agt aac aac ggt caa cgc gtg gta ttt acc atc ctc      1479
Trp Met Leu Gly Ser Asn Asn Gly Gln Arg Val Val Phe Thr Ile Leu
        175                 180                 185

ctg ctg ttg gtc gct ccg gct tac agt ttt aat tgt ctg gga atg ggc      1527
Leu Leu Leu Val Ala Pro Ala Tyr Ser Phe Asn Cys Leu Gly Met Gly
    190                 195                 200

aat cgt gac ttc ata gaa gga gcc agt gga gcc act tgg gtg gac ttg      1575
Asn Arg Asp Phe Ile Glu Gly Ala Ser Gly Ala Thr Trp Val Asp Leu
205                 210                 215                 220

gtg ctg gaa gga gat agc tgc ttg aca atc atg gca aac gac aaa cca      1623
Val Leu Glu Gly Asp Ser Cys Leu Thr Ile Met Ala Asn Asp Lys Pro
                225                 230                 235

aca ttg gac gtc cgc atg att aac atc gaa gct agc caa ctt gct gag      1671
Thr Leu Asp Val Arg Met Ile Asn Ile Glu Ala Ser Gln Leu Ala Glu
            240                 245                 250

gtc aga agt tac tgc tat cat gct tca gtc act gac atc tcg acg gtg      1719
```

```
Val Arg Ser Tyr Cys Tyr His Ala Ser Val Thr Asp Ile Ser Thr Val
    255                 260                 265

gct cgg tgc ccc acg act gga gaa gcc cac aac gag aag cga gct gat      1767
Ala Arg Cys Pro Thr Thr Gly Glu Ala His Asn Glu Lys Arg Ala Asp
    270                 275                 280

agt agc tat gtg tgc aaa caa ggc ttc act gac cgt ggg tgg ggc aac      1815
Ser Ser Tyr Val Cys Lys Gln Gly Phe Thr Asp Arg Gly Trp Gly Asn
285                 290                 295                 300

gga tgt gga ctt ttc ggg aag gga agc att gac aca tgt gca aaa ttc      1863
Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile Asp Thr Cys Ala Lys Phe
                305                 310                 315

tcc tgc acc agt aaa gcg att ggg aga aca atc cag cca gaa aac atc      1911
Ser Cys Thr Ser Lys Ala Ile Gly Arg Thr Ile Gln Pro Glu Asn Ile
            320                 325                 330

aaa tac gaa gtt ggc att ttt gtg cat gga acc acc act tcg gaa aac      1959
Lys Tyr Glu Val Gly Ile Phe Val His Gly Thr Thr Thr Ser Glu Asn
        335                 340                 345

cat ggg aat tat tca gcg caa gtt ggg gcg tcc cag gcg gca aag ttt      2007
His Gly Asn Tyr Ser Ala Gln Val Gly Ala Ser Gln Ala Ala Lys Phe
    350                 355                 360

aca gta aca ccc aat gct cct tcg ata acc ctc aaa ctt ggt gac tac      2055
Thr Val Thr Pro Asn Ala Pro Ser Ile Thr Leu Lys Leu Gly Asp Tyr
365                 370                 375                 380

gga gaa gtc aca ctg gac tgt gag cca agg agt gga ctg aac act gaa      2103
Gly Glu Val Thr Leu Asp Cys Glu Pro Arg Ser Gly Leu Asn Thr Glu
                385                 390                 395

gcg ttt tac gtc atg acc gtg ggg tca aag tca ttt ctg gtc cat agg      2151
Ala Phe Tyr Val Met Thr Val Gly Ser Lys Ser Phe Leu Val His Arg
            400                 405                 410

gag tgg ttt cat gac ctc gct ctc ccc tgg acg tcc cct tcg agc aca      2199
Glu Trp Phe His Asp Leu Ala Leu Pro Trp Thr Ser Pro Ser Ser Thr
        415                 420                 425

gcg tgg aga aac aga gaa ctc ctc atg gaa ttt gaa gag gcg cac gcc      2247
Ala Trp Arg Asn Arg Glu Leu Leu Met Glu Phe Glu Glu Ala His Ala
    430                 435                 440

aca aaa cag tcc gtt gtt gct ctt ggg tca cag gaa gga ggc ctc cat      2295
Thr Lys Gln Ser Val Val Ala Leu Gly Ser Gln Glu Gly Gly Leu His
445                 450                 455                 460

cag gcg ttg gca gga gcc atc gtg gtg gag tac tca agc tca gtg aag      2343
Gln Ala Leu Ala Gly Ala Ile Val Val Glu Tyr Ser Ser Ser Val Lys
                465                 470                 475
```

```
tta aca tca ggc cac ctg aaa tgt agg ctg aaa atg gac aaa ctg gct      2391
Leu Thr Ser Gly His Leu Lys Cys Arg Leu Lys Met Asp Lys Leu Ala
            480                 485                 490

ctg aaa ggc aca acc tat ggc atg tgt aca gaa aaa ttc tcg ttc gcg      2439
Leu Lys Gly Thr Thr Tyr Gly Met Cys Thr Glu Lys Phe Ser Phe Ala
        495                 500                 505

aaa aat ccg gcg gac act ggt cac gga aca gtt gtc att gaa ctc tcc      2487
Lys Asn Pro Ala Asp Thr Gly His Gly Thr Val Val Ile Glu Leu Ser
    510                 515                 520

tac tct ggg agt gat ggc ccc tgc aaa att ccg att gct tcc gtt gcg      2535
Tyr Ser Gly Ser Asp Gly Pro Cys Lys Ile Pro Ile Ala Ser Val Ala
525                 530                 535                 540

agc ctc aat gac atg acc ccc gtt ggg cgg ctg gtg aca gtg aac ccc      2583
Ser Leu Asn Asp Met Thr Pro Val Gly Arg Leu Val Thr Val Asn Pro
                545                 550                 555

ttc gtc gcg act tcc agt gcc agc tca aag gtg ctg gtc gag atg gaa      2631
Phe Val Ala Thr Ser Ser Ala Ser Ser Lys Val Leu Val Glu Met Glu
            560                 565                 570

ccc ccc ttc gga gac tcc tac atc gta gtt gga agg gga gac aag cag      2679
Pro Pro Phe Gly Asp Ser Tyr Ile Val Val Gly Arg Gly Asp Lys Gln
        575                 580                 585

atc aac cac cat tgg cac aaa gct gga agc acg ctg ggc aag gcc ttt      2727
Ile Asn His His Trp His Lys Ala Gly Ser Thr Leu Gly Lys Ala Phe
    590                 595                 600

tca aca act ttg aag gga gct caa aga ctg gca gcg ttg ggc gac aca      2775
Ser Thr Thr Leu Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr
605                 610                 615                 620

gcc tgg gac ttt ggc tct att gga ggg gtc ttc aac tcc ata gga aaa      2823
Ala Trp Asp Phe Gly Ser Ile Gly Gly Val Phe Asn Ser Ile Gly Lys
                625                 630                 635

gcc gtt cac caa gtg ttt ggt ggt gcc ttc aga aca ctc ttt ggg gga      2871
Ala Val His Gln Val Phe Gly Gly Ala Phe Arg Thr Leu Phe Gly Gly
            640                 645                 650

atg tct tgg atc aca caa ggg cta atg ggt gcc cta ctg ctc tgg atg      2919
Met Ser Trp Ile Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met
        655                 660                 665

ggc gtc aac gca cga gac cga tca att gct ttg gcc ttc tta gcc aca      2967
Gly Val Asn Ala Arg Asp Arg Ser Ile Ala Leu Ala Phe Leu Ala Thr
    670                 675                 680

ggg ggt gtg ctc gtg ttc tta gcg acc aat gtg cat gct t aattagtttg     3017
```

Gly Gly Val Leu Val Phe Leu Ala Thr Asn Val His Ala
685 690 695

agcggccgct cgagcatgca tctagagggc cctattctat agtgtcacct aaatgctaga 3077
gctcgctgat cagcctcgac tgtgccttct agttgccagc catctgttgt ttgcccctcc 3137
cccgtgcctt ccttgaccct ggaaggtgcc actcccactg tcctttccta ataaaatgag 3197
gaaattgcat cgcattgtct gagtaggtgt cattctattc tgggggggtgg ggtggggcag 3257
gacagcaagg gggaggattg ggaagacaat agcaggcatg ctggggatgc ggtgggctct 3317
atggcttctg aggcggaaag aaccagctgc attaatgaat cggccaacgc gcggggagag 3377
gcggtttgcg tattgggcgc tcttccgctt cctcgctcac tgactcgctg cgctcggtcg 3437
ttcggctgcg gcgagcggta tcagctcact caaaggcggt aatacggtta tccacagaat 3497
caggggataa cgcaggaaag aacatgtgag caaaaggcca gcaaaaggcc aggaaccgta 3557
aaaaggccgc gttgctggcg tttttccata ggctccgccc ccctgacgag catcacaaaa 3617
atcgacgctc aagtcagagg tggcgaaacc cgacaggact ataaagatac caggcgtttc 3677
cccctggaag ctccctcgtg cgctctcctg ttccgaccct gccgcttacc ggatacctgt 3737
ccgcctttct cccttcggga agcgtggcgc tttctcatag ctcacgctgt aggtatctca 3797
gttcggtgta ggtcgttcgc tccaagctgg gctgtgtgca cgaaccccc gttcagcccg 3857
accgctgcgc cttatccggt aactatcgtc ttgagtccaa cccggtaaga cacgacttat 3917
cgccactggc agcagccact ggtaacagga ttagcagagc gaggtatgta ggcggtgcta 3977
cagagttctt gaagtggtgg cctaactacg gctacactag aagaacagta tttggtatct 4037
gcgctctgct gaagccagtt accttcggaa aaagagttgg tagctcttga tccggcaaac 4097
aaaccaccgc tggtagcggt ggtttttttg tttgcaagca gcagattacg cgcagaaaaa 4157
aaggatctca agaagatcct ttgatctttt ctacggggtc tgacgctcag tggaacgaaa 4217
actcacgtta agggattttg gtcatgagat tatcaaaaag gatcttcacc tagatccttt 4277
taaattaaaa atgaagtttt aaatcaatct aaagtatata tgagtaaact tggtctgaca 4337
gttaccaatg cttaatcagt gaggcaccta tctcagcgat ctgtctattt cgttcatcca 4397
tagttgcctg actccccgtc gtgtagataa ctacgatacg ggagggctta ccatctggcc 4457
ccagtgctgc aatgataccg cgagacccac gctcaccggc tccagattta tcagcaataa 4517
accagccagc cggaagggcc gagcgcagaa gtggtcctgc aactttatcc gcctccatcc 4577
agtctattaa ttgttgccgg gaagctagag taagtagttc gccagttaat agtttgcgca 4637
acgttgttgc cattgctaca ggcatcgtgg tgtcacgctc gtcgtttggt atggcttcat 4697
tcagctccgg ttcccaacga tcaaggcgag ttacatgatc ccccatgttg tgcaaaaaag 4757
cggttagctc cttcggtcct ccgatcgttg tcagaagtaa gttggccgca gtgttatcac 4817
tcatggttat ggcagcactg cataattctc ttactgtcat gccatccgta agatgctttt 4877
ctgtgactgg tgagtactca accaagtcat tctgagaata gtgtatgcgg cgaccgagtt 4937
gctcttgccc ggcgtcaata cgggataata ccgcgccaca tagcagaact ttaaaagtgc 4997
tcatcattgg aaaacgttct tcggggcgaa aactctcaag gatcttaccg ctgttgagat 5057
ccagttcgat gtaacccact cgtgcaccca actgatcttc agcatctttt actttcacca 5117
gcgtttctgg gtgagcaaaa acaggaaggc aaaatgccgc aaaaaaggga ataagggcga 5177
cacggaaatg ttgaatactc atactcttcc tttttcaata ttattgaagc atttatcagg 5237
gttattgtct catgagcgga tacatatttg aatgtattta gaaaaataaa caaatagggg 5297
ttccgcgcac atttccccga aaagtgccac ctgacgt 5334

<210> 18
<211> 697
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<223> pCBJE 1-14

<400> 18
Met Gly Arg Lys Gln Asn Lys Arg Gly Gly Asn Glu Gly Ser Ile Met
1 5 10 15
Trp Leu Ala Ser Leu Ala Val Val Ile Ala Cys Ala Gly Ala Met Lys
20 25 30
Leu Ser Asn Phe Gln Gly Lys Leu Leu Met Thr Ile Asn Asn Thr Asp
35 40 45
Ile Ala Asp Val Ile Val Ile Pro Thr Ser Lys Gly Glu Asn Arg Cys
50 55 60
Trp Val Arg Ala Ile Asp Val Gly Tyr Met Cys Glu Asp Thr Ile Thr
65 70 75 80
Tyr Glu Cys Pro Lys Leu Thr Met Gly Asn Asp Pro Glu Asp Val Asp
85 90 95
Cys Trp Cys Asp Asn Gln Glu Val Tyr Val Gln Tyr Gly Arg Cys Thr
100 105 110
Arg Thr Arg His Ser Lys Arg Ser Arg Arg Ser Val Ser Val Gln Thr
115 120 125
His Gly Glu Ser Ser Leu Val Asn Lys Lys Glu Ala Trp Leu Asp Ser
130 135 140
Thr Lys Ala Thr Arg Tyr Leu Met Lys Thr Glu Asn Trp Ile Ile Arg
145 150 155 160
Asn Pro Gly Tyr Ala Phe Leu Ala Ala Val Leu Gly Trp Met Leu Gly
165 170 175
Ser Asn Asn Gly Gln Arg Val Val Phe Thr Ile Leu Leu Leu Leu Val
180 185 190
Ala Pro Ala Tyr Ser Phe Asn Cys Leu Gly Met Gly Asn Arg Asp Phe
195 200 205
Ile Glu Gly Ala Ser Gly Ala Thr Trp Val Asp Leu Val Leu Glu Gly
210 215 220
Asp Ser Cys Leu Thr Ile Met Ala Asn Asp Lys Pro Thr Leu Asp Val
225 230 235 240
Arg Met Ile Asn Ile Glu Ala Ser Gln Leu Ala Glu Val Arg Ser Tyr
245 250 255
Cys Tyr His Ala Ser Val Thr Asp Ile Ser Thr Val Ala Arg Cys Pro
260 265 270
Thr Thr Gly Glu Ala His Asn Glu Lys Arg Ala Asp Ser Ser Tyr Val
275 280 285
Cys Lys Gln Gly Phe Thr Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu
290 295 300
Phe Gly Lys Gly Ser Ile Asp Thr Cys Ala Lys Phe Ser Cys Thr Ser
305 310 315 320
Lys Ala Ile Gly Arg Thr Ile Gln Pro Glu Asn Ile Lys Tyr Glu Val
325 330 335
Gly Ile Phe Val His Gly Thr Thr Thr Ser Glu Asn His Gly Asn Tyr
340 345 350
Ser Ala Gln Val Gly Ala Ser Gln Ala Ala Lys Phe Thr Val Thr Pro
355 360 365
Asn Ala Pro Ser Ile Thr Leu Lys Leu Gly Asp Tyr Gly Glu Val Thr
370 375 380
Leu Asp Cys Glu Pro Arg Ser Gly Leu Asn Thr Glu Ala Phe Tyr Val
385 390 395 400
Met Thr Val Gly Ser Lys Ser Phe Leu Val His Arg Glu Trp Phe His
405 410 415

Asp Leu Ala Leu Pro Trp Thr Ser Pro Ser Ser Thr Ala Trp Arg Asn
420 425 430
Arg Glu Leu Leu Met Glu Phe Glu Glu Ala His Ala Thr Lys Gln Ser
435 440 445
Val Val Ala Leu Gly Ser Gln Glu Gly Gly Leu His Gln Ala Leu Ala
450 455 460
Gly Ala Ile Val Val Glu Tyr Ser Ser Ser Val Lys Leu Thr Ser Gly
465 470 475 480
His Leu Lys Cys Arg Leu Lys Met Asp Lys Leu Ala Leu Lys Gly Thr
485 490 495
Thr Tyr Gly Met Cys Thr Glu Lys Phe Ser Phe Ala Lys Asn Pro Ala
500 505 510
Asp Thr Gly His Gly Thr Val Val Ile Glu Leu Ser Tyr Ser Gly Ser
515 520 525
Asp Gly Pro Cys Lys Ile Pro Ile Ala Ser Val Ala Ser Leu Asn Asp
530 535 540
Met Thr Pro Val Gly Arg Leu Val Thr Val Asn Pro Phe Val Ala Thr
545 550 555 560
Ser Ser Ala Ser Ser Lys Val Leu Val Glu Met Glu Pro Pro Phe Gly
565 570 575
Asp Ser Tyr Ile Val Val Gly Arg Gly Asp Lys Gln Ile Asn His His
580 585 590
Trp His Lys Ala Gly Ser Thr Leu Gly Lys Ala Phe Ser Thr Thr Leu
595 600 605
Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr Ala Trp Asp Phe
610 615 620
Gly Ser Ile Gly Gly Val Phe Asn Ser Ile Gly Lys Ala Val His Gln
625 630 635 640
Val Phe Gly Gly Ala Phe Arg Thr Leu Phe Gly Gly Met Ser Trp Ile
645 650 655
Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met Gly Val Asn Ala
660 665 670
Arg Asp Arg Ser Ile Ala Leu Ala Phe Leu Ala Thr Gly Gly Val Leu
675 680 685
Val Phe Leu Ala Thr Asn Val His Ala
690 695

<210> 19
<211> 5283
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (910)...(2965)

<400> 19
gacggatcgg gagatctccc gatcccctat ggtcgactct cagtacaatc tgctctgatg 60
ccgcatagtt aagccagtat ctgctccctg cttgtgtgtt ggaggtcgct gagtagtgcg 120
cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc 180
ttagggttag gcgttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt 240

```
gattattgac tagttattaa tagtaatcaa ttacggggtc attagttcat agcccatata     300
tggagttccg cgttacataa cttacggtaa atggcccgcc tggctgaccg cccaacgacc     360
cccgcccatt gacgtcaata atgacgtatg ttcccatagt aacgccaata gggactttcc     420
attgacgtca atgggtggac tatttacggt aaactgccca cttggcagta catcaagtgt     480
atcatatgcc aagtacgccc cctattgacg tcaatgacgg taaatggccc gcctggcatt     540
atgcccagta catgacctta tgggactttc ctacttggca gtacatctac gtattagtca     600
tcgctattac catggtgatg cggttttggc agtacatcaa tgggcgtgga tagcggtttg     660
actcacgggg atttccaagt ctccacccca ttgacgtcaa tgggagtttg ttttggcacc     720
aaaatcaacg ggactttcca aaatgtcgta acaactccgc cccattgacg caaatgggcg     780
gtaggcgtgt acggtgggag gtctatataa gcagagctct ctggctaact agagaaccca     840
ctgcttactg gcttatcgaa attaatacga ctcactatag ggagacccaa gcttggtacc     900
gccgccgcc atg ggc aag agg tcc gcc ggc tca atc atg tgg ctc gcg agc     951
          Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser
           1               5                  10

ttg gca gtt gtc ata gct ggt aca agc gct acc acc atc cac cgg gac      999
Leu Ala Val Val Ile Ala Gly Thr Ser Ala Thr Thr Ile His Arg Asp
 15                  20                  25                  30

agg gaa gga tac atg gtt atg cgg gcc agt gga agg gac gct gca agc     1047
Arg Glu Gly Tyr Met Val Met Arg Ala Ser Gly Arg Asp Ala Ala Ser
                35                  40                  45

cag gtc agg gta caa aac gga acg tgc gtc atc ctg gca aca gac atg     1095
Gln Val Arg Val Gln Asn Gly Thr Cys Val Ile Leu Ala Thr Asp Met
            50                  55                  60

gga gag tgg tgt gaa gat tca atc acc tac tct tgc gtc acg att gac     1143
Gly Glu Trp Cys Glu Asp Ser Ile Thr Tyr Ser Cys Val Thr Ile Asp
        65                  70                  75

cag gag gaa gaa ccc gtt gac gtg gac tgc ttc tgc cga ggt gtt gat     1191
Gln Glu Glu Glu Pro Val Asp Val Asp Cys Phe Cys Arg Gly Val Asp
    80                  85                  90

agg gtt aag tta gag tat gga cgc tgt gga agg caa gct gga tct agg     1239
Arg Val Lys Leu Glu Tyr Gly Arg Cys Gly Arg Gln Ala Gly Ser Arg
 95                  100                 105                 110

ggg aaa agg tct gtg gtc att cca aca cat gca caa aaa gac atg gtc     1287
Gly Lys Arg Ser Val Val Ile Pro Thr His Ala Gln Lys Asp Met Val
                115                 120                 125

ggg cga ggt cat gca tgg ctt aaa ggt gac aat att cga gat cat gtc     1335
Gly Arg Gly His Ala Trp Leu Lys Gly Asp Asn Ile Arg Asp His Val
            130                 135                 140

acc cga gtc gag ggc tgg atg tgg aag aac aag ctt cta act gcc gcc     1383
Thr Arg Val Glu Gly Trp Met Trp Lys Asn Lys Leu Leu Thr Ala Ala
        145                 150                 155

att gtg gcc ttg gct tgg ctc atg gtt gat agt tgg atg gcc aga gtg     1431
Ile Val Ala Leu Ala Trp Leu Met Val Asp Ser Trp Met Ala Arg Val
    160                 165                 170
```

act gtc atc ctc ttg gcg ttg agt cta ggg cca gtg tac gcc acg agg 1479
Thr Val Ile Leu Leu Ala Leu Ser Leu Gly Pro Val Tyr Ala Thr Arg
175 180 185 190

tgc acg cat ctt gag aac aga gat ttt gtg aca gga act caa ggg acc 1527
Cys Thr His Leu Glu Asn Arg Asp Phe Val Thr Gly Thr Gln Gly Thr
195 200 205

acc aga gtg tcc cta gtt ttg gaa ctt gga ggc tgc gtg acc atc aca 1575
Thr Arg Val Ser Leu Val Leu Glu Leu Gly Gly Cys Val Thr Ile Thr
210 215 220

gct gag ggc aag cca tcc att gat gta tgg ctc gaa gac att ttt cag 1623
Ala Glu Gly Lys Pro Ser Ile Asp Val Trp Leu Glu Asp Ile Phe Gln
225 230 235

gaa agc ccg gct gaa acc aga gaa tac tgc ctg cac gcc aaa ttg acc 1671
Glu Ser Pro Ala Glu Thr Arg Glu Tyr Cys Leu His Ala Lys Leu Thr
240 245 250

aac aca aaa gtg gag gct cgc tgt cca acc act gga ccg gcg aca ctt 1719
Asn Thr Lys Val Glu Ala Arg Cys Pro Thr Thr Gly Pro Ala Thr Leu
255 260 265 270

ccg gag gag cat cag gct aat atg gtg tgc aag aga gac caa agc gac 1767
Pro Glu Glu His Gln Ala Asn Met Val Cys Lys Arg Asp Gln Ser Asp
275 280 285

cgt gga tgg gga aac cac tgc ggg ttt ttt ggg aag ggc agt ata gtg 1815
Arg Gly Trp Gly Asn His Cys Gly Phe Phe Gly Lys Gly Ser Ile Val
290 295 300

gct tgt gca aag ttt gaa tgc gag gaa gca aaa aaa gct gtg ggc cac 1863
Ala Cys Ala Lys Phe Glu Cys Glu Glu Ala Lys Lys Ala Val Gly His
305 310 315

gtc tat gac tcc aca aag atc acg tat gtt gtc aag gtt gag ccc cac 1911
Val Tyr Asp Ser Thr Lys Ile Thr Tyr Val Val Lys Val Glu Pro His
320 325 330

aca ggg gat tac ttg gct gca aat gag acc aat tca aac agg aaa tca 1959
Thr Gly Asp Tyr Leu Ala Ala Asn Glu Thr Asn Ser Asn Arg Lys Ser
335 340 345 350

gca cag ttt acg gtg gca tcc gag aaa gtg atc ctg cgg ctc ggc gac 2007
Ala Gln Phe Thr Val Ala Ser Glu Lys Val Ile Leu Arg Leu Gly Asp
355 360 365

tat gga gat gtg tcg ctg acg tgt aaa gtg gca agt ggg att gat gtc 2055
Tyr Gly Asp Val Ser Leu Thr Cys Lys Val Ala Ser Gly Ile Asp Val
370 375 380

gcc caa act gtg gtg atg tca ctc gac agc agc aag gac cac ctg cct 2103

Ala Gln Thr Val Val Met Ser Leu Asp Ser Ser Lys Asp His Leu Pro
385 390 395

tct gca tgg caa gtg cac cgt gac tgg ttt gag gac ttg gcg ctg ccc 2151
Ser Ala Trp Gln Val His Arg Asp Trp Phe Glu Asp Leu Ala Leu Pro
400 405 410

tgg aaa cac aag gac aac caa gat tgg aac agt gtg gag aaa ctt gtg 2199
Trp Lys His Lys Asp Asn Gln Asp Trp Asn Ser Val Glu Lys Leu Val
415 420 425 430

gaa ttt gga cca cca cat gct gtg aaa atg gat gtt ttc aat ctg ggg 2247
Glu Phe Gly Pro Pro His Ala Val Lys Met Asp Val Phe Asn Leu Gly
435 440 445

gac cag acg gct gtg ctg ctc aaa tca ctg gca gga gtt ccg ctg gcc 2295
Asp Gln Thr Ala Val Leu Leu Lys Ser Leu Ala Gly Val Pro Leu Ala
450 455 460

agt gtg gag ggc cag aaa tac cac ctg aaa agc ggc cat gtt act tgt 2343
Ser Val Glu Gly Gln Lys Tyr His Leu Lys Ser Gly His Val Thr Cys
465 470 475

gat gtg gga ctg gaa aag ctg aaa ctg aaa ggc aca acc tac tcc atg 2391
Asp Val Gly Leu Glu Lys Leu Lys Leu Lys Gly Thr Thr Tyr Ser Met
480 485 490

tgt gac aaa gca aag ttc aaa tgg aag aga gtt cct gtg gac agc ggc 2439
Cys Asp Lys Ala Lys Phe Lys Trp Lys Arg Val Pro Val Asp Ser Gly
495 500 505 510

cat gac aca gta gtc atg gag gta tca tac aca gga agc gac aag cca 2487
His Asp Thr Val Val Met Glu Val Ser Tyr Thr Gly Ser Asp Lys Pro
515 520 525

tgt cgg atc ccg gtg cgg gct gtg gca cat ggt gtc cca gcg gtt aat 2535
Cys Arg Ile Pro Val Arg Ala Val Ala His Gly Val Pro Ala Val Asn
530 535 540

gta gcc atg ctc ata acc ccc aat cca acc att gaa aca aat ggt ggc 2583
Val Ala Met Leu Ile Thr Pro Asn Pro Thr Ile Glu Thr Asn Gly Gly
545 550 555

gga ttc ata gaa atg cag ctg cca cca ggg gat aac atc atc tat gtg 2631
Gly Phe Ile Glu Met Gln Leu Pro Pro Gly Asp Asn Ile Ile Tyr Val
560 565 570

gga gac ctt agc cag cag tgg ttt cag aaa ggc agt acc att ggt aga 2679
Gly Asp Leu Ser Gln Gln Trp Phe Gln Lys Gly Ser Thr Ile Gly Arg
575 580 585 590

atg ttt gaa aaa acc cgc agg gga ttg gaa agg ctc tct gtg gtt gga 2727
Met Phe Glu Lys Thr Arg Arg Gly Leu Glu Arg Leu Ser Val Val Gly
595 600 605

```
gaa cat gca tgg gac ttt ggc tca gta ggc ggg gta ctg tct tct gtg      2775
Glu His Ala Trp Asp Phe Gly Ser Val Gly Gly Val Leu Ser Ser Val
            610                 615                 620

ggg aag gca atc cac acg gtg ctg ggg gga gct ttc aac acc ctt ttt      2823
Gly Lys Ala Ile His Thr Val Leu Gly Gly Ala Phe Asn Thr Leu Phe
        625                 630                 635

ggg ggg gtt gga ttc atc cct aag atg ctg ctg ggg gtt gct ctg gtc      2871
Gly Gly Val Gly Phe Ile Pro Lys Met Leu Leu Gly Val Ala Leu Val
    640                 645                 650

tgg ttg gga cta aat gcc agg aat cca acg atg tcc atg acg ttt ctt      2919
Trp Leu Gly Leu Asn Ala Arg Asn Pro Thr Met Ser Met Thr Phe Leu
655                 660                 665                 670

gct gtg ggg gct ttg aca ctg atg atg aca atg gga gtt ggg gca t        2965
Ala Val Gly Ala Leu Thr Leu Met Met Thr Met Gly Val Gly Ala
                675                 680                 685

gagcggccgc tcgagcatgc atctagaggg ccctattcta tagtgtcacc taaatgctag   3025
agctcgctga tcagcctcga ctgtgccttc tagttgccag ccatctgttg tttgcccctc   3085
ccccgtgcct tccttgaccc tggaaggtgc cactcccact gtcctttcct aataaaatga   3145
ggaaattgca tcgcattgtc tgagtaggtg tcattctatt ctgggggggtg gggtggggca  3205
ggacagcaag ggggaggatt gggaagacaa tagcaggcat gctggggatg cggtgggctc   3265
tatggcttct gaggcggaaa gaacagctgc attaatgaat cggccaacgc gcggggagag   3325
gcggtttgcg tattgggcgc tcttccgctt cctcgctcac tgactcgctg cgctcggtcg   3385
ttcggctgcg gcgagcggta tcagctcact caaaggcggt aatacggtta tccacagaat   3445
caggggataa cgcaggaaag aacatgtgag caaaaggcca gcaaaaggcc aggaaccgta   3505
aaaaggccgc gttgctggcg tttttccata ggctccgccc ccctgacgag catcacaaaa   3565
atcgacgctc aagtcagagg tggcgaaacc cgacaggact ataaagatac caggcgtttc   3625
cccctggaag ctccctcgtg cgctctcctg ttccgaccct gccgcttacc ggatacctgt   3685
ccgcctttct cccttcggga agcgtggcgc tttctcaatg ctcacgctgt aggtatctca   3745
gttcggtgta ggtcgttcgc tccaagctgg gctgtgtgca cgaaccccccc gttcagcccg   3805
accgctgcgc cttatccggt aactatcgtc ttgagtccaa cccggtaaga cacgacttat   3865
cgccactggc agcagccact ggtaacagga ttagcagagc gaggtatgta ggcggtgcta   3925
cagagttctt gaagtggtgg cctaactacg gctacactag aaggacagta tttggtatct   3985
gcgctctgct gaagccagtt accttcggaa aaagagttgg tagctcttga tccggcaaac   4045
aaaccaccgc tggtagcggt ggtttttttg tttgcaagca gcagattacg cgcagaaaaa   4105
aaggatctca agaagatcct ttgatctttt ctacggggtc tgacgctcag tggaacgaaa   4165
actcacgtta agggattttg gtcatgagat tatcaaaaag gatcttcacc tagatccttt   4225
taaattaaaa atgaagtttt aaatcaatct aaagtatata tgagtaaact tggtctgaca   4285
gttaccaatg cttaatcagt gaggcaccta tctcagcgat ctgtctattt cgttcatcca   4345
tagttgcctg actccccgtc gtgtagataa ctacgatacg ggagggctta ccatctggcc   4405
ccagtgctgc aatgataccg cgagacccac gctcaccggc tccagattta tcagcaataa   4465
accagccagc cggaagggcc gagcgcagaa gtggtcctgc aactttatcc gcctccatcc   4525
agtctattaa ttgttgccgg gaagctagag taagtagttc gccagttaat agtttgcgca   4585
acgttgttgc cattgctaca ggcatcgtgg tgtcacgctc gtcgtttggt atggcttcat   4645
tcagctccgg ttcccaacga tcaaggcgag ttacatgatc ccccatgttg tgcaaaaaag   4705
cggttagctc cttcggtcct ccgatcgttg tcagaagtaa gttggccgca gtgttatcac   4765
tcatggttat ggcagcactg cataattctc ttactgtcat gccatccgta agatgctttt   4825
ctgtgactgg tgagtactca accaagtcat tctgagaata gtgtatgcgg cgaccgagtt   4885
gctcttgccc ggcgtcaata cgggataata ccgcgccaca tagcagaact ttaaaagtgc   4945
```

tcatcattgg aaaacgttct tcggggcgaa aactctcaag gatcttaccg ctgttgagat 5005
ccagttcgat gtaacccact cgtgcaccca actgatcttc agcatctttt actttcacca 5065
gcgtttctgg gtgagcaaaa acaggaaggc aaaatgccgc aaaaaaggga ataagggcga 5125
cacggaaatg ttgaatactc atactcttcc tttttcaata ttattgaagc atttatcagg 5185
gttattgtct catgagcgga tacatatttg aatgtattta gaaaaataaa caaatagggg 5245
ttccgcgcac atttccccga aaagtgccac ctgacgtc 5283

<210> 20
<211> 681
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<400> 20
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser Leu Ala
1 5 10 15
Val Val Ile Ala Gly Thr Ser Ala Val Thr Leu Val Arg Lys Asn Arg
20 25 30
Trp Leu Leu Leu Asn Val Thr Ser Glu Asp Leu Gly Lys Thr Phe Ser
35 40 45
Val Gly Thr Gly Asn Cys Thr Thr Asn Ile Leu Glu Ala Lys Tyr Trp
50 55 60
Cys Pro Asp Ser Met Glu Tyr Asn Cys Pro Asn Leu Ser Pro Arg Glu
65 70 75 80
Glu Pro Asp Asp Ile Asp Cys Trp Cys Tyr Gly Val Glu Asn Val Arg
85 90 95
Val Ala Tyr Gly Lys Cys Asp Ser Ala Gly Arg Ser Arg Arg Ser Arg
100 105 110
Arg Ala Ile Asp Leu Pro Thr His Glu Asn His Gly Leu Lys Thr Arg
115 120 125
Gln Glu Lys Trp Met Thr Gly Arg Met Gly Glu Arg Gln Leu Gln Lys
130 135 140
Ile Glu Arg Trp Phe Val Arg Asn Pro Phe Phe Ala Val Thr Ala Leu
145 150 155 160
Thr Ile Ala Tyr Leu Val Gly Ser Asn Met Thr Gln Arg Val Val Ile
165 170 175
Ala Leu Leu Val Leu Ala Val Gly Pro Ala Tyr Ser Ala His Cys Ile
180 185 190
Gly Ile Thr Asp Arg Asp Phe Ile Glu Gly Val His Gly Gly Thr Trp
195 200 205
Val Ser Ala Thr Leu Glu Gln Asp Lys Cys Val Thr Val Met Ala Pro
210 215 220
Asp Lys Pro Ser Leu Asp Ile Ser Leu Glu Thr Val Ala Ile Asp Arg
225 230 235 240
Pro Ala Glu Val Arg Lys Val Cys Tyr Asn Ala Val Leu Thr His Val
245 250 255
Lys Ile Asn Asp Lys Cys Pro Ser Thr Gly Glu Ala His Leu Ala Glu
260 265 270
Glu Asn Glu Gly Asp Asn Ala Cys Lys Arg Thr Tyr Ser Asp Arg Gly
275 280 285

```
Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile Val Ala Cys
    290                 295                 300
Ala Lys Phe Thr Cys Ala Lys Ser Met Ser Leu Phe Glu Val Asp Gln
305                 310                 315                 320
Thr Lys Ile Gln Tyr Val Ile Arg Ala Gln Leu His Val Gly Ala Lys
                325                 330                 335
Gln Glu Asn Trp Thr Thr Asp Ile Lys Thr Leu Lys Phe Asp Ala Leu
            340                 345                 350
Ser Gly Ser Gln Glu Val Glu Phe Ile Gly Tyr Gly Lys Ala Thr Leu
        355                 360                 365
Glu Cys Gln Val Gln Thr Ala Val Asp Phe Gly Asn Ser Tyr Ile Ala
    370                 375                 380
Glu Met Glu Thr Glu Ser Trp Ile Val Asp Arg Gln Trp Ala Gln Asp
385                 390                 395                 400
Leu Thr Leu Pro Trp Gln Ser Gly Ser Gly Gly Val Trp Arg Glu Met
                405                 410                 415
His His Leu Val Glu Phe Glu Pro Pro His Ala Ala Thr Ile Arg Val
            420                 425                 430
Leu Ala Leu Gly Asn Gln Glu Gly Ser Leu Lys Thr Ala Leu Thr Gly
        435                 440                 445
Ala Met Arg Val Thr Lys Asp Thr Asn Asp Asn Asn Leu Tyr Lys Leu
    450                 455                 460
His Gly Gly His Val Ser Cys Arg Val Lys Leu Ser Ala Leu Thr Leu
465                 470                 475                 480
Lys Gly Thr Ser Tyr Lys Ile Cys Thr Asp Lys Met Phe Phe Val Lys
                485                 490                 495
Asn Pro Thr Asp Thr Gly His Gly Thr Val Val Met Gln Val Lys Val
            500                 505                 510
Ser Lys Gly Ala Pro Cys Arg Ile Pro Val Ile Val Ala Asp Asp Leu
        515                 520                 525
Thr Ala Ala Ile Asn Lys Gly Ile Leu Val Thr Val Asn Pro Ile Ala
    530                 535                 540
Ser Thr Asn Asp Asp Glu Val Leu Ile Glu Val Asn Pro Pro Phe Gly
545                 550                 555                 560
Asp Ser Tyr Ile Ile Val Gly Arg Gly Asp Ser Arg Leu Thr Tyr Gln
                565                 570                 575
Trp His Lys Glu Gly Ser Ser Ile Gly Lys Leu Phe Thr Gln Thr Met
            580                 585                 590
Lys Gly Val Glu Arg Leu Ala Val Met Gly Asp Thr Ala Trp Asp Phe
        595                 600                 605
Ser Ser Ala Gly Gly Phe Phe Thr Ser Val Gly Lys Gly Ile His Thr
    610                 615                 620
Val Phe Gly Ser Ala Phe Gln Gly Leu Phe Gly Gly Leu Asn Trp Ile
625                 630                 635                 640
Thr Lys Val Ile Met Gly Ala Val Leu Ile Trp Val Gly Ile Asn Thr
                645                 650                 655
Arg Asn Met Thr Met Ser Met Ser Met Ile Leu Val Gly Val Ile Met
            660                 665                 670
Met Phe Leu Ser Leu Gly Val Gly Ala
        675                 680
```

<210> 21
<211> 5304
<212> DNA

<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (910)...(2986)

<400> 21
gacggatcgg gagatctccc gatcccctat ggtcgactct cagtacaatc tgctctgatg 60
ccgcatagtt aagccagtat ctgctccctg cttgtgtgtt ggaggtcgct gagtagtgcg 120
cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc 180
ttagggttag gcgttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt 240
gattattgac tagttattaa tagtaatcaa ttacggggtc attagttcat agcccatata 300
tggagttccg cgttacataa cttacggtaa atggcccgcc tggctgaccg cccaacgacc 360
cccgcccatt gacgtcaata atgacgtatg ttcccatagt aacgccaata gggactttcc 420
attgacgtca atgggtggac tatttacggt aaactgccca cttggcagta catcaagtgt 480
atcatatgcc aagtacgccc cctattgacg tcaatgacgg taaatggccc gcctggcatt 540
atgcccagta catgacctta tgggactttc ctacttggca gtacatctac gtattagtca 600
tcgctattac catggtgatg cggttttggc agtacatcaa tgggcgtgga tagcggtttg 660
actcacgggg atttccaagt ctccacccca ttgacgtcaa tgggagtttg ttttggcacc 720
aaaatcaacg ggactttcca aaatgtcgta acaactccgc cccattgacg caaatgggcg 780
gtaggcgtgt acggtgggag gtctatataa gcagagctct ctggctaact agagaaccca 840
ctgcttactg gcttatcgaa attaatacga ctcactatag ggagacccaa gcttggtacc 900
gccgccgcc atg ggc aag agg tcc gcc ggc tca atc atg tgg ctc gcg agc 951
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser
1 5 10

ttg gca gtt gtc ata gct ggt aca agc gct ttg cag tta tca acc tat 999
Leu Ala Val Val Ile Ala Gly Thr Ser Ala Leu Gln Leu Ser Thr Tyr
15 20 25 30

cag ggg aaa gtg tta atg tca atc aac aag act gac gct caa agc gcc 1047
Gln Gly Lys Val Leu Met Ser Ile Asn Lys Thr Asp Ala Gln Ser Ala
35 40 45

ata aac att cct agt gcc aac gga gca aac act tgc att gtg agg gct 1095
Ile Asn Ile Pro Ser Ala Asn Gly Ala Asn Thr Cys Ile Val Arg Ala
50 55 60

cta gat gtg ggg gtc atg tgc aaa gat gac atc aca tac ctg tgc cca 1143
Leu Asp Val Gly Val Met Cys Lys Asp Asp Ile Thr Tyr Leu Cys Pro
65 70 75

gtg ctt tca gcg gga aat gat ccc gag gac att gac tgt tgg tgt gac 1191
Val Leu Ser Ala Gly Asn Asp Pro Glu Asp Ile Asp Cys Trp Cys Asp
80 85 90

gtc gaa gag gtg tgg gtg cac tac ggc aga tgc acg cgc atg gga cat 1239
Val Glu Glu Val Trp Val His Tyr Gly Arg Cys Thr Arg Met Gly His
95 100 105 110

tcg agg cgt agc cga cgg tca atc tct gtg cag cat cat gga gat tcc 1287

```
Ser Arg Arg Ser Arg Arg Ser Ile Ser Val Gln His His Gly Asp Ser
                115                 120                 125

aca ctg gca aca aag aac acg cca tgg ttg gac acc gtg aaa acc acc      1335
Thr Leu Ala Thr Lys Asn Thr Pro Trp Leu Asp Thr Val Lys Thr Thr
            130                 135                 140

aaa tac ttg aca aaa gta gaa aac tgg gtt ttg cgc aat cct gga tat      1383
Lys Tyr Leu Thr Lys Val Glu Asn Trp Val Leu Arg Asn Pro Gly Tyr
        145                 150                 155

gcc cta gtt gcg ctg gcg att gga tgg atg ctc ggt agc aac aac aca      1431
Ala Leu Val Ala Leu Ala Ile Gly Trp Met Leu Gly Ser Asn Asn Thr
    160                 165                 170

cag aga gtg gtt ttt gtg atc atg ctg atg ctg att gct ccg gca tac      1479
Gln Arg Val Val Phe Val Ile Met Leu Met Leu Ile Ala Pro Ala Tyr
175                 180                 185                 190

agc ttc aac tgt ctg gga aca tca aac agg gac ttt gtc gag gga gcc      1527
Ser Phe Asn Cys Leu Gly Thr Ser Asn Arg Asp Phe Val Glu Gly Ala
                195                 200                 205

agt ggg gca aca tgg att gac ttg gta ctt gaa ggg gga agc tgt gtc      1575
Ser Gly Ala Thr Trp Ile Asp Leu Val Leu Glu Gly Gly Ser Cys Val
            210                 215                 220

aca gtg atg gca cca gag aaa cca aca ctg gac ttc aaa gtg atg aag      1623
Thr Val Met Ala Pro Glu Lys Pro Thr Leu Asp Phe Lys Val Met Lys
        225                 230                 235

atg gag gct acc gag tta gcc act gtg cgt gag tat tgt tac gaa gca      1671
Met Glu Ala Thr Glu Leu Ala Thr Val Arg Glu Tyr Cys Tyr Glu Ala
    240                 245                 250

acc ttg gac acg ctg tca aca gtg gca agg tgc ccc aca aca gga gaa      1719
Thr Leu Asp Thr Leu Ser Thr Val Ala Arg Cys Pro Thr Thr Gly Glu
255                 260                 265                 270

gct cac aac acc aaa agg agt gac cca aca ttt gtc tgc aaa aga gat      1767
Ala His Asn Thr Lys Arg Ser Asp Pro Thr Phe Val Cys Lys Arg Asp
                275                 280                 285

gtt gtg gac cgc gga tgg ggt aac gga tgt ggt ctg ttt gga aaa ggg      1815
Val Val Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly
            290                 295                 300

agc att gac aca tgc gct aag ttc aca tgc aaa aac aag gca aca ggg      1863
Ser Ile Asp Thr Cys Ala Lys Phe Thr Cys Lys Asn Lys Ala Thr Gly
        305                 310                 315

aag acg atc ttg aga gaa aac atc aag tat gag gtt gca atc ttt gtg      1911
Lys Thr Ile Leu Arg Glu Asn Ile Lys Tyr Glu Val Ala Ile Phe Val
    320                 325                 330
```

```
cat ggt tca acg gac tct acg tca cat ggc aat tac tct gag cag att      1959
His Gly Ser Thr Asp Ser Thr Ser His Gly Asn Tyr Ser Glu Gln Ile
335                 340                 345                 350

gga aaa aac caa gcg gct aga ttc acc ata agc ccg caa gca ccg tcc      2007
Gly Lys Asn Gln Ala Ala Arg Phe Thr Ile Ser Pro Gln Ala Pro Ser
                355                 360                 365

ttt acg gcc aac atg ggc gag tat gga aca gtt acc att gat tgt gaa      2055
Phe Thr Ala Asn Met Gly Glu Tyr Gly Thr Val Thr Ile Asp Cys Glu
            370                 375                 380

gca aga tca gga atc aac acg gag gat tat tat gtt ttc act gtc aag      2103
Ala Arg Ser Gly Ile Asn Thr Glu Asp Tyr Tyr Val Phe Thr Val Lys
        385                 390                 395

gag aag tca tgg cta gtg aac agg gac tgg ttt cac gac ttg aac ctt      2151
Glu Lys Ser Trp Leu Val Asn Arg Asp Trp Phe His Asp Leu Asn Leu
    400                 405                 410

cca tgg acg agc cct gcc aca act gat tgg cgc aac aga gaa aca ctg      2199
Pro Trp Thr Ser Pro Ala Thr Thr Asp Trp Arg Asn Arg Glu Thr Leu
415                 420                 425                 430

gtg gaa ttt gag gaa ccg cat gcc acc aag caa act gta gta gcc cta      2247
Val Glu Phe Glu Glu Pro His Ala Thr Lys Gln Thr Val Val Ala Leu
                435                 440                 445

gga tcg caa gaa ggt gcc ctg cac aca gca ttg gct gga gcc att cca      2295
Gly Ser Gln Glu Gly Ala Leu His Thr Ala Leu Ala Gly Ala Ile Pro
            450                 455                 460

gcc act gtt agc agc tca acc cta acc ttg caa tca ggg cat ttg aaa      2343
Ala Thr Val Ser Ser Ser Thr Leu Thr Leu Gln Ser Gly His Leu Lys
        465                 470                 475

tgc aga gct aag ctt gac aag gtc aaa atc aag gga acg aca tat ggc      2391
Cys Arg Ala Lys Leu Asp Lys Val Lys Ile Lys Gly Thr Thr Tyr Gly
    480                 485                 490

atg tgt gac tct gcc ttc acc ttc agc aag aac cca act gac aca ggg      2439
Met Cys Asp Ser Ala Phe Thr Phe Ser Lys Asn Pro Thr Asp Thr Gly
495                 500                 505                 510

cac ggg aca gtg att gtg gaa ctg cag tat act gga agc aac gga ccc      2487
His Gly Thr Val Ile Val Glu Leu Gln Tyr Thr Gly Ser Asn Gly Pro
                515                 520                 525

tgc cga gtt ccc atc tcc gtg act gca aac ctc atg gat ttg aca ccg      2535
Cys Arg Val Pro Ile Ser Val Thr Ala Asn Leu Met Asp Leu Thr Pro
            530                 535                 540

gtt gga aga ttg gtc acg gtc aat ccc ttt ata agc aca ggg gga gcg      2583
```

```
Val Gly Arg Leu Val Thr Val Asn Pro Phe Ile Ser Thr Gly Gly Ala
        545                 550                 555

aac aac aag gtc atg atc gaa gtt gaa cca ccc ttt ggc gat tct tac      2631
Asn Asn Lys Val Met Ile Glu Val Glu Pro Pro Phe Gly Asp Ser Tyr
    560                 565                 570

atc gtc gtc gga aga ggc acc acc cag att aac tac cac tgg cac aaa      2679
Ile Val Val Gly Arg Gly Thr Thr Gln Ile Asn Tyr His Trp His Lys
575                 580                 585                 590

gag gga agc agc att ggg aag gct ttg gcg acc aca tgg aaa gga gcc      2727
Glu Gly Ser Ser Ile Gly Lys Ala Leu Ala Thr Thr Trp Lys Gly Ala
                595                 600                 605

caa cgg cta gcc gtc tta ggg gac aca gcg tgg gac ttt gga tct att      2775
Gln Arg Leu Ala Val Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser Ile
            610                 615                 620

gga gga gtt ttc aat tca att ggc aaa gct gtc cac caa gtt ttc gga      2823
Gly Gly Val Phe Asn Ser Ile Gly Lys Ala Val His Gln Val Phe Gly
        625                 630                 635

gga gcg ttc agg act ctg ttc ggg gga atg tcc tgg atc aca cag ggg      2871
Gly Ala Phe Arg Thr Leu Phe Gly Gly Met Ser Trp Ile Thr Gln Gly
    640                 645                 650

cta ctt gga gct ctt ctc ctg tgg atg ggg ttg cag gcc cgc gac agg      2919
Leu Leu Gly Ala Leu Leu Leu Trp Met Gly Leu Gln Ala Arg Asp Arg
655                 660                 665                 670

agc atc tcg ctg act cta ctg gct gtc gga ggg att ctc atc ttt ctg      2967
Ser Ile Ser Leu Thr Leu Leu Ala Val Gly Gly Ile Leu Ile Phe Leu
                675                 680                 685

gca acc agc gtg caa gcc t gagcggccgc tcgagcatgc atctagaggg           3016
Ala Thr Ser Val Gln Ala
            690

ccctattcta tagtgtcacc taaatgctag agctcgctga tcagcctcga ctgtgccttc   3076
tagttgccag ccatctgttg tttgcccctc ccccgtgcct tccttgaccc tggaaggtgc   3136
cactcccact gtcctttcct aataaaatga ggaaattgca tcgcattgtc tgagtaggtg   3196
tcattctatt ctgggggggtg gggtggggca ggacagcaag ggggaggatt gggaagacaa   3256
tagcaggcat gctggggatg cggtgggctc tatggcttct gaggcggaaa gaacagctgc   3316
attaatgaat cggccaacgc gcggggagag gcggtttgcg tattgggcgc tcttccgctt   3376
cctcgctcac tgactcgctg cgctcggtcg ttcggctgcg gcgagcggta tcagctcact   3436
caaaggcggt aatacggtta tccacagaat caggggataa cgcaggaaag aacatgtgag   3496
caaaaggcca gcaaaaggcc aggaaccgta aaaaggccgc gttgctggcg tttttccata   3556
ggctccgccc ccctgacgag catcacaaaa atcgacgctc aagtcagagg tggcgaaacc   3616
cgacaggact ataaagatac caggcgtttc cccctggaag ctccctcgtg cgctctcctg   3676
ttccgaccct gccgcttacc ggatacctgt ccgcctttct cccttcggga agcgtggcgc   3736
tttctcaatg ctcacgctgt aggtatctca gttcggtgta ggtcgttcgc tccaagctgg   3796
gctgtgtgca cgaaccccc gttcagcccg accgctgcgc cttatccggt aactatcgtc   3856
ttgagtccaa cccggtaaga cacgacttat cgccactggc agcagccact ggtaacagga   3916
```

ttagcagagc gaggtatgta ggcggtgcta cagagttctt gaagtggtgg cctaactacg 3976
gctacactag aaggacagta tttggtatct gcgctctgct gaagccagtt accttcggaa 4036
aaagagttgg tagctcttga tccggcaaac aaaccaccgc tggtagcggt ggtttttttg 4096
tttgcaagca gcagattacg cgcagaaaaa aaggatctca agaagatcct ttgatctttt 4156
ctacggggtc tgacgctcag tggaacgaaa actcacgtta agggattttg gtcatgagat 4216
tatcaaaaag gatcttcacc tagatccttt taaattaaaa atgaagtttt aaatcaatct 4276
aaagtatata tgagtaaact tggtctgaca gttaccaatg cttaatcagt gaggcaccta 4336
tctcagcgat ctgtctattt cgttcatcca tagttgcctg actccccgtc gtgtagataa 4396
ctacgatacg ggagggctta ccatctggcc ccagtgctgc aatgataccg cgagacccac 4456
gctcaccggc tccagattta tcagcaataa accagccagc cggaagggcc gagcgcagaa 4516
gtggtcctgc aactttatcc gcctccatcc agtctattaa ttgttgccgg gaagctagag 4576
taagtagttc gccagttaat agtttgcgca acgttgttgc cattgctaca ggcatcgtgg 4636
tgtcacgctc gtcgtttggt atggcttcat tcagctccgg ttcccaacga tcaaggcgag 4696
ttacatgatc ccccatgttg tgcaaaaaag cggttagctc cttcggtcct ccgatcgttg 4756
tcagaagtaa gttggccgca gtgttatcac tcatggttat ggcagcactg cataattctc 4816
ttactgtcat gccatccgta agatgctttt ctgtgactgg tgagtactca accaagtcat 4876
tctgagaata gtgtatgcgg cgaccgagtt gctcttgccc ggcgtcaata cgggataata 4936
ccgcgccaca tagcagaact ttaaaagtgc tcatcattgg aaaacgttct tcggggcgaa 4996
aactctcaag gatcttaccg ctgttgagat ccagttcgat gtaacccact cgtgcaccca 5056
actgatcttc agcatctttt actttcacca gcgtttctgg gtgagcaaaa acaggaaggc 5116
aaaatgccgc aaaaaaggga ataagggcga cacggaaatg ttgaatactc atactcttcc 5176
tttttcaata ttattgaagc atttatcagg gttattgtct catgagcgga tacatatttg 5236
aatgtattta gaaaaataaa caaatagggg ttccgcgcac atttccccga aaagtgccac 5296
ctgacgtc 5304

<210> 22
<211> 692
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<400> 22
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser Leu Ala
1 5 10 15
Val Val Ile Ala Gly Thr Ser Ala Leu Gln Leu Ser Thr Tyr Gln Gly
20 25 30
Lys Val Leu Met Ser Ile Asn Lys Thr Asp Ala Gln Ser Ala Ile Asn
35 40 45
Ile Pro Ser Ala Asn Gly Ala Asn Thr Cys Ile Val Arg Ala Leu Asp
50 55 60
Val Gly Val Met Cys Lys Asp Asp Ile Thr Tyr Leu Cys Pro Val Leu
65 70 75 80
Ser Ala Gly Asn Asp Pro Glu Asp Ile Asp Cys Trp Cys Asp Val Glu
85 90 95
Glu Val Trp Val His Tyr Gly Arg Cys Thr Arg Met Gly His Ser Arg
100 105 110
Arg Ser Arg Arg Ser Ile Ser Val Gln His His Gly Asp Ser Thr Leu
115 120 125
Ala Thr Lys Asn Thr Pro Trp Leu Asp Thr Val Lys Thr Thr Lys Tyr
130 135 140

Leu Thr Lys Val Glu Asn Trp Val Leu Arg Asn Pro Gly Tyr Ala Leu
145 150 155 160
Val Ala Leu Ala Ile Gly Trp Met Leu Gly Ser Asn Asn Thr Gln Arg
165 170 175
Val Val Phe Val Ile Met Leu Met Leu Ile Ala Pro Ala Tyr Ser Phe
180 185 190
Asn Cys Leu Gly Thr Ser Asn Arg Asp Phe Val Glu Gly Ala Ser Gly
195 200 205
Ala Thr Trp Ile Asp Leu Val Leu Glu Gly Gly Ser Cys Val Thr Val
210 215 220
Met Ala Pro Glu Lys Pro Thr Leu Asp Phe Lys Val Met Lys Met Glu
225 230 235 240
Ala Thr Glu Leu Ala Thr Val Arg Glu Tyr Cys Tyr Glu Ala Thr Leu
245 250 255
Asp Thr Leu Ser Thr Val Ala Arg Cys Pro Thr Thr Gly Glu Ala His
260 265 270
Asn Thr Lys Arg Ser Asp Pro Thr Phe Val Cys Lys Arg Asp Val Val
275 280 285
Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile
290 295 300
Asp Thr Cys Ala Lys Phe Thr Cys Lys Asn Lys Ala Thr Gly Lys Thr
305 310 315 320
Ile Leu Arg Glu Asn Ile Lys Tyr Glu Val Ala Ile Phe Val His Gly
325 330 335
Ser Thr Asp Ser Thr Ser His Gly Asn Tyr Ser Glu Gln Ile Gly Lys
340 345 350
Asn Gln Ala Ala Arg Phe Thr Ile Ser Pro Gln Ala Pro Ser Phe Thr
355 360 365
Ala Asn Met Gly Glu Tyr Gly Thr Val Thr Ile Asp Cys Glu Ala Arg
370 375 380
Ser Gly Ile Asn Thr Glu Asp Tyr Tyr Val Phe Thr Val Lys Glu Lys
385 390 395 400
Ser Trp Leu Val Asn Arg Asp Trp Phe His Asp Leu Asn Leu Pro Trp
405 410 415
Thr Ser Pro Ala Thr Thr Asp Trp Arg Asn Arg Glu Thr Leu Val Glu
420 425 430
Phe Glu Glu Pro His Ala Thr Lys Gln Thr Val Val Ala Leu Gly Ser
435 440 445
Gln Glu Gly Ala Leu His Thr Ala Leu Ala Gly Ala Ile Pro Ala Thr
450 455 460
Val Ser Ser Ser Thr Leu Thr Leu Gln Ser Gly His Leu Lys Cys Arg
465 470 475 480
Ala Lys Leu Asp Lys Val Lys Ile Lys Gly Thr Thr Tyr Gly Met Cys
485 490 495
Asp Ser Ala Phe Thr Phe Ser Lys Asn Pro Thr Asp Thr Gly His Gly
500 505 510
Thr Val Ile Val Glu Leu Gln Tyr Thr Gly Ser Asn Gly Pro Cys Arg
515 520 525
Val Pro Ile Ser Val Thr Ala Asn Leu Met Asp Leu Thr Pro Val Gly
530 535 540
Arg Leu Val Thr Val Asn Pro Phe Ile Ser Thr Gly Gly Ala Asn Asn
545 550 555 560
Lys Val Met Ile Glu Val Glu Pro Pro Phe Gly Asp Ser Tyr Ile Val
565 570 575

```
Val Gly Arg Gly Thr Thr Gln Ile Asn Tyr His Trp His Lys Glu Gly
            580                 585                 590
Ser Ser Ile Gly Lys Ala Leu Ala Thr Thr Trp Lys Gly Ala Gln Arg
        595                 600                 605
Leu Ala Val Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser Ile Gly Gly
    610                 615                 620
Val Phe Asn Ser Ile Gly Lys Ala Val His Gln Val Phe Gly Gly Ala
625                 630                 635                 640
Phe Arg Thr Leu Phe Gly Gly Met Ser Trp Ile Thr Gln Gly Leu Leu
                645                 650                 655
Gly Ala Leu Leu Leu Trp Met Gly Leu Gln Ala Arg Asp Arg Ser Ile
            660                 665                 670
Ser Leu Thr Leu Leu Ala Val Gly Gly Ile Leu Ile Phe Leu Ala Thr
        675                 680                 685
Ser Val Gln Ala
    690
```


<210> 23
<211> 5271
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> CDS
<222> (910)...(2953)

<400> 23

```
gacggatcgg gagatctccc gatcccctat ggtcgactct cagtacaatc tgctctgatg      60
ccgcatagtt aagccagtat ctgctccctg cttgtgtgtt ggaggtcgct gagtagtgcg     120
cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc     180
ttagggttag gcgttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt     240
gattattgac tagttattaa tagtaatcaa ttacggggtc attagttcat agcccatata     300
tggagttccg cgttacataa cttacggtaa atggcccgcc tggctgaccg cccaacgacc     360
cccgcccatt gacgtcaata atgacgtatg ttcccatagt aacgccaata gggactttcc     420
attgacgtca atgggtggac tatttacggt aaactgccca cttggcagta catcaagtgt     480
atcatatgcc aagtacgccc cctattgacg tcaatgacgg taaatggccc gcctggcatt     540
atgcccagta catgacctta tgggactttc ctacttggca gtacatctac gtattagtca     600
tcgctattac catggtgatg cggttttggc agtacatcaa tgggcgtgga tagcggtttg     660
actcacgggg atttccaagt ctccacccca ttgacgtcaa tgggagtttg ttttggcacc     720
aaaatcaacg ggactttcca aaatgtcgta acaactccgc cccattgacg caaatgggcg     780
gtaggcgtgt acggtgggag gtctatataa gcagagctct ctggctaact agagaaccca     840
ctgcttactg gcttatcgaa attaatacga ctcactatag ggagacccaa gcttggtacc     900
gccgccgcc atg ggc aag agg tcc gcc ggc tca atc atg tgg ctc gcg agc      951
          Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser
           1               5                  10

ttg gca gtt gtc ata gct ggt aca agc gct gtg acc ttg gtg cgg aaa       999
Leu Ala Val Val Ile Ala Gly Thr Ser Ala Val Thr Leu Val Arg Lys
 15                  20                  25                  30

aac aga tgg ttg ctc cta aat gtg aca tct gag gac ctc ggg aaa aca      1047
```

```
Asn Arg Trp Leu Leu Leu Asn Val Thr Ser Glu Asp Leu Gly Lys Thr
                35                  40                  45

ttc tct gtg ggc aca ggc aac tgc aca aca aac att ttg gaa gcc aag      1095
Phe Ser Val Gly Thr Gly Asn Cys Thr Thr Asn Ile Leu Glu Ala Lys
            50                  55                  60

tac tgg tgc cca gac tca atg gaa tac aac tgt ccc aat ctc agt cca      1143
Tyr Trp Cys Pro Asp Ser Met Glu Tyr Asn Cys Pro Asn Leu Ser Pro
        65                  70                  75

aga gag gag cca gat gac att gat tgc tgg tgc tat ggg gtg gaa aac      1191
Arg Glu Glu Pro Asp Asp Ile Asp Cys Trp Cys Tyr Gly Val Glu Asn
    80                  85                  90

gtt aga gtc gca tat ggt aag tgt gac tca gca ggc agg tct agg agg      1239
Val Arg Val Ala Tyr Gly Lys Cys Asp Ser Ala Gly Arg Ser Arg Arg
95                  100                 105                 110

tca aga agg gcc att gac ttg cct acg cat gaa aac cat ggt ttg aag      1287
Ser Arg Arg Ala Ile Asp Leu Pro Thr His Glu Asn His Gly Leu Lys
                115                 120                 125

acc cgg caa gaa aaa tgg atg act gga aga atg ggt gaa agg caa ctc      1335
Thr Arg Gln Glu Lys Trp Met Thr Gly Arg Met Gly Glu Arg Gln Leu
            130                 135                 140

caa aag att gag aga tgg ttc gtg agg aac ccc ttt ttt gca gtg acg      1383
Gln Lys Ile Glu Arg Trp Phe Val Arg Asn Pro Phe Phe Ala Val Thr
        145                 150                 155

gct ctg acc att gcc tac ctt gtg gga agc aac atg acg caa cga gtc      1431
Ala Leu Thr Ile Ala Tyr Leu Val Gly Ser Asn Met Thr Gln Arg Val
    160                 165                 170

gtg att gcc cta ctg gtc ttg gct gtt ggt ccg gcc tac tca gct cac      1479
Val Ile Ala Leu Leu Val Leu Ala Val Gly Pro Ala Tyr Ser Ala His
175                 180                 185                 190

tgc att gga att act gac agg gat ttc att gag ggg gtg cat gga gga      1527
Cys Ile Gly Ile Thr Asp Arg Asp Phe Ile Glu Gly Val His Gly Gly
                195                 200                 205

act tgg gtt tca gct acc ctg gag caa gac aag tgt gtc act gtt atg      1575
Thr Trp Val Ser Ala Thr Leu Glu Gln Asp Lys Cys Val Thr Val Met
            210                 215                 220

gcc cct gac aag cct tca ttg gac atc tca cta gag aca gta gcc att      1623
Ala Pro Asp Lys Pro Ser Leu Asp Ile Ser Leu Glu Thr Val Ala Ile
        225                 230                 235

gat aga cct gct gag gtg agg aaa gtg tgt tac aat gca gtt ctc act      1671
Asp Arg Pro Ala Glu Val Arg Lys Val Cys Tyr Asn Ala Val Leu Thr
    240                 245                 250
```

```
cat gtg aag att aat gac aag tgc ccc agc act gga gag gcc cac cta     1719
His Val Lys Ile Asn Asp Lys Cys Pro Ser Thr Gly Glu Ala His Leu
255                 260                 265                 270

gct gaa gag aac gaa ggg gac aat gcg tgc aag cgc act tat tct gat     1767
Ala Glu Glu Asn Glu Gly Asp Asn Ala Cys Lys Arg Thr Tyr Ser Asp
                275                 280                 285

aga ggc tgg ggc aat ggc tgt ggc cta ttt ggg aaa ggg agc att gtg     1815
Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile Val
            290                 295                 300

gca tgc gcc aaa ttc act tgt gcc aaa tcc atg agt ttg ttt gag gtt     1863
Ala Cys Ala Lys Phe Thr Cys Ala Lys Ser Met Ser Leu Phe Glu Val
        305                 310                 315

gat cag acc aaa att cag tat gtc atc aga gca caa ttg cat gta ggg     1911
Asp Gln Thr Lys Ile Gln Tyr Val Ile Arg Ala Gln Leu His Val Gly
    320                 325                 330

gcc aag cag gaa aat tgg act acc gac att aag act ctc aag ttt gat     1959
Ala Lys Gln Glu Asn Trp Thr Thr Asp Ile Lys Thr Leu Lys Phe Asp
335                 340                 345                 350

gcc ctg tca ggc tcc cag gaa gtc gag ttc att ggg tat gga aaa gct     2007
Ala Leu Ser Gly Ser Gln Glu Val Glu Phe Ile Gly Tyr Gly Lys Ala
                355                 360                 365

aca ctg gaa tgc cag gtg caa act gcg gtg gac ttt ggt aac agt tac     2055
Thr Leu Glu Cys Gln Val Gln Thr Ala Val Asp Phe Gly Asn Ser Tyr
            370                 375                 380

atc gct gag atg gaa aca gag agc tgg ata gtg gac aga cag tgg gcc     2103
Ile Ala Glu Met Glu Thr Glu Ser Trp Ile Val Asp Arg Gln Trp Ala
        385                 390                 395

cag gac ttg acc ctg cca tgg cag agt gga agt ggc ggg gtg tgg aga     2151
Gln Asp Leu Thr Leu Pro Trp Gln Ser Gly Ser Gly Gly Val Trp Arg
    400                 405                 410

gag atg cat cat ctt gtc gaa ttt gaa cct ccg cat gcc gcc act atc     2199
Glu Met His His Leu Val Glu Phe Glu Pro Pro His Ala Ala Thr Ile
415                 420                 425                 430

aga gta ctg gcc ctg gga aac cag gaa ggc tcc ttg aaa aca gct ctt     2247
Arg Val Leu Ala Leu Gly Asn Gln Glu Gly Ser Leu Lys Thr Ala Leu
                435                 440                 445

act ggc gca atg agg gtt aca aag gac aca aat gac aac aac ctt tac     2295
Thr Gly Ala Met Arg Val Thr Lys Asp Thr Asn Asp Asn Asn Leu Tyr
            450                 455                 460

aaa cta cat ggt gga cat gtt tct tgc aga gtg aaa ttg tca gct ttg     2343
```

```
Lys Leu His Gly Gly His Val Ser Cys Arg Val Lys Leu Ser Ala Leu
        465                 470                 475

aca ctc aag ggg aca tcc tac aaa ata tgc act gac aaa atg ttt ttt     2391
Thr Leu Lys Gly Thr Ser Tyr Lys Ile Cys Thr Asp Lys Met Phe Phe
    480                 485                 490

gtc aag aac cca act gac act ggc cat ggc act gtt gtg atg cag gtg     2439
Val Lys Asn Pro Thr Asp Thr Gly His Gly Thr Val Val Met Gln Val
495                 500                 505                 510

aaa gtg tca aaa gga gcc ccc tgc agg att cca gtg ata gta gct gat     2487
Lys Val Ser Lys Gly Ala Pro Cys Arg Ile Pro Val Ile Val Ala Asp
                515                 520                 525

gat ctt aca gcg gca atc aat aaa ggc att ttg gtt aca gtt aac ccc     2535
Asp Leu Thr Ala Ala Ile Asn Lys Gly Ile Leu Val Thr Val Asn Pro
            530                 535                 540

atc gcc tca acc aat gat gat gaa gtg ctg att gag gtg aac cca cct     2583
Ile Ala Ser Thr Asn Asp Asp Glu Val Leu Ile Glu Val Asn Pro Pro
        545                 550                 555

ttt gga gac agc tac att atc gtt ggg aga gga gat tca cgt ctc act     2631
Phe Gly Asp Ser Tyr Ile Ile Val Gly Arg Gly Asp Ser Arg Leu Thr
    560                 565                 570

tac cag tgg cac aaa gag gga agc tca ata gga aag ttg ttc act cag     2679
Tyr Gln Trp His Lys Glu Gly Ser Ser Ile Gly Lys Leu Phe Thr Gln
575                 580                 585                 590

acc atg aaa ggc gtg gaa cgc ctg gcc gtc atg gga gac acc gcc tgg     2727
Thr Met Lys Gly Val Glu Arg Leu Ala Val Met Gly Asp Thr Ala Trp
                595                 600                 605

gat ttc agc tcc gct gga ggg ttc ttc act tcg gtt ggg aaa gga att     2775
Asp Phe Ser Ser Ala Gly Gly Phe Phe Thr Ser Val Gly Lys Gly Ile
            610                 615                 620

cat acg gtg ttt ggc tct gcc ttt cag ggg cta ttt ggc ggc ttg aac     2823
His Thr Val Phe Gly Ser Ala Phe Gln Gly Leu Phe Gly Gly Leu Asn
        625                 630                 635

tgg ata aca aag gtc atc atg ggg gcg gta ctt ata tgg gtt ggc atc     2871
Trp Ile Thr Lys Val Ile Met Gly Ala Val Leu Ile Trp Val Gly Ile
    640                 645                 650

aac aca aga aac atg aca atg tcc atg agc atg atc ttg gta gga gtg     2919
Asn Thr Arg Asn Met Thr Met Ser Met Ser Met Ile Leu Val Gly Val
655                 660                 665                 670

atc atg atg ttt ttg tct cta gga gtt ggg gcg t gagcggccgc            2963
Ile Met Met Phe Leu Ser Leu Gly Val Gly Ala
                675                 680
```

tcgagcatgc atctagaggg ccctattcta tagtgtcacc taaatgctag agctcgctga 3023
tcagcctcga ctgtgccttc tagttgccag ccatctgttg tttgcccctc ccccgtgcct 3083
tccttgaccc tggaaggtgc cactcccact gtcctttcct aataaaatga ggaaattgca 3143
tcgcattgtc tgagtaggtg tcattctatt ctgggggggtg gggtggggca ggacagcaag 3203
ggggaggatt gggaagacaa tagcaggcat gctggggatg cggtgggctc tatggcttct 3263
gaggcggaaa gaacagctgc attaatgaat cggccaacgc gcggggagag gcggtttgcg 3323
tattgggcgc tcttccgctt cctcgctcac tgactcgctg cgctcggtcg ttcggctgcg 3383
gcgagcggta tcagctcact caaaggcggt aatacggtta tccacagaat caggggataa 3443
cgcaggaaag aacatgtgag caaaaggcca gcaaaaggcc aggaaccgta aaaaggccgc 3503
gttgctggcg tttttccata ggctccgccc ccctgacgag catcacaaaa atcgacgctc 3563
aagtcagagg tggcgaaacc cgacaggact ataaagatac caggcgtttc cccctggaag 3623
ctccctcgtg cgctctcctg ttccgaccct gccgcttacc ggatacctgt ccgccttttct 3683
cccttcggga agcgtggcgc tttctcaatg ctcacgctgt aggtatctca gttcggtgta 3743
ggtcgttcgc tccaagctgg gctgtgtgca cgaacccccc gttcagcccg accgctgcgc 3803
cttatccggt aactatcgtc ttgagtccaa cccggtaaga cacgacttat cgccactggc 3863
agcagccact ggtaacagga ttagcagagc gaggtatgta ggcggtgcta cagagttctt 3923
gaagtggtgg cctaactacg gctacactag aaggacagta tttggtatct gcgctctgct 3983
gaagccagtt accttcggaa aaagagttgg tagctcttga tccggcaaac aaaccaccgc 4043
tggtagcggt ggtttttttg tttgcaagca gcagattacg cgcagaaaaa aaggatctca 4103
agaagatcct ttgatctttt ctacggggtc tgacgctcag tggaacgaaa actcacgtta 4163
agggattttg gtcatgagat tatcaaaaag gatcttcacc tagatccttt taaattaaaa 4223
atgaagtttt aaatcaatct aaagtatata tgagtaaact tggtctgaca gttaccaatg 4283
cttaatcagt gaggcaccta tctcagcgat ctgtctattt cgttcatcca tagttgcctg 4343
actccccgtc gtgtagataa ctacgatacg ggagggctta ccatctggcc ccagtgctgc 4403
aatgataccg cgagacccac gctcaccggc tccagattta tcagcaataa accagccagc 4463
cggaagggcc gagcgcagaa gtggtcctgc aactttatcc gcctccatcc agtctattaa 4523
ttgttgccgg gaagctagag taagtagttc gccagttaat agtttgcgca acgttgttgc 4583
cattgctaca ggcatcgtgg tgtcacgctc gtcgtttggt atggcttcat tcagctccgg 4643
ttcccaacga tcaaggcgag ttacatgatc ccccatgttg tgcaaaaaag cggttagctc 4703
cttcggtcct ccgatcgttg tcagaagtaa gttggccgca gtgttatcac tcatggttat 4763
ggcagcactg cataattctc ttactgtcat gccatccgta agatgctttt ctgtgactgg 4823
tgagtactca accaagtcat tctgagaata gtgtatgcgg cgaccgagtt gctcttgccc 4883
ggcgtcaata cgggataata ccgcgccaca tagcagaact ttaaaagtgc tcatcattgg 4943
aaaacgttct tcggggcgaa aactctcaag gatcttaccg ctgttgagat ccagttcgat 5003
gtaacccact cgtgcaccca actgatcttc agcatctttt actttcacca gcgtttctgg 5063
gtgagcaaaa acaggaaggc aaaatgccgc aaaaaaggga ataagggcga cacggaaatg 5123
ttgaatactc atactcttcc tttttcaata ttattgaagc atttatcagg gttattgtct 5183
catgagcgga tacatatttg aatgtattta gaaaaataaa caaatagggg ttccgcgcac 5243
atttccccga aaagtgccac ctgacgtc 5271

<210> 24
<211> 681
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<400> 24
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser Leu Ala
1 5 10 15

Val Val Ile Ala Gly Thr Ser Ala Val Thr Leu Val Arg Lys Asn Arg
20 25 30

Trp Leu Leu Leu Asn Val Thr Ser Glu Asp Leu Gly Lys Thr Phe Ser
35 40 45

Val Gly Thr Gly Asn Cys Thr Thr Asn Ile Leu Glu Ala Lys Tyr Trp
50 55 60

Cys Pro Asp Ser Met Glu Tyr Asn Cys Pro Asn Leu Ser Pro Arg Glu
65 70 75 80

Glu Pro Asp Asp Ile Asp Cys Trp Cys Tyr Gly Val Glu Asn Val Arg
85 90 95

Val Ala Tyr Gly Lys Cys Asp Ser Ala Gly Arg Ser Arg Arg Ser Arg
100 105 110

Arg Ala Ile Asp Leu Pro Thr His Glu Asn His Gly Leu Lys Thr Arg
115 120 125

Gln Glu Lys Trp Met Thr Gly Arg Met Gly Glu Arg Gln Leu Gln Lys
130 135 140

Ile Glu Arg Trp Phe Val Arg Asn Pro Phe Phe Ala Val Thr Ala Leu
145 150 155 160

Thr Ile Ala Tyr Leu Val Gly Ser Asn Met Thr Gln Arg Val Val Ile
165 170 175

Ala Leu Leu Val Leu Ala Val Gly Pro Ala Tyr Ser Ala His Cys Ile
180 185 190

Gly Ile Thr Asp Arg Asp Phe Ile Glu Gly Val His Gly Gly Thr Trp
195 200 205

Val Ser Ala Thr Leu Glu Gln Asp Lys Cys Val Thr Val Met Ala Pro
210 215 220

Asp Lys Pro Ser Leu Asp Ile Ser Leu Glu Thr Val Ala Ile Asp Arg
225 230 235 240

Pro Ala Glu Val Arg Lys Val Cys Tyr Asn Ala Val Leu Thr His Val
245 250 255

Lys Ile Asn Asp Lys Cys Pro Ser Thr Gly Glu Ala His Leu Ala Glu
260 265 270

Glu Asn Glu Gly Asp Asn Ala Cys Lys Arg Thr Tyr Ser Asp Arg Gly
275 280 285

Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile Val Ala Cys
290 295 300

Ala Lys Phe Thr Cys Ala Lys Ser Met Ser Leu Phe Glu Val Asp Gln
305 310 315 320

Thr Lys Ile Gln Tyr Val Ile Arg Ala Gln Leu His Val Gly Ala Lys
325 330 335

Gln Glu Asn Trp Thr Thr Asp Ile Lys Thr Leu Lys Phe Asp Ala Leu
340 345 350

Ser Gly Ser Gln Glu Val Glu Phe Ile Gly Tyr Gly Lys Ala Thr Leu
355 360 365

Glu Cys Gln Val Gln Thr Ala Val Asp Phe Gly Asn Ser Tyr Ile Ala
370 375 380

Glu Met Glu Thr Glu Ser Trp Ile Val Asp Arg Gln Trp Ala Gln Asp
385 390 395 400

Leu Thr Leu Pro Trp Gln Ser Gly Ser Gly Gly Val Trp Arg Glu Met
405 410 415

His His Leu Val Glu Phe Glu Pro Pro His Ala Ala Thr Ile Arg Val
420 425 430

Leu Ala Leu Gly Asn Gln Glu Gly Ser Leu Lys Thr Ala Leu Thr Gly
435 440 445

Ala Met Arg Val Thr Lys Asp Thr Asn Asp Asn Asn Leu Tyr Lys Leu
450 455 460
His Gly Gly His Val Ser Cys Arg Val Lys Leu Ser Ala Leu Thr Leu
465 470 475 480
Lys Gly Thr Ser Tyr Lys Ile Cys Thr Asp Lys Met Phe Phe Val Lys
485 490 495
Asn Pro Thr Asp Thr Gly His Gly Thr Val Val Met Gln Val Lys Val
500 505 510
Ser Lys Gly Ala Pro Cys Arg Ile Pro Val Ile Val Ala Asp Asp Leu
515 520 525
Thr Ala Ala Ile Asn Lys Gly Ile Leu Val Thr Val Asn Pro Ile Ala
530 535 540
Ser Thr Asn Asp Asp Glu Val Leu Ile Glu Val Asn Pro Pro Phe Gly
545 550 555 560
Asp Ser Tyr Ile Ile Val Gly Arg Gly Asp Ser Arg Leu Thr Tyr Gln
565 570 575
Trp His Lys Glu Gly Ser Ser Ile Gly Lys Leu Phe Thr Gln Thr Met
580 585 590
Lys Gly Val Glu Arg Leu Ala Val Met Gly Asp Thr Ala Trp Asp Phe
595 600 605
Ser Ser Ala Gly Gly Phe Phe Thr Ser Val Gly Lys Gly Ile His Thr
610 615 620
Val Phe Gly Ser Ala Phe Gln Gly Leu Phe Gly Gly Leu Asn Trp Ile
625 630 635 640
Thr Lys Val Ile Met Gly Ala Val Leu Ile Trp Val Gly Ile Asn Thr
645 650 655
Arg Asn Met Thr Met Ser Met Ser Met Ile Leu Val Gly Val Ile Met
660 665 670
Met Phe Leu Ser Leu Gly Val Gly Ala
675 680

<210> 25
<211> 35
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-35
<223> POW 454

<400> 25
aaaagaaaaa gcgctaccac catccaccgg gacag 35

<210> 26
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-41
<223> CPOW 2417

<400> 26
actgttaccc tcaaccccgt actcgccggc gaaaagaaa a 41

<210> 27
<211> 24
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<223> Modified JE Signal

<400> 27
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser Leu Ala
1 5 10 15
Val Val Ile Ala Gly Thr Ser Ala
20

<210> 28
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-36
<223> YF 482

<400> 28
aaaagaaaaa gcgctgtgac cttggtgcgg aaaaac 36

<210> 29
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-41
<223> CYF 2433

<400> 29
acagagatcc tcaaccccgc actcgccggc gaaaaagaaa a 41

<210> 30
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-41
<223> SLE 463

<400> 30
aaaagaaaaa gcgctttgca gttatcaacc tatcagggga a 41

<210> 31
<211> 40
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature
<222> 1-40
<223> CSLE 2477

<400> 31
accgttggtc gcacgttcgg actcgccggc gaaaaagaaa 40